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Schmeissneria: A missing link to angiosperms?

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Abstract

Background: The origin of angiosperms has been under debate since the time of Darwin. While there has been much speculation in past decades about pre-Cretaceous angiosperms, including Archaefructus, these reports are controversial. The earliest reliable fossil record of angiosperms remains restricted to the Cretaceous, even though recent molecular phylogenetic studies suggest an origin for angiosperms much earlier than the current fossil record.

Results: In this paper, after careful SEM and light microscopic work, we report fossils with angiospermous traits of the Jurassic age. The fossils were collected from the Haifanggou Formation (middle Jurassic) in western Liaoning, northeast China. They include two female structures and an associated leaf on the same slab. One of the female structures is physically connected to the apex of a short shoot. The female organs are borne in pairs on short peduncles that are arranged along the axis of the female structure. Each of the female organs has a central unit that is surrounded by an envelope with characteristic longitudinal ribs. Each central unit has two locules completely separated by a vertical septum. The apex of the central unit is completely closed. The general morphology places these fossils into the scope of *Schmeissneria*, an early Jurassic genus that was previously attributed to Ginkgoales.

Conclusion: Because the closed carpel is a character only found in angiosperms, the closed apex of the central unit suggests the presence of angiospermy in *Schmeissneria*. This angiospermous trait implies either a Jurassic angiosperm or a new seed plant group parallel to angiosperms and other known seed plants. As an angiosperm, the Liassic age (earliest Jurassic) of *Schmeissneria microstachys* would suggest an origin of angiosperms during the Triassic. Although still uncertain, this could have a great impact on our perspective of the history, diversity and systematics of seed plants and angiosperms.

Background

Angiosperms are the dominating plant group in current vegetation. They account for the majority of terrestrial primary production, demonstrate the highest diversity in the plant kingdom, and dominate highly diversified habitats [1-4]. The rapid radiation and diversification of angiosperms during the Cretaceous led to major ecological changes on the earth, the latter were a prerequisite for many critical evolutionary events [3] including the later evolution of human beings. However, the origin of this

important plant group has remained obscure since the time of Darwin [3,5-8]. Although palaeobotanists have been searching for pre-Cretaceous angiosperms and some speculate that the angiosperm line may extend back to the Triassic [8,9], the earliest angiosperm fossil records are still restricted to the Cretaceous hitherto [3-5,10-12]. Therefore, it is not surprising that whenever something related to the origin of angiosperms is discovered, such as Sanmiguelia [13] and Archaefructus [6,7,14], it not only receives attention but also triggers controversy within and beyond academic circles. Western Liaoning has been a focus of palaeontological research because of its wealth of fossil plants [7,15-20] and animals [21,22]. Pan's claims of Jurassic angiosperms [15-17] once raised great interest in fossil plants in this region, but they are currently not generally accepted [18,23].

Schmeissneria was first identified as a member of the Ginkgoales [24], with a history dating back to 1838 [25]. Schmeissneria (Stachyopitys) was once thought to be related to conifers [25,26]. Its ginkgoalean affinity was initially proposed based on association: Schenk (1890) classified it as a premature male flower of Baiera, therefore placing Schmeissneria (Stachyopitys) in the Ginkgoales [27]. Based on the data available now, Schenk's conclusion on Schmeissneria (Stachyopitys) has been proven erroneous. First, the male nature of Schmeissneria (Stachyopitys) has been disproved [28]. Second, the connection between Baiera and Schmeissneria has been nullified [24,29]. Third, Schmeissneria has been proven connected with Glossophyllum? sp. A [30], which is dissimilar to any known ginkgoalean leaf [24]. All of this evidence refuted Schenk's initial proposal on the ginkgoalean affinity of Schmeissneria (Stachyopitys), and thus Schmeissneria lost its affinity. Apparently, Kirchner and Van Konijnenburg-Van Cittert did not realize what they had accomplished and conveniently put Schmeissneria in Ginkgoales [24]. In addition, there are a few characters that make Schmeissneria even more mysterious: 1) the internal structure of the reproductive organ, which is important for systematics, is hitherto unknown; 2) its connected vegetative parts do not provide enough information to resolve its systematic position; and 3) its winged seeds are never found in other Ginkgoales.

Here new specimens of *Schmeissneria*, *S. sinensis* Wang sp. nov., are reported from the Haifanggou Formation (middle Jurassic) in western Liaoning, China. The new information from these materials allowed us to re-examine the affinity of *Schmeissneria*.

Results

Description of the specimens

A leaf was closely associated with one of the female structures (Fig. 1a). The leaf was incomplete, over 19 mm long and up to 1.8 mm wide (Fig. 1a). It was slender and cunei-

form, but its apex was unknown (Fig. 1a). The venation was probably parallel (Fig. 1a). No cuticle could be obtained.

Only the apex of a short shoot was organically connected to a female structure (Figs. 1a,c). It was about 2.4 mm long and 2.3 mm wide, with leaf cushions (Fig. 1c). The leaf cushion was about 0.56 mm high and 1.8 mm wide (Fig. 1c).

The female structures were spicate, up to 9.4 mm wide, at least 6 cm (Fig. 1a) and 1.8 cm (Fig. 1b) long, respectively

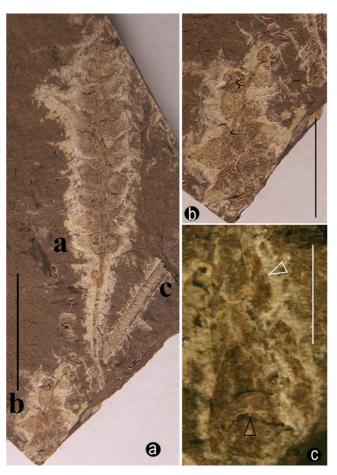


Figure I A general view of female structures, leaf, and short shoot. a. A general view of two female structures (a, b) and one leaf (c) on the same slab. The specimen A is the holotype, and specimens B and C are the paratypes. Specimen numbers 8604a, 8604b, and 8604c. Bar = 2 cm. b. A detailed view of specimen B in Fig. Ia. Note the twisted axis of the female structure and the attached female organs. Specimen number 8604b. Bar = 1 cm. c. A detailed view of the short shoot. Note the leaf cushion (black arrow) and the axis of the connected female structure (white arrow). Specimen number 8604a. Bar = 2 mm.

and generally tapering apically (Figs. 1a-b, 4d). The axes of the female structures were up to 1.3 mm across basally and only 0.2 mm across apically (Figs. 1a-c, 2a-d). The structures were either straight (Fig. 1a) or sinuous (Fig. 1b), longitudinally ribbed (Figs. 2a-c,f, 4d), but free of female organ for about 1.8 cm at the base (Figs. 1a, 4d). One of the female structures was organically connected to the apex of a short shoot (Figs. 1a,c). A female structure had more than 21 female organs attached (Fig. 1a). Generally, the female organs at the basal part were larger and more mature than those at the distal (Figs. 1a-b), except for an isolated female organ pair at the base (Figs. 1a, 2c, 4d). At least some female organs were connated basally (Figs. 2b-e, 4a,d). The peduncle of the female organ pair, rarely seen, was about 0.5 mm long (Fig. 2a).

The female organs were about 1.6-4.6 mm long, about 1.2-4 mm in diameter, widest at the base and constricted at the apex (Figs. 2a-f, 3a, 4a,c). They had an onion-like appearance, including a central unit and a sheathing envelope (Figs. 2b,e, 3a,d, 4c,d). The envelope was longitudinally ribbed internally and externally (Figs. 2a-b,f, 3a,j, 4a,c,d). The apices of the female organs pointed away from the axis of the female structure (Figs. 2a-b, 4a,d). The envelope apex of a large female organ (Figs. 2a-b, 2ef) was more extended than that of a smaller one (Figs. 2cd, 3d). The envelope, sheathing around the central unit, was of multiple parts (Fig. 3c), and inflated (Figs. 2a-b, 3a). The surface of the envelope was smooth locally with elongated epidermal cells (Fig. 3b). The cells within the envelope were elongated: 18-33 μm long and 6-12 μm wide (Fig. 3b). The rugulate pollen grains found on the internal surface of the envelope apex were about 26 µm in diameter (Figs. 3j-o). The central units were about 1.5 to 3.3 mm long and 1 to 3.2 mm in diameter, widest basally and constricted apically (Figs. 2a-b,e, 3a,d, 4a,c). The central unit was attached to a receptacle by a wide base about 1.6 mm in diameter (Fig. 3a), with its apex approaching that of the envelope (Figs. 3d-f, 4c). The distal part of the central unit wall was longitudinally ribbed internally and externally (Figs. 3a,e). No elaborated pollen reception site was observed (Figs. 3d-f,i). The central unit was bilocular, completely separated by a 9 to 19 µm thick vertical septum (Figs. 2f-h, 3d-i, 4b-c). The central unit internal walls were smooth (Figs. 2g, 4b) and flat at the base and rough in the upper part (Figs. 2g, 3e,f, 4b). The septum was complete, extending from the base (Figs. 2g, 4b), through the middle part (Figs. 3g-h) to the apex (Figs. 2h, 3e-f,i, 4c) of the central unit. The septum was papillate (Fig. 2h and its inset).

Remarks: Although two rugulate pollen grains (Figs. 3j-o) were found on the internal surface of the envelope apex, their relationship to *S. sinensis* was tentative and subject to further validation. *Stachyopitys preslii* (possible

male organ of *Schmeissneria* [24]) was only associated with but never connected to *Schmeissneria microstachys* [24], and may also be related to other Ginkgoalean plants [24].

Discussion

The specimens studied here were generally identical to *Schmeissneria* [24], especially the holotype of *S. microstachys* (pl. I, fig. 1) [24]. The latter closely resembled *S. sinensis* in spicate structure, paired female organs, female organ morphology, longitudinally ribbed axes of female structures, and the insertion of a female structure on the apex of a short shoot, even though the specimens described here and *S. microstachys* were from different continents and geological epochs, and *S. sinensis* had more densely clustered female organs and much shorter peduncles of female organ pairs (0.5 mm rather than 2 mm long). Fortunately, the internal structure was preserved in *S. sinensis* and thus shed new light on its affinity.

Structure and Interpretation

In all non-angiospermous seed plants, there is an opening at the nucellar apices to allow the pollen grains to approach the ovules [31,32]. The dimension of the opening has to be large enough to let pollen grains pass through. Unlike seed ferns or other gymnosperms, no opening was found at the apex of the central unit in S. sinensis (Figs. 3d-f). In the view from the interior of the central unit to its apex (Fig. 3f), any opening larger than 20 μm (the normal size of an average pollen grain) should be visible if present on the central unit wall. Although the preservation of the fossils reported here was not perfect, the female organs had both the cellular details (Fig. 3b) and a septum only about 10 µm thick (Fig. 3i) preserved. The preservation of such fine details suggested that preservation fidelity was high enough for structures above the cellular level. Therefore the central unit apex of S. sinensis was physically closed, at least to pollen grains. This was essentially different from the situation in Caytonia, in which there are numerous ovules within a cupule that has an opening for pollen grain entry before pollination [33,34]. There were two alternative interpretations for the absence of pollen entry in Schmeissneria: one was that the pollen entry did not exist at all; the other was that the pollen entry had been obliterated. The latter situation has been seen in Gnetum [32,35], Ephedra, Pinus, Cedrus, Cephalotaxus [36], and possibly in Caytonia [33,34], in all of which the pollen canals were plugged or obliterated by tissue outgrowth or cell proliferation after pollination. This tissue growth or cell proliferation is concomitant with morphological changes [35,36]. However, in the case of Schmeissneria this alternative appeared unlikely for the following reasons: 1) the female organs in Figs. 3d-f were in their early stages (pre-pollination), suggested by their smaller sizes and morphology different from winged

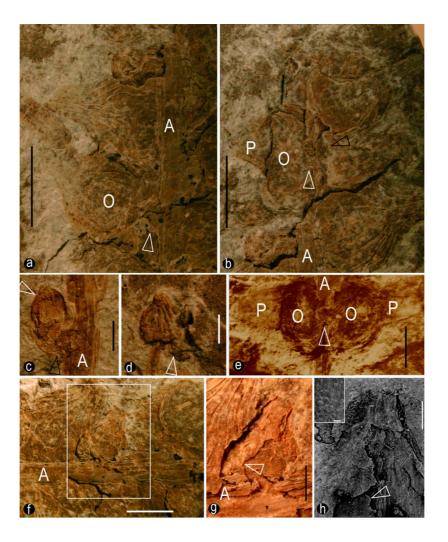


Figure 2

The morphology and structures of the female organs. a. Several female organs attached to the axis of the female structure (A). Note the longitudinal ribs on the sheathing envelope and the central unit (O), the short peduncle (arrow), the extended envelope apex, and dark coaly residues. Specimen number 8604a. Bar = 3 mm. b. Several female organs attached to the axis of the female structure (A). Note the envelope (P) and central unit (O), the fused bases of the female organ pair (white arrow), the mark on the central unit left by the fallen envelope (black arrow), and the extended envelope apex. Specimen number 8604a. Bar = 3 mm. c. An isolated female organ pair in the proximal portion of specimen A. Only one of the pair is evident (white arrow); the other one (black arrow) is obscure due to preservation. Note the longitudinal ribs on the axis of the female structure (A) and the less-extended envelope apex. Specimen number 8604a. Bar = 1 mm. d. The top female organ pair in specimen B. Note the longitudinal ribs on the sheathing envelope, the relic of the missing female organ of the pair (arrow), and the less-extended envelope apex. Specimen number 8604b. Bar = 1 mm. e. A female organ pair in the proximal portion of specimen A. Note the axis of the female structure (A), central units (O), sheathing envelope (P), fused female organ bases (arrow), their spatial relationship (the axis of the female structure is in the foreground), and the extended envelope apices. Colored from an original greyscale picture. Refer to Fig. 4a. Specimen number 8604a. Bar = 2 mm. f. Female organs attached to the axis of the female structure. Note the longitudinal ribs on the axis of the female structure (A), female organs of various sizes and orientations, longitudinal ribs on the envelope, and the extended envelope apex. Specimen number 8604a. Bar = 2 mm. g. A detailed view of the rectangular region in Fig. 2f. Note the exposed internal details of the central unit, smooth wall in the lower part, rough wall in the upper part, large locule, and dark coaly residue. Because the septum and part of the central unit are raised above the level of the side wall, they cast a dark shadow on the latter (arrow) and the shadow extends from the base to the upper parts of the central unit. Refer to Fig. 4b. Specimen number 8604a. Bar = 1 mm. h. A view of a female organ with the exposed interior details of the central unit with the SEM. Note the central unit margin (black arrow) and papillate septum (white arrow and upper left inset) distinct from the interior wall with longitudinal ribs. Specimen number 8604a. Inset bar $= 20 \mu m$, Bar = 0.5 mm.

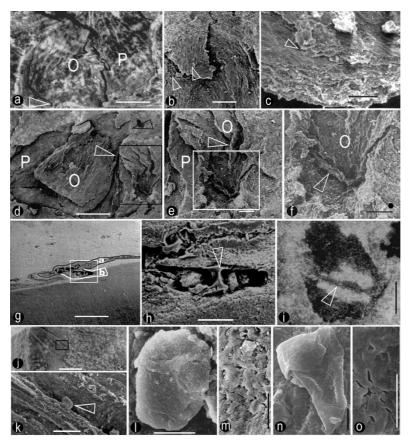


Figure 3 Detailed views of the internal structure of female organs. a. A female organ with a broken envelope and its exposed central unit. Note the sheathing envelope (P), central unit (O), and longitudinal ribs with dark coaly residues on them. The central unit is attached to the receptacle by a wide base (arrow). Specimen number 8604a. Bar = I mm. b. A view of the envelope apex of the female organ in Fig. 3d with the SEM. Note the elongated cells on the smooth epidermis and slightly elongated cells in the tissue of the envelope (arrows). Specimen number 8604b. Bar = 0.1 mm. c. A view of an envelope fragment with the SEM. Note the smooth surfaces and the border between different parts of the envelope (arrow). Specimen number 8604a. Bar = 50 µm. d. A view of a female organ with the envelope (P) and the central unit (O) with the SEM. The texture and the central unit outline (white arrow) could be traced to the envelope apex (black arrow). On the right, there is another female organ with its apex plunging into the sediment matrix (black rectangle). Specimen number 8604b. Bar = 0.5 mm. e. A detailed view of the rectangular region in Fig. 3d with the SEM. Note the spatial relationship between the envelope (P) and the central unit (O), and the longitudinal ribs (arrow) on the internal walls of the central unit. Specimen number 8604b. Bar = 0.1 mm. f. A detailed view of the apex of the central unit in Fig. 3e (rectangle) with the SEM. Note the septum (arrow) across the central unit (O) apex. The black dot beside the bar is about 20 µm in diameter, the size of an average pollen grain. An entry point for a pollen grain of similar size, if present, would be hard to ignore in this image. Therefore, at least the tip of the upper locule (carpel), which is not eclipsed by the septum vestige or in its shadow, is closed. Specimen number 8604b. Bar = 0.1 mm. g. A cross section of two female organs embedded in the sediment. Note the pale sediment (upper half), dark resin (lower half), and darker stripes of two fused female organs (a and b, outlined by black and white lines, respectively). Specimen number 8604a. Bar = 0.5 mm. h. A detailed view of the female organ in Fig. 3g. Note the septum (arrow) and its smooth connection to the side walls of the central unit. Specimen number 8604a. Bar = 0.1 mm. i. A thin section across the apex of the central unit in Fig. 3f. Note the septum (arrow) separating two locules and its smooth connection to the side walls. Light microscope. Specimen number 8604b. Bar = 0.1 mm. j. A view of the internal surface of an envelope apex with the SEM. Note the converging longitudinal ribs. Specimen number 8604a. Bar = 0.5 mm. k. A detailed view of the rectangular region in Fig. 3j. Note the pollen grain (arrow) adherent to one of the longitudinal ribs on the internal surface of the envelope. Specimen number 8604a. Bar = 50 μm. l. A detailed view of the pollen grain in Fig. 3k. Specimen number 8604a. Bar = 10 μm. m. The rugulate sculpture on the pollen grain in Fig. 3I. Specimen number 8604a. Bar = $2 \mu m$. n. Another pollen grain adherent to the internal surface of the envelope apex. Specimen number 8604a. Bar = 5 μ m. o. The rugulate sculpture on the pollen grain in Fig. 3n. Specimen number 8604a. Bar = 2 μm.

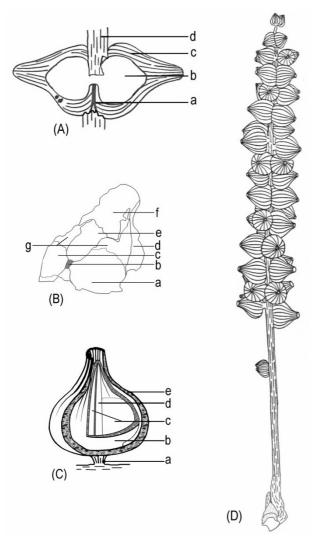


Figure 4 Diagrams of the female organs and the reconstructions of S. sinensis. a. Schematic diagram of the female organ pair shown in Fig. 2e. Note the female organs' fused bases (a), the central unit (b), the sheathing envelope (c), and the axis of the female structure (d). b. Schematic diagram of the female organ in Fig. 2g. Note the smooth locule walls (a and c, probably due to the fallen ovules) and the rough internal walls (g) on each side of the septum (b), different parts of the envelope (d and f), the relic of the broken central unit (e), the vertical septum (b) connected to the base of the central unit. c. A reconstruction of a female organ of S. sinensis. Note the short peduncle of the female organ pair (a) connected to the axis of the female structure (for simplicity, only one organ of the female organ pair is shown here), the central unit (b), two locules (c), the septum (d) separating the two locules, and the sheathing envelope (e). d. Schematic diagram of S. sinensis. From the bottom, note the apex of the short shoot, axis of the female structure, isolated immature female organ pair, clustered female organ pairs along the axis of the female structure, and terminal female organ pair.

seeds [24]; 2) there was neither any trace of abnormal tissue outgrowth nor sudden changes in the wall at the apex of the central unit (Figs. 3d-f), unlike what is observed in *Gnetum* [35], *Cephalotaxus*, and *Ephedra* [36]. Consequently, the other alternative was more reasonable and acceptable. This was the major reason that the authors correlated the structures of *Schmeissneria*'s central unit, envelope, female organ and female structure with an angiosperm's gynoecium, perianth, flower and inflorescence.

A septum extended from the base (Figs. 2g, 4b), through the middle (Figs. 3g-h), to the apex (Figs. 2h, 3e,f,i, 4c) of the central unit. The septum separated the central unit into two independent locules (Figs. 2f-h, 3h-i, 4b-c). The latter were suggestive of two carpels in a central unit (equivalent to gynoecium in angiosperm) [37]. The septum appeared as a ridge raised above the smooth wall of the central unit in the longitudinal view (Fig. 2g). Its smooth connections to the side walls (Figs. 3h-i) and base (Fig. 2g) of the central units suggested that the septum represented an original structure rather than an artefact or alteration. The repeated presence of a septum of various poses in four individual female organs (Figs. 2gh, 3d-i) also strongly suggested its truthful existence. Up until now there has been no report of a complete septum in a seed or ovule in any gymnosperm. Sometimes paired ovules in Ginkgo may appear to have a membraneous division in between. However, their ovules have never been completely enclosed before pollination, the tips of the paired ovules point to different directions, and there is no empty interior space within any ovule. Therefore the paired ovules of Ginkgo were distinct from the central unit of Schmeissneria.

Schmeissneria sinensis had female organs of various ontogenetic stages, from small premature ones at the top to large mature ones at the base of the female structures (Figs. 1, 2, 3, 4). Considering the slight morphological difference between the topmost (most immature) and bottommost (most mature) female organs, it was reasonable to assume that the female organs reported here, especially the smaller ones, were not yet pollinated. The rough internal surface in the upper portion of the central unit (Fig. 2g and Fig. 4b(g)) and the longitudinal ribs on it (Fig. 3e), in contrast to the smooth surface in the lower portion of the internal surface of the central unit (Fig. 2g and Fig. 4b(a,c)), suggested that the upper portion of the central unit was empty, while the lower part was occupied by another inherent substructure (probably an ovule). This situation was unlikely in ovules or seeds, which rarely, if ever, have an empty interior space. It was also unlikely to be a result of differentiated preservation related to histological differences because the presence or absence of tissues is a morphological rather than histological character.

The winged seeds connected to vegetative parts reported by Kirchner and Van Konijnenburg-Van Cittert (Fig. 1b, [24]) were distinct from the female organs reported here, implying the immaturity of *S. sinensis*. Considering the aforementioned details, since Wcislo-Luraniec [28] and Kirchner and Van Konijnenburg-Van Cittert [24] have proven the female nature of *Schmeissneria*, the immature central unit with two locules and a closed apex sheathed by an envelope can be interpreted as a gynoecium of two carpels surrounded by a perianth. This suggested that the carpels (gynoecium) in *Schmeissneria* were closed before pollination, a situation quite different from the "angiospermy" after pollination in gymnosperms [38].

Affinity

In 1838, Presl (pl. 33, fig. 12 only) reported fructifications from Keuper Sandstone (actually Liassic age [24]) of Reundorf near Bamberg, Germany [25]. He considered them male flowers of Pinites microstachys (Conifers). Schenk (pl. XLIV, figs. 11, 12 only, 1867) investigated similar fossils from Veitlahm near Kulmbach, describing them as female flowers of Stachyopitys preslii, which was associated with Schizolepis (a coniferous genus) [26]. Later Schenk (fig. 180b only, 1890) assigned Stachyopitys preslii as male fructifications of Baiera münsteriana (Ginkgoales), and interpreted the formerly "female fructifications" as male flowers in an early stage [27]. This association between Baiera and Stachyopitys proposed by Schenk [27] was later widely accepted (taf. 29, abb. 4 [39]; fig. 239c [40]; fig. 375d [41]; abb. 303h [42]; fig. 198B [31]; taf. 1, fig. lower left [43]; fig. 35d [32]). Actually, fossils of various affinities had been lumped into Stachyopitys preslii [44]. Wcislo-Luraniec (pl. 1 and textfigs. 1, 2, 1992) cast doubt on the male nature of Stachyopitys preslii because of the discovery of "cupules," thereby proving the female nature of the fossil [28]. At the same time, Stachyopitys preslii was found connected with Weber's (p58, taf. 5, fig. 51 [30]) Glossophyllum? sp. A [29]. With more complete fossil materials available, a new genus, Schmeissneria (Ginkgoales), was established for the female Stachyopitys and officially separated from the male Stachyopitys (Figure 1 and pls. I-III, pl. IV, fig. 1, [24]). It appeared that Kirchner and Van Konijnenburg-Van Cittert did not realize that they had undermined the ground for the ginkgoalean affinity of the fossil and uncritically accepted the previous conclusion as true [24]. Furthermore, they did not give any valid reason for why they put the genus in Ginkgoales except for comparing the paired "ovules" of Schmeissneria and the paired ovules of Ginkgo in two sentences (p.207) [24]. Apparently, their comparison was not sufficient to place the fossil in Ginkgoales with confidence because the characteristic collar at the base of Ginkgo seed was absent in all specimens of Schmeissneria and the winged seeds of Schmeissneria were never seen in any Ginkgoales. However, re-examining the references indicated that 1) the discovery of the connection between *Stachyopitys* (*Schmeissneria*) *preslii* and Weber's *Glossophyllum*? sp. A disproved the relationship between *Stachyopitys* (*Schmeissneria*) *preslii* and *Baiera münsteriana* proposed by Schenk [27]; and 2) *Glossophyllum*? sp. A did not belong to the ginkgoalean *Glossophyllum* [24]. Therefore, *Schmeissneria* was dangling phylogenetically. Since the connected vegetative organs could not resolve its affinity, the affinity of *Schmeissneria* had to be resolved based on the internal structure of its own female reproductive organs.

Seed plants include two major groups, gymnosperms and angiosperms. Currently the known major Mesozoic and extant gymnospermous groups include Ginkgoales, Cycadales, Bennettitales, Coniferales, Glossopteridales, Caytoniales, and Gnetales. Among them, Cycadales have pinnate leaves and ovules/seeds attached to the margins of megasporophylls [5,31,32]; Bennettitales have pinnate leaves and conspicuous cone-like reproductive organs with ovules/seeds arranged on a central receptacle [5,31,32]; Coniferales have needle-like or various leaves and more or less cone-like structures composed of bractscale complexes, except for all or some elements of Taxaceae and Podocarpaceae [5,31,32,45], the latter two are distinctly different from Schmeissneria; Glossopteridales have reticulate leaves and their megasporophylls adnated to the adaxial of the foliage [5,31,32,46]; Caytoniales have reticulate leaves and cupules with inverted openings oppositely arranged along the laterals of the megasporophyll axis [5,32]; and Gnetales have a micropylar tube, articulate shoot, opposite/whorled leaves, opposite/ whorled bracts in the reproductive cone, ovuliferous units in the bract axil [31,32,47,48] (Table 1). These characteristics clearly distinguished these groups from Schmeissneria. Therefore these groups will not be considered further in the following discussion on the affinity of Schmeissneria, and the only possibilities remaining are Ginkgoales, Angiosperms, or a new group of seed plants.

Several characters are used to distinguish angiosperms and gymnosperms, including enclosed ovules/seeds, double fertilization, vessel elements, reticulate venation, and tectate-columellate pollen wall structure. However, it is harder to draw a line between these two groups than it might appear: neither of the above characters is a touchstone for angiosperms [1,3]. None of the above characters is unique to angiosperms. All the ovules in gymnosperms are exposed when pollination occurs, but at least some of the ovules/seeds are enclosed after fertilization (Caytoniales [33]; Gnetales [35]; Coniferales and Gnetales [36]; Coniferales [38]). Angiosperms have completely closed carpels. However, in some basal groups this closure is by secretion, and not by postgenital fusion (e.g., Amborellaceae, Schisandraceae, Austrobaileyaceae, Trimeniaceae [49]). Double fertilization has been reported in non-

Table 1: Comparison of Schmeissneria to Ginkgoales, Gnetales, and angiosperms.

Characters	Schmeissneria [24,*]	Ginkgoales[24,31,32,45]	Gnetales[31-32,47,83-84]	Angiosperms[1]		
Long/short shoot	+	+	-	+/-		
Shoot articulate	-	<u>-</u>	+	+/-		
Phyllotaxis	Irregular spiral	Spiral on short shoot	Opposite/whorled	Opposite, whorled, spiral		
Leaf	Slender-cuneiform, obtuse apex	Fan-shaped, or cuneiform	Strap-like, triangular, linear or oval-shaped	Various		
Venation	Parallel	Dichotomous	Parallel or reticulate	Various		
Unisexuality	+	+	+ (-)	+/-		
Reproductive axis articulate	-	-	+	+/-		
Bract	-	-	+	+/- (?)		
Bract arrangement	N/A	-(? bract scale seed complex)	Whorled/paired	Whorled, paired, or N/A		
Ovuliferous unit position	On peduncle tip, in pair	On peduncle tip, single, pair or more in group	In bract axil, one to many in a group	Terminal/axillary		
Ovuliferous unit with septum	+	-	-	+/-		
Övule	Completely enclosed by central unit wall	Exposed through micropyle	Exposed through micropylar tube	Completely enclosed by carpel or secretion		
Micropylar tube	-	<u>-</u>	+	-		

Note that the character of enclosed ovule distinguishes Schmeissneria and angiosperms from the other two groups.*: information from this paper.

angiosperms, as in Ephedra [31,50] and Abies [31]. Vessel elements are also present in various non-angiosperms, such as Selaginella, Equisetum, Pteridium [1], Gigantopteriales [51], and Gnetales [1,4,31,32]. Reticulate venation has been reported in non-angiosperms, including Dipteridaceae [52-55], Gigantopteriales [56,57], Caytoniales [33,58], Glossopteridales [5,40,46,58], Bennettitales [32], and Gnetales [31,32]. Pollen grains with an angiospermlike wall structure have been reported in many pre-Cretaceous plants [59-61] that are regarded as nonangiosperms by others [62]. According to Tomlinson and Takaso [38], the only consistent difference between angiosperms and gymnosperms is that the ovules at pollination are exposed in gymnosperms, but enclosed in angiosperms. Fortunately, the closed carpel at or before pollination is a character that is sufficient to identify an angiosperm alone. This was one of the characters used here to resolve the affinity of Schmeissneria.

Two characters separated *Schmeissneria* from known gymnosperms: the vertical complete septum and the closed apex of the central unit. Considering all available information and the definitions of plant groups, there were two alternatives left to us: 1) accepting that *Schmeissneria* as a new angiosperm in the Jurassic, or 2) proposing *Schmeissneria* as a new gymnosperm. Although the specialized features of the early Cretaceous angiosperm *Archaefructus* [14] and other data [8,9] may imply the possible existence of angiosperms before the Cretaceous, *Schmeissneria* did not look like any known typical angiosperm. However, this dissimilarity was conceivable and understandable since 1) "angiophytes" had not

evolved any typical identifiable angiospermous character [8], and 2) many angiospermous taxa were much more diversified then and much of that diversification had since become extinct [62]. Extreme caution should be exercised when a Jurassic angiosperm, along with their relatives of which we know very little, is compared with the extant angiosperms.

If accepted as an angiosperm, because of its early Jurassic age in Europe, *Schmeissneria* would push the origin of angiospermy back to the Triassic. This would make the claims of Triassic angiosperms [13,59-61], [63,64] less surprising, and also help to bridge the gap between the fossil record [3,5,10-12,65] and molecular data [66-72]. However, it should be kept in mind that *Schmeissneria* might well represent early angiosperms still sporadic in the vegetation dominated by gymnosperms, that it might bear no direct relationship with any known angiosperm, and that the presence of *Schmeissneria* in the flora was still far different from the radiation and diversification of angiosperms.

Conclusion

Schmeissneria is an interesting Jurassic plant that bears a trait of angiosperms (two separated locules in a central unit with a closed apex). This feature has a counterpart in angiosperms (two carpels in the gynoecium). Based on the current definition of angiosperms, Schmeissneria could be classified as an angiosperm. Otherwise, a new group, Schmeissneriales, would have to be established for Schmeissneria. The bottom line is that in whatever position Schmeissneria is placed in the future, it increases the diver-

sity of seed plants and challenges people regarding the systematics of seed plants. Undoubtedly, *Schmeissneria* requires further study.

Systematics

Schmeissneria Kirchner et Van Konijnenburg-Van Cittert 1994, emend.

Type species: *Schmeissneria microstachys* (Presl 1838) Kirchner and Van Konijnenburg-Van Cittert 1994.

Emended diagnosis (based on previous work and present observation)

Plants with long- and short-shoots. Leaves inserted helically on short shoots. Short shoots with leaf cushions. Leaf slender, slightly cuneiform, apex obtuse. Veins parallel, more than two in the proximal part of the leaf, branching in the lower third of the lamina.

Female structures spicate, with a slender axis. Axis of the female structure longitudinally ribbed. Female organ pairs connated basally, borne on a peduncle, arranged along the axis of the female structure. Female organ with a central unit and a sheathing envelope. Envelope of undetermined number of parts, inflated, longitudinally ribbed internally and externally. Central unit bilocular, astylous, with a vertical septum, with longitudinal ribs distally both internally and externally. Seed winged (?).

Type locality: Reundorf near Bamberg, Germany.

Further locality: Oberwaiz, Unternschreez (Lautner) and Schnabelwaid (Creußen) near Bayreuth, Veitlahm, Pechgraben near Kulmbach, Großbellhofen, Rollhofen (Wolfshöhe) northeast of Nuremberg (all in Germany); Odrowaz, Holy Cross Mounts, Poland; western Liaoning, China.

Stratigraphic horizon: Liassic, lower Jurassic (Germany and Poland); middle Jurassic (China).

Remarks: There are a few fossil taxa similar to Schmeissneria, including Ktalenia, Schizolepis, Drepanolepis, Caytonia, Leptostrobus, and Karkenia. Among them, Schizolepis has spirally arranged bilobate two-seed-bearing scales in bract axils [73]; Drepanolepis has spirally arranged sickle-shaped appendages bearing a single seed [74]; Ktalenia has oppositely arranged globose seed-bearing cupules with micropyle pointing downward [75]; Caytonia has oppositely arranged globose multiple-seed-bearing cupules with micropyles pointing to the axis [33,58]; Leptostrobus has spirally arranged bivalvate multiple-seed-bearing cupules in which the two valves share a common slit-like opening pointing outward [76,44]; and Karkenia is an oval-elon-

gate fructification of irregularly disposed atropous, pedunculate ovules/seeds with micropyle pointing to the axis [24,77,78], a genus distinctly different from *Schmeissneria* [24]. These characters distinguish these genera from *Schmeissneria*, which has paired female organs on a peduncle that are arranged along the axis of the female structure [24, also in this paper] (for details, see Table 2).

Schmeissneria sinensis Wang sp. nov.

Diagnosis: The same as that of the genus, except that female organs are densely clustered along the axis of the female structure and the peduncles of the female organ pairs are short in this new species.

Description (see Results)

Holotype: 8604a.

Paratype: 8604b, 8604c.

Etymology: sinensis referring to China, for specimens found in China.

Type locality: Sanjiaochengcun, Jinxi, western Liaoning, China.

Stratigraphic horizon: the Haifanggou Formation, middle Jurassic; equivalent to Aalenian-Bajocian.

Depository: Institute of Botany, the Chinese Academy of Sciences, Beijing, China.

Remarks: The specimens described here and *S. microstachys* were from different continents and geological epochs, and *S. sinensis* had more densely clustered female organs and much shorter peduncles of female organ pairs (0.5 mm rather than 2 mm long). Therefore, a new species, *S. sinensis*, was established for the specimens from Liaoning, China.

Methods

The specimens were collected from Sanjiaochengcun (120°21'E, 40°58'N), Jinxi, Liaoning, China (Fig. 5) in 1988. The bed yielding the present materials belongs to the Haifanggou Formation [15-17,19]. Most people think that the age of the Formation is middle Jurassic [15-17,19,79-81]. The flora of the Haifanggou Formation (including 122 species in 48 genera, dominated by a *Coniopteris-Phoenicopsis* assemblage) iscomparable to the Yorkshire flora (middle Jurassic) of England [79]. This conclusion is supported by chronostratigraphy and other fossils, including ostracodes, bivalves, insects, vertebrates, and palynology [79]. Based on K-Ar, ⁴⁰Ar-³⁹Ar, Rb-Sr, U-Pb, and Sm-Nd datings, the volcanic rocks of the Lanqi

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Table 2: Comparison between Schmeissneria and other possibly related fossil taxa.

	Appendage arrangement	Appendage shape	No. appendages per peduncle	Appendage opening orientation	Appendage apex	Seed/central unit position	Ovule completely enclosed	Vertical septum	No. ovules/seeds per cupule/ central unit	Peduncle connects to
Schmeissneria(young) [24,*]	Irregular	Globose, in pair	2	Away from axis	More or less extended	Within the sheathing envelope	+	+	2?	Envelope
Schmeissneria (mature) [24]	Irregular	Sickle-like, single	I – 2	Away from axis	Wing-shaped	Below the wing, enclosed (?)	?	?	1?	Seed?
Ktalenia[75]	Opposite, subopposite	Globose	I	Down	More or less extended	Within cupule	-	-	I-2	Cupule
Schizolepis[73]	Spiral	Bilobate	l	Up	Lobate	Axil of scale	-	-	2	Scale
Drepanolepis[74]	Spiral	Sickle-like	I	Up	Wing-shaped	Axil of scale	-	-	1	Scale
Leptostrobus[44,76]	Spiral	Round bivalvate	I (2 valves)	Away from axis	Round, thickened margin	Within capsule, axil of valves	-	-	Several	Cupule
Caytonia[33,58]	Opposite	Globose	I	To axis	Extended lip	Within capsule	-	-	Several	Cupule
Karkenia[24,77-78]	Irregular	Round-oval	I	To axis	Acuminate apex	On peduncle	-	-	I	Ovule/seed

Note the characters (enclosed ovule, vertical septum) distinguishing Schmeissneria from the others. *: information from this paper.

Formation (the formation just above the Haifanggou Formation) in Liaoning are 160 to 170 Ma old [79]. Thus, under any circumstance, S. sinensis found in the Haifanggou Formation is older than the Lanqi Formation [79] and at least 160 Ma old. The latest dating, based on volcanic rock and biota, of the Daohugou Formation (equivalent to the Haifanggou Formation) is Callovian to Oxfordian (155 to 165 Ma) [82]. Furthermore, the European Schmeissneria are from the Triassic-Jurassic boundary (Lias α [24]; lower Lias [28]). Considering all the evidence, the authors accept the age of S. sinensis as middle Jurassic.

The studied specimens included two weakly coalified compressions of female structures and one associated leaf. Coaly membranes were found only on the surface of the specimens (Fig. 3a), while most parts of the specimens were replaced by unidentified minerals. The fossils were exposed by trimming away the fine matrix. They were observed under a Zeiss stereomicroscope, photographed with a Nikon 4500 and a Nikon Stereomicroscope SMZ1000 with digital camera DMX1200F. Some of the specimens (Figs. 3b–f, 3j–o) were coated with gold and observed under a Hitachi S800 SEM at Institute of Botany, the Chinese Academy of Sciences. After SEM observations,

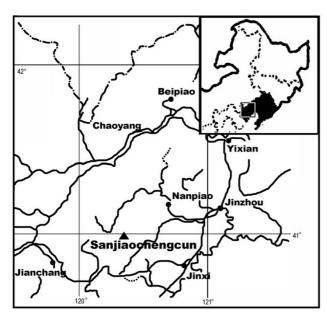


Figure 5
The geographic position of the type fossil locality of *S. sinensis*. The upper right inset shows northeast China, and the black area within is Liaoning Province. The rectangular area in the inset is shown in detail in the main map. The black dots are the major cities in the region, and the black triangle is the type locality of *S. sinensis*, Sanjiaochengcun, Jinxi, Liaoning, China (120°21'E, 40°58'N).

a piece of specimen (Fig. 3d-f) was embedded in an Epon 812 resin, ground into a thin section, and observed with the light microscope (Fig. 3i). One of the trimmed fragments was embedded in Epon 812 resin, ground, coated with gold, and observed with the SEM again (Figs. 3g-h). The SEM images were recorded on black-white negatives. All photographs were later scanned, processed, and pieced together for publication using Photoshop 7.0.

Authors' contributions

XW carried out the fossil processing, photography, SEM, figure preparation, data analysis and interpretation, manuscript drafting and finalization. SD & BG did the field work, collected the specimen, participated in the data analysis and interpretation, and manuscript modification. JC participated in the data analysis and interpretation, and manuscript modification. YY participated in the data analysis and interpretation, and manuscript modification. All authors read and approved the final manuscript.

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References

- Eames AJ: Morphology of the angiosperms New York: McGraw-Hill Book Company; 1961.
- Heywood VH: Flowering plants of the world Oxford: Oxford University Press: 1979.
- Friis EM, Chaloner WG, Crane PR: Introduction to angiosperms. In Origins of angiosperms and their biological consequences Edited by: Friis EM, Chaloner WG, Crane PR. Cambridge: Cambridge University Press; 1987:1-15.
- 4. Cronquist A: The evolution and classification of flowering plants Bronx: New York Botanical Garden; 1988.
- 5. Hughes NF: The enigma of angiosperm origins Cambridge: Cambridge University Press; 1994.
- Sun G, Dilcher DL, Zheng S-L, Zhou Z-K: In search of the first flower: a Jurassic angiosperm, Archaefructus, from Northeast China. Science 1998, 282:1692-1695.
- Sun G, Ji Q, Dilcher DL, Zheng S, Nixon KC, Wang X: Archaefructaceae, a new basal angiosperm family. Science 2002, **296:**899-904.
- De Bodt S, Maere S, Van de Peer Y: Genome duplication and the 8. origin of angiosperms. Trends Ecol Evol 2005, 20:591-597
- Doyle JA, Donoghue MJ: Seed plant phylogeny and the origin of angiosperms: an experimental cladistic approach. Bot Rev 1986, **52:**321-431.
- 10. Friis EM, Pedersen KR, Crane PR: Reproductive structure and organization of basal angiosperms from the early Cretaceous (Barremian or Aptian) of Western Portugal. Int J Plant Sci 2000:S169-S182.
- 11. Friis EM, Pedersen KR, Crane PR: Cretaceous angiosperm flowers: innovation and evolution in plant reproduction. Palaeogeo Palaeoclimat Palaeoeco 2006, 232:251-293.
- 12. Feild TS, Arens NC: Form, function and environments of the early angiosperms: merging extant phylogeny and ecophysiology with fossils. New Phytol 2005, 166:383-408
- 13. Cornet B: The reproductive morphology and biology of Sanmiguelia lewisii, and its bearing on angiosperm evolution in the late Triassic. Evol Trends Plant 1989, 3:25-51.
- 14. Friis EM, Doyle JA, Endress PK, Leng Q: Archaefructus angiosperm precursor or specialized early angiosperm? Trends Plant Sci 2003, 8:369-373.
- Pan G: The Jurassic precursors of angiosperms from Yanliao region of North China and the origin of angiosperms. Chin Sci Bull 1983, 28:15-20
- 16. Pan G: Juglandaceous plant (Pterocarya) from middle Jurassic of Yanliao region, north China. Acta Sci Nat Univ Sunyatseni 1997,
- Pan K: Rhamnaceous plants from middle Jurassic of Yanliao region, north China. Bull Geol Soc Liaoning Prov, China 1990, 2:1-8.
- 18. Xu R: Do fossil angiosperms really occur in Jurassic beds of the Yanshan-Liaoning area, north China. Chin Sci Bull 1987, 32:1712-1714.
- Kimura T, Ohana T, Zhao LM, Geng BY: Pankuangia haifanggouensis gen. et sp. nov., a fossil plant with unknown affinity from the middle Jurassic Haifanggou Fromation, western Liaoning, Northeast China. Bull Kitakyushu Mus Nat Hist 1994, 13:255-261.
- Ji Q, Li H, Bowe M, Liu Y, Taylor DW: Early Cretaceous Archaefructus eoflora sp. nov. with bisexual flowers from Beipiao, Western Liaoning, China. Acta Geol Sin 2004, 78:883-896.
 21. Wang X-L, Zhou Z-H, He H-Y, Jin F, Wang Y-Q, Zhang J-Y, Wang Y,
- Xu X, Zhang F-C: On the stratigraphic relationship and age of the fossil-bearing layer at Daohugou, Ningcheng, Inner Mongolia. Chin Sci Bull 2005, 50:2127-2135.
- 22. Li G, Luo ZX: A Cretaceous symmetrodont therian with some monotreme-like postcranial features. Nature 439:195-200.
- Zheng S-L, Zhang L-J, Gong E-P: A discovery of Anomozamites with reproductive organs. Acta Bot Sin 2003, 45:667-672.
- Kirchner M, Van Konijnenburg-Van Cittert JHA: Schmeissneria microstachys (Presl, 1833) Kirchner et Van Konijnenburg-Van Cittert, comb. nov. and Karkenia hauptmannii Kirchner et Van Konijnenburg-Van Cittert, sp. nov., plants with ginkgoalean affinities from the Liassic of Germany. Rev Palaeobot Palynol 1994, 83:199-215.

- 25. Presl KB: Versuch einer geognostisch-botanischen Darstellung der Flora der Vorwelt Volume 2. Issue 7-8 Edited by: Sternberg KM. Prag: Johann Spurny; 1838:81-220.
- Schenk A: Die fossile Flora der Grenzschichten des Keupers und Lias Frankens Wiesbaden: C.W. Kreidel's Verlag; 1867.
- Schenk A: Paläophytologie Volume 2. München: Druck und Verlag von R. Oldenbourg; 1890.
- Wcislo-Luraniec E: A fructification of Stachyopitys preslii Schenk from the lower Jurassic of Poland. Cour Forsch-Inst Senck 1992, 147:247-253.
- Kirchner M: Untersuchungen an einigen Gymnospermen der Fränkischen Rhät-Lias-Grenzschichten. Paläontograph B 1992, 224:17-61
- Weber R: Die fossile Flora der Rhät-Lias-Übergangsschichten von Bayreuth (Oberfranken) unter besonder Berücksichtigung der Coenologie. Erl Geol Abhandl 1968, 72:1-73. Chamberlain CJ: Gymnosperms, structure and evolution New York: John-
- son Reprint Corporation; 1957.
- Sporne KR: The morphology of gymnosperms, the structure and evolution of primitive seed plants London: Hutchinson University Library; 1971.
- 33. Harris TM: Caytoniales, Cycadales & Pteridosperms, Volume 2. In The Yorkshire Jurassic Flora Edited by: Harris TM. London: Trustees of the British Museum (Natural History); 1964:1-191.
- Harris TM: Caytonia. Ann Bot 1940, 4:713-734.
- Berridge EM: On some points of resemblance between gnetalean and bennettitean seeds. New Phytol 1911, 10:140-144.
- Singh H: Embryology of gymnosperms Berlin: Gebrüder Borntraeger; 1978.
- 37. Harris JG, Harris MW: Plant identification terminology, an illustrated glossary Springer Lake: Spring Lake Publishing; 2000.
- Tomlinson PB, Takaso T: Seed cone structure in conifers in relation to development and pollination: a biological approach. Can | Bot 2002, 80:1250-1273.
- 39. Gothan W: Die unterliassische (rhätische) Flora der Umgegend von Nürnberg. Abh Nat Ges Nürnberg 1914, 19:91-186.
- Potonie H: Lehrbuch der Paläobotanik 1st edition. Berlin: Verlag von Gebrüder Borntraeger; 1921.
- 41. Emberger L: Les plantes fossiles dans leurs rapports avec les végétaux, vivants Paris: Boulevard Saint-Germain; 1944.
- Gothan W, Weyland H: Lehrbuch der Paläobotanik Berlin: Akadmie-Verlag; 1954.
- Zürlick VF: Neue Pflanzen aus dem Rhätolias. Aufschluß 1958, 9:58-60.
- Harris TM, Millington W: Ginkgoales. In The Yorkshire Jurassic Flora Edited by: Harris TM. London: Trustees of the British Museum (Natural History); 1974:1-150.
- Yang Y, Fu DZ: Review on the megastrobilus theories of conifers. Acta Phytotax Sin 2001, 39:169-191.
- Retallack G, Dilcher DL: Arguments for glossopterid ancestry of angiosperms. Palaeobiol 1981, 7:54-67.
- Thoday WG, Berridge EM: The anatomy and morphology of theinflorescences and flowers of Ephedra. Ann Bot 1912, 26:953-985.
- Tao JR, Yang Y: Alloephedra xingxueii gen. et sp. nov., an early Cretaceous member of Ephedraceae from Dalazi Formation in Yanji Basin, Jilin Province of China. Acta Palaeontol Sin 2003, **42**:208-215
- Endress PK, Igersheim A: Gynoecium structure and evolution in basal angiosperms. Int J Plant Sci 2000:S211-S223
- Yang Y, Fu DZ, Wen LH: On double fertilization in Ephedra. Adv Plant Sci 2000, 3:67-74.
- 51. Li H, Taylor EL, Taylor TN: Permian vessel elements. Science 1996, 271:188-189.
- Kryshtofovich A: Pleuromeia and Hausmannia in eastern Sibiria, with a summary of recent contribution to the palaeobotany of the region. Am J Sci 1923, 5:200-208.
- Shen GL, Gu ZG, Li KD: More material of Hausmannia ussuriensis from Jingyuan, Gansu. J Lanzhou Univ 1976, 3:71-81
- Sun G: Discovery of Dipteridaceae from the upper Triassic of eastern Jilin. Acta Palaeontol Sin 1981, 20:459-467
- Sun G: Late Triassic flora from Tianqiaoling of Jilin, China Changchun: Jilin Science & Technology Press; 1993. Li H, Tian B, Taylor EL, Taylor TN: Foliar anatomy of Gigantono-
- clea guizhouensis (Gigantopteridales) from the upper Permian of Guizhou province, China. Am J Bot 1994, 81:678-689.

- 57. Li H, Taylor DW: Aculeovinea yunguiensis gen. et sp. nov. (Gigantopteridales), a new taxon of gigantopterid stem from the upper Permian of Guizhou province, China. Int J Plant Sci 1998, **159:**1023-1033.
- 58. Thomas HH: The Caytoniales, a new group of angiospermous plants from the Jurassic rocks of Yorkshire. Phil Trans Roy Soc London 1925, 213B:299-363.
- Cornet B: Late Triassic angiosperm-like pollen from the Richmond rift basin of Virginia, USA. Paläontograph B 1989,
- 60. Cornet B, Habib D: Angiosperm-like pollen from the ammonite-dated Oxfordian (upper Jurassic) of France. Rev Palaeobot Palynol 1992, 71:269-294.
- Hochuli PA, Feist-Burkhardt S: A boreal early cradle of angiosperms? angiosperm-like pollen from the middle Triassic of the Barents Sea (Norway). J Micropalaeontol 2004, 23:97-104.
- 62. Friis EM, Pedersen KR, Crane PR: When earth started blooming: insights from the fossil record. Curr Opin Plant Biol 2005, 8:5-12
- 63. Cornet B: The leaf venation and reproductive structures of a late Triassic angiosperm, Sanmiguelia lewisii. Evol Theo 1986, 7:231-308
- 64. Taylor DW, Li H, Dahl J, Fago FJ, Zinniker D, Moldowan JM: Biogeochemical evidence for the presence of the angiosperm molecular fossil oleanane in Paleozoic and Mesozoic nonangiospermous fossils. Paleobiol 2006, 32:179-190.
- Crane PR: Comment on Cornet's Late Triassic angiospermlike pollen from the Richmond rift basin of Virginia, USA. Taxon 1987, 36:778-779.
- Martin W, Gierl A, Saedler H: Molecular evidence for pre-Cretaceous angiosperm origins. Nature 1989, 339:46-48.
- Li W-H, Gouy M, Wolfe KH, Sharp PM: Angiosperm origins. Nature 1989, 342:131-132 [http://www.nature.com/nature/journal/
- v342/n6246/pdf/342131c0.pdf].
 Soltis PS, Soltis DE: The origin and diversification of 68. angiosperms. Am J Bot 2004, 91:1614-1626.
- 69. Crepet WL, Nixon KC, Gandolfo MA: Fossil evidence and phylogeny: the age of major angiosperm clades based on mesofossil and macrofossil evidence from Cretaceous deposits. Am J Bot 2004, 91:1666-1682.
- 70. Sanderson MJ, Thorne JL, Wikström N, Bremer K: Molecular evi-
- dence on plant divergence times. Am J Bot 2004, 91:1656-1665.
 71. Anderson CL, Bremer K, Friis EM: Dating phylogenetically basal eudicots using rbcL sequences and multiple fossil reference points. Am J Bot 2005, 92:1737-1748.
- Bell CD, Soltis DE, Soltis PS: The age of the angiosperms: a molecular timescale without a clock. Evol 2005, 59:1245-1258.
- Wang X, Duan SY, Cui JZ: Several species of Schizolepis and their significance on the evolution of conifers. Taiwania 1997,
- 74. Nathorst AG: Zur mesozoischen Flora Spitzbergens gegründet auf die Sammlungen der schwedischen Expeditionen. Kungl Sven Veten Hangl 1897, 30:1-77.
- Taylor TN, Archangelsky S: The Cretaceous pteridosperms of Ruflorinia and Ktalenia and implication on cupule and carpel evolution. Am | Bot 1985, 72:1842-1853.
- Krassilov VA: Mesozoic flora of Bureya River, Ginkgoales and Czekanowskiales Moskow: Nauka; 1972.
- 77. Archangelsky S: Fossil Ginkgoales from the Tico Flora, Santa Cruz Province, Argentina. Bull Br Mus Nat Hist Geol 1965, 10:121-137.
- 78. Zhou Z, Zhang B, Wang Y, Guignard G: A new Karkenia (Ginkgoales) from the Jurassic Yima formation, Henan, China and its megaspore membrane ultrastructure. Rev Palaeobot Palynol 2002, **120:**91-105.
- Deng S, Yao Y, Ye D, Chen P, Jin F, Zhang Y, Xu K, Zhao Y, Yuan X, Zhang S: Stratum introduction, Volume 1. In Jurassic System in the North China Edited by: Guan D, Luo Z, Ma J, Deng S, Ye D, Li X, Yang S, Shen H, Yao Y, Zhao C, Zhong X, Liang H, Fu Z. Beijing: Petroleum Industry Press; 2003:1-399.
- Wang W-L, Zhang H, Zhang L-J, Zheng S-L, Yang F-L, Li Z-T, Zheng Y-J, Ding Q-H: Standard sections of Tuchengzi stage and Yixian stage and their stratigraphy, palaeontology and tectonic-volcanic actions Beijing: Geological Publishing House; 2004.

- 81. Zhang L-I, Wang L-X: Mesozoic salamander fossil-bearing strata in Reshuitang area near Lingyuan, Western Liaoning. Geol Res 2004, 13:202-206.
- 82. Zhang J: New winter crane flies (Insecta: Diptera: Trichoceridae) from the Jurassic Daohugou Formation (Inner Mongolia, China) and their associated biota. Can J Earth Sci 2006, 43:9-22.
- 83. Yang Y: Ontogeny of triovulate cones of Ephedraintermedia and origin of the outer envelope of ovules of Ephedraceae. Am | Bot 2004, 91:361-368 [http://www.amjbot.org/cgi/content/ abstract/91/3/361]
- Yang Y, Geng B-Y, Dilcher DL, Chen Z-D, Lott TA: Morphology and affinities of an early Cretaceous Ephedra (Ephedraceae) from China. Am | Bot 2005, 92:231-241 [http://www.amjbot.org/cgi/ content/abstract/92/2/231].

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