RESEARCH

Open Access

Colonization of the ocean floor by jawless vertebrates across three mass extinctions



Chase Doran Brownstein^{1*} and Thomas Near^{1,2}

Abstract

Background The deep (> 200 m) ocean floor is often considered to be a refugium of biodiversity; many benthic marine animals appear to share ancient common ancestry with nearshore and terrestrial relatives. Whether this pattern holds for vertebrates is obscured by a poor understanding of the evolutionary history of the oldest marine vertebrate clades. Hagfishes are jawless vertebrates that are either the living sister to all vertebrates or form a clade with lampreys, the only other surviving jawless fishes.

Results We use the hagfish fossil record and molecular data for all recognized genera to construct a novel hypothesis for hagfish relationships and diversification. We find that crown hagfishes persisted through three mass extinctions after appearing in the Permian ~ 275 Ma, making them one of the oldest living vertebrate lineages. In contrast to most other deep marine vertebrates, we consistently infer a deep origin of continental slope occupation by hagfishes that dates to the Paleozoic. Yet, we show that hagfishes have experienced marked body size diversification over the last hundred million years, contrasting with a view of this clade as morphologically stagnant.

Conclusion Our results establish hagfishes as ancient members of demersal continental slope faunas and suggest a prolonged accumulation of deep sea jawless vertebrate biodiversity.

Keywords Hagfishes, Phylogenetics, Jawless Vertebrates, Continental Slope, Habitat

Introduction

The asymmetrical accumulation of ancient biodiversity across ecosystems has fascinated biologists for centuries. Charles Darwin recognized that several ancient clades he dubbed 'living fossils' all lived in freshwater ecosystems and suggested that these habitats might offer refuge to lineages that have faced competition and extinction elsewhere [1]. Since the biota of continental slope and abyssal oceanic ecosystems (>200 m depth) was first documented, these complex habitats have been regarded as oases for very old lineages that have since

Chase Doran Brownstein

become depauperate or wholly extinct in shallow-water environments. Classic examples of ancient deep-sea lineages include clades as varied as coelacanths (Latimeria) [2], the Vampire Squid Vampyroteuthis infernalis [3, 4], crinoids and brittlestars in Echinodermata [5-7], black corals [8], the living graptolite genus Rhabdopleura and other colonial hemichordates [9–12], and potentially several lineages of deep-sea isopods [13–15], all of which last share common ancestry with related shallow-water and terrestrial forms well over 200 million years ago. At the same time, recent work has shown that these ancient components of the deep sea biota do not represent a simple accumulation, or sink, of remnant biodiversity over geological time [16]. Rather, many lineages have undergone extensive diversification in continental slope, bathypelagic, and abyssal ecosystems [7, 8, 17, 18].

Vertebrates, which comprise tens of thousands of species distributed across all major ecosystems, underwent



© The Author(s) 2024. **Open Access** This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit http://creativecommons.gr/licenses/by/4.0/. The Creative Commons Public Domain Dedication waiver (http://creativecommons.gr/licenses/by/4.0/. The CreativeCommons Public Domain Dedication waiver (http://creativecommons.gr/licenses/by/4.0/. The CreativeCommons.gr/licenses/by/4.0/. The CreativeCommons Public Domain Dedication waiver (h

^{*}Correspondence:

chase.brownstein@yale.edu

¹ Department of Ecology and Evolutionary Biology, Yale University, New Haven, CT 06511, USA

² Yale Peabody Museum, Yale University, New Haven, CT 06511, USA

their initial diversification in marine habitats during the Ordovician to Devonian Periods, around 480-360 million years ago [19–21]. Despite this ancient history in the ocean, evidence from the fossil record [22] and molecular phylogenetics [23–34] suggests that most living deep marine vertebrate diversity originates from radiations that took place over the last 100 million years. This observation raises the question of whether any living vertebrate diversity in the ocean benthos can truly be considered geologically ancient.

By end of the Devonian, jawed vertebrates had eclipsed the once dominant jawless lineages in diversity and morphological disparity [21, 35, 36]. Presently, only two lineages of jawless vertebrates, lampreys and hagfishes, survive. The resolution of living jawless vertebrates relationships and divergence times is essential for understanding the evolutionary context of living vertebrate diversity [36–42], but these remain disputed [9, 37, 39, 42–55]; only in the past few years have comparative genomic analyses provided strong support for a hagfishlamprey sister relationship [45, 48].

Hagfishes (Myxiniformes) are a globally-distributed clade of eel-like demersal marine species that are important for comprising a large proportion of vertebrate biomass in ocean floor ecosystems [50, 56]. Although several hagfishes live on the continental shelf [56, 57], most species periodically or permanently occupy continental slope habitats at depths over 200 and up to 3000 m [56– 62], where they are ecologically important predators and scavengers [56, 63].

The hagfish body plan, which includes features like a poorly developed eye and skeleton, auxiliary hearts, and a single semicircular canal, departs so much from other vertebrates that this clade has often been placed as the sister to all other living vertebrate clades [35, 42, 49, 53, 64]. Most molecular, morphological, and combined evidence analyses posit that hagfishes are the sister clade of lampreys [9, 37–39, 44–48, 51, 52, 65–68]. However, the interrelationships of hagfishes and the age of living hagfish diversity remain underexplored [37, 38, 40, 43, 69]. This problem is compounded by the poor fossil record of the two living jawless vertebrate clades [36-39, 64-67, 64-67]70-74]; only a handful of fossils with well-characterized anatomy are identifiable as early total clade lampreys and hagfishes, and many fossil taxa identified as putative stem-group hagfishes and lampreys [75] may not even be jawless vertebrates [76–78].

Here, we pair data from the fossil record with a dataset including all genera of living hagfishes and $\sim 60\%$ of the species diversity to reconstruct the interrelationships of hagfishes and their tempo of diversification. By using fossils to produce a tip-dated phylogeny of living jawless vertebrates, we recover an age for the most recent common

ancestor of hagfishes that is over twice as old as previously suggested [37, 38, 51]. Our results show that living hagfishes compose one of the oldest major vertebrate clades, with isolated species potentially representing up to 160 million years of unique evolutionary history. Ancestral state reconstructions support an ancient origin for continental slope ecologies in living hagfishes. These results suggest that hagfishes are an ancient vertebrate clade and clarify their diversification in the deep sea through at least three mass extinctions.

Methods

Sequence dataset collection

To produce a comprehensive hypothesis of hagfish interrelationships, we aimed to maximize the number of species sampled. Previous taxonomic and phylogenetic studies concentrated on the mitochondrial genes COI and 16S ribosomal DNA [40, 57, 60, 69, 79] and we gathered sequences of these two genes for all available species (n=44) on GenBank. Our taxon sampling included two species of Rubicundus, two species of Neomyxine, 14 species of Myxine, and 26 species of Eptatretus, with an additional three potentially distinct species of Eptatretus from India, Japan, and Korea. Our sample consists of all recognized genera and over 50% of species diversity (https://www.calacademy.org/scientists/projects/ eschmeyers-catalog-of-fishes), including the problematic 'Notomyxine' (=Myxine) tridentiger and several species previously classified in 'Quadratus' and 'Paramyxine.' Nemamyxine, which includes two species known from preserved specimens collected in the twentieth century [80], does not have any available genetic material. Thus, its relationships are untestable with molecular phylogenetics. We note that the specimens assigned to Nemamyxine have potential affinities to Rubicundus, and that the current diagnosis for Nemamyxine, which includes an extremely slender body and an anteriorly placed ventral finfold that originates anterior of the ventral gill apertures, is not sufficient to differentiate this lineage from other hagfishes. First, numerous species in Myxine, Rubicundus, and Eptatretus, as well as the extinct species *+Tethymyxine tapirostrum*, show a highly elongated body [37, 40]. Further, slender body depth and high slime pore counts were cited as diagnostic characters for both species of Nemamyxine, and both these features are widely distributed among other elongated hagfishes [37, 40]. The placement of the finfold relative to the apertures is a more convincing character. Thus, without genetic material, the phylogenetic position of Nemamyxine remains unclear and the available diagnosis is largely uninformative. All sequence data, along with corresponding numbers in GenBank, is included in the Supplementary Data.

Gene tree inferences

The DNA sequences of the COI and 16S gene were aligned by eye using the translated amino acid sequences as a guide. We also used the online Clustal Omega tool at the EMBL-EBI online resource portal to aid in 16S alignments (www.ebi.ac.uk/Tools/msa/clustalo/). Phylogenies were inferred using both maximum likelihood and Bayesian methods. We used the gnathostomes Polypterus ornatipinnis, Protopterus annectens, and Callorhincus milii and the lampreys Geotria australis, Petromyzon marinus, and Lampetra fluviatilis as outgroups for the analysis of the COI gene and rooted the 16 s tree on Rubicundus. Maximum likelihood analyses were conducted on each of the COI and 16S alignments and with both concatenated together using the software IQTREE v. 2.2.0.3 [81] with branch support assessed using 100 standard bootstrap replicates. We allowed IQTREE to find the optimal partitions and molecular evolutionary models using AIC values via ModelFinder [82]. Preferred models were the TN93+F+I+G4 model for COI and TIM2+F+I+G4 for 16S. Bayesian analyses were conducted using the program MrBayes v. 3.2 [83] using the GTR+G evolutionary model. Analyses were run for 1.0×10^7 generations, and two simultaneous runs were conducted within each of four chains. We assessed chain convergence and stationarity by inspecting chain likelihoods and monitoring average standard deviations of split frequencies between the two runs to ensure values less than 5.0×10^{-3} after 1.0 X 10⁶ generations. We discarded the first 50% of sampled generations as burn-in and summarized the posterior tree set in a 50% majorityrule tree.

Bayesian tip-dating analyses

We jointly estimated the phylogenetic relationships and divergence times of hagfishes and a subset of lampreys using a tip-dating approach as implemented in the program BEAST 2.6.6 [84, 85] using the fossilized birthdeath (FBD) model [86]. Lampreys were included to allow for use of several fossils as tip-calibrations and because they are phylogenetically proximal to hagfishes among living vertebrates. We generated modified input molecular data files that included all hagfish species for which DNA sequences were available and our included fossil tip calibrators whose phylogenetic positions we constrained based on the results of previous studies using monophyletic MRCA priors. A complete list of calibrations, along with age and placement justifications, is included in the Supplementary Information. In total, we selected two fossil lampreys (one total group and one crown group), one crown-group fossil hagfish, and putative stem-hagfish *†Myxinikela siroka* as fossil tip calibrators. We set the origin prior at 439.0 Ma (bounds of 400 and 600 Ma), which is the age of the oldest gnathostome *+Fanjingsha*nia renovata from the early Silurian of China [87], to account for the uncertainty surrounding the monophyly of cyclostomes. We applied a TN93 model of nucleotide evolution to our COI sequence data and a GTR model to our 16S sequence data following the model choices of the IQTREE analysis. We set the rho parameter of the FBD model to 0.57, which is the proportion of known living species included in the dataset, and set the diversification rate prior to 0.1 based on the ratio of living species in the dataset to the origin prior, with bounds of 0.0 and infinity. We used a gamma prior with a default value of 1.0 for the mean and 0.33 for the standard deviation, and relaxed uncorrelated lognormal clock model. Two BEAST runs were conducted over 1.0 X 108 generations with 1.0 X 10^7 pre-burnin, and convergence of the posteriors was checked using Tracer v. 1.7.1 [88]. We combined the posterior tree sets in LogCombiner v. 2.6.6 with 10% burnins and summarized them in a maximum clade credibility (MCC) tree with median node heights using TreeAnnotator 2.6.4. We conducted a set of three BEAST analyses without using the putative stem-hagfish +Myxinikela *siroka* as a tip calibration for the hagfish total clade to test its influence on divergence time estimates. Finally, we conducted BEAST analyses of including and excluding *†Myxinikela siroka* as a tip calibration but only sampling from prior values, in order to test whether our divergence time estimates for the analyses using sequence data were driven by only the priors. We compared the posterior mean and 95% highest posterior density intervals for the divergence times of selected major hagfish clades across both iterations of our BEAST analyses (Fig. 2). All input xml files and output files for both BEAST run iterations are included in the Supplementary Information.

In order to assess the phylogenetic informativeness of the sequences we used for reconstructing ancient relationships in hagfishes, we used the program hyphy as implemented through the phydesign online tool [89, 90] to plot the phylogenetic informativeness of both mitochondrial loci used in this study for reconstructing hagfish relationships through time.

Age comparisons

We compared the estimated ages of hagfish divergences to previous ones [37, 51] (Table S1) and the ages of the hagfish and lamprey crown groups estimated from the Bayesian tip-dating analysis including †*Myxinikela siroka* to other major vertebrate clades by extracting crown age estimates from TimeTree.org [91] We recorded the age of the MRCA of living gars from our recent relaxed molecular clock estimation [92]. The complete list of crown ages are listed in Table S2.

Ancestral habitat reconstruction

In order to assess the evolution of depth preference among hagfishes, we assembled a dataset from FishBase and the literature (Table S3) on observed depth ranges for hagfish species. Values of 200 m or less were coded as continental shelf ranges, and values of over 200 m were coded as continental slope regions following the literature [8, 93, 94]. Most studies attempting to reconstruct habitat preference evolution have either divided habitat characters into a discrete states [7, 93, 95-98] or treated different habitats as independent areas [8]. However, because ecological character states can form a spectrum and species can show polymorphic ecological states (i.e., appearing in more than one habitat), modeling overlap as discrete characters (e.g., denoting slope, shelf, and shelfslope as distinct, discrete characters) is nonideal [99]. To account for this problem, we used the R package phytools [100] to perform stochastic ancestral character mapping where habitat preference was treated as a polymorphic character using the fitpolyMk function with unordered transition rates ("ARD" model) and the root prior distribution π proposed by Fitzjohn et al. [101]. Given that there were only two primary states (shelf, slope), we selected the ARD model a priori because it is the most generalized unordered model available for polymorphic traits in phytools. The results of fitpolyMk were used for stochastic character mapping over 1000 simulated topologies, and the results were summarized into a single reconstruction (Fig. 1). All code necessary to replicate this analysis is in the supplement.

Results

Phylogeny and divergence times of Myxiniformes

Maximum likelihood and Bayesian analyses of our hagfish sequence dataset supported the reciprocal monophyly of three major lineages (Fig. 1, Figure S1-S4): Rubicundinae, Eptatretinae and Myxininae. *Neomyxine* is resolved as the sister lineage of *Myxine* (Fig. 1a, Figure S1, Figure S2, Figure S3). The resolution of Rubicundinae as the sister clade of all other hagfishes is consistent with some previous phylogenetic analyses [37, 38, 40, 60] but incongruent with other studies that variably resolve *Neomyxine* as the sister lineage of all other hagfishes [79].

(See figure on next page.)

Our phylogenetic results affirm the synonymy of several genera of hagfishes, including *Quadratus* and *Paramyx-ine*, with *Eptatretus* and the enigmatic *Notomyxine tri-dentiger* with *Myxine* (Fig. 1a) [40, 57, 79].

Our tip-dated Bayesian analyses of hagfish phylogeny consistently recovered Paleozoic ages for the major hagfish crown clades (Fig. 1; Fig. 2). Relaxed clock analyses using BEAST with the inclusion and exclusion of †Myx*inikela siroka* resulted in closely comparable posterior divergence time estimates for all major hagfish lineages (Fig. 2A), demonstrating that the tip-dating scheme employed in this study is robust to the use of putative Mazon Creek hagfishes as calibrations. This is especially important given that we excluded the wildcard taxon.

+Gilpichthys greenei, known from numerous poorly preserved specimens from the Mazon Creek, from all of our analyses. Recent phylogenetic analyses place +G. greenei as an indeterminate jawless vertebrate [37, 49, 66, 67] or as the most stemward hagfish [38, 53, 73]. Because +G. greenei is classically united with Myxiniformes based on the *absence* of features like mineralized teeth, the placement of this taxon among early vertebrates might be biased by the phenomenon of stemward slippage, whereby decomposition of the body can lead to erroneous interpretations of a fossil's phylogenetic affinities [102, 103].

Across different analyses, we consistently estimate an Early Permian origin for the hagfish crown and an early Triassic age for the split between Eptatretus and Myxininae (Fig. 1, Fig. 2A). These age ranges are substantially older than previous estimates of hagfish diversification (Fig. 2A), which invariably place the origins of the major living hagfish clades far later in the middle to Late Cretaceous [10, 17, 33]. Although we use mitochondrial gene sequences (mtDNA), which have been shown to overestimate divergence times for ray-finned fish ingroups relative to nuclear DNA sequences [104], sequence data type cannot explain the discrepancy between our results and previous estimates, which also used mtDNA [37]. Instead, our results are best explained by a combination of more extensive hagfish species sampling (including several deeply-divergent singletons like Eptatretus cheni) and more stringent criteria for including fossils as tip

Fig. 1 Hagfish phylogeny and tempo of diversification. Tip-dated Bayesian maximum clade credibility phylogeny of jawless fishes from two independent runs in BEAST 2.6.6 showing the interrelationships of the major lineages of hagfishes. Bars indicate 95% highest posterior density (HPD) intervals for divergence times at nodes. Outgroups not shown. Grey bars are at nodes supported by posterior values of 0.90 or more, clear bars are at nodes supported by posterior values of 0.89 or less. Gray columns indicate mass extinction events. Dagger (†) indicates extinct species known from the fossil record. Pie charts indicate ancestral state reconstructions of habitat for each node, where purple represents the probability of a slope component (either slope or shelf-slope) at each node and salmon indicates the probability of continental shelf habitat being ancestral. Inset includes the transition matrix from the polymorphic character ancestral reconstruction analysis (note that purple here is exclusively slope, as opposed to purple denoting slope/shelf-slope at nodes in the phylogeny). Photograph of *Eptatretus stoutii* is courtesy Douglas Fudge





Fig. 2 The Paleozoic age of the hagfish radiation. Boxplots (**A**) generated using the R package ggplot2 for divergence times of major hagfish lineages under different fossil calibration schemes and those reported in a previous study. Note the similar ages estimated for these lineages regardless of the inclusion of *†Myxinikela siroka* as a fossil tip calibration, and the much older ages for all major hagfish clades except Rubicundinae estimated in this study compared to previous estimates based on smaller molecular datasets and less-inclusive taxon sampling. Panel (**B**) shows the ages of major vertebrate crown groups. Hagfishes are the third oldest, exceeded only by the far more species rich and morphologically disparate lissamphibians (frogs, salamanders, and caecilians) and chondrichthyans (sharks, rays, skates, and chimaeras). Silhouettes are public from phylopic.org, Wikimedia commons, and by CDB

calibrations. For example, +*Gilpichthys greenei*, a problematic fossil chordate [37, 38, 73] from the Mazon Creek Lagerstätten, was excluded in our analyses but placed as a stem-lamprey with little evidence in the previous phylogenetic analyses [37]. Phylogenetic informativeness analyses [89] suggest that, while the informativeness of *COI* declines for hagfish divergences over 100 million years ago, *16S* provides steady (albeit reduced relative to *COI*) phylogenetic information across the age of hagfish evolutionary history (Fig. 3).

The revised timescale of hagfish diversification (Figs. 1 and 2) reduces the long branch separating living hagfishes from other vertebrates by over 120 million years [9, 10, 17, 19, 33, 44] and demonstrates that the hagfish crown clade has persisted through the last three mass extinctions. Further, our age estimates place crown hagfishes among the oldest vertebrate crown clades (Fig. 2B), suggesting that living hagfish diversity is far more ancient than most other marine vertebrates. These age estimates are not attributable to our choice of prior settings, as runs sampling only from priors produce far older ages for hagfish divergence times.

Ancestral habitats of hagfishes

Ancestral habitat reconstruction along the time-calibrated phylogeny indicates that hagfishes have occupied continental slope habitats with depths of over 200 m since the origins of the crown group during the late Paleozoic (Fig. 1). Although there is a high degree of variability in living hagfish habitat utilization and fossil hagfishes are only known from estuarine and continental shelf settings (depth < 200 m) [37, 38], polymorphic character ancestral state reconstruction favors a continental slope component (either slope or shelf + slope) as ancestral for nearly all deep nodes in crown hagfish phylogeny (Fig. 1; note the root node for Myxiniforms is less clearly resolved). Continental slope components are more strongly inferred for the ancestral habitat of crown *Rubicundus* and several recent divergences inside *Eptatretus* (Fig. 1), whereas the continental shelf appears to be supported as the ancestral habitat for the clade containing *Myxine fernholmi* and *Myxine affinis*.

Discussion

Paleozoic origins of hagfish diversity

Since the end of the Triassic, only hagfishes and lampreys have persisted as the survivors of the once more diverse grade of jawless vertebrates [36, 37, 53–55]. This accident of deep time makes these two clades particularly important for comparative studies of phylogenetic relationships of early vertebrates and the context of their diversification [36, 37, 48, 51]. Because the anatomy of lampreys and hagfishes diverges considerably from other living vertebrates and both lineages have poor fossil records, molecular phylogenetics provides key information for reconstructing the relationships and timescale of diversification in living jawless vertebrates.

Our hypothesis of hagfish phylogeny and relaxed molecular clock divergence time estimates reveals an ancient origin of crown Myxiniformes during the Permian period (Fig. 1, Fig. 2). We infer that the initial divergences of living hagfishes occurred in the Paleozoic and



Fig. 3 Phylogenetic Informativeness Analysis. Plot shows the phylogenetic informativeness of both mitochondrial loci used in this study through time. The grey shaded area represents the 95% HPD interval for crown Myxiniformes found in the tip-dated phylogeny where †*Myxinikela siroka* was used as a fossil tip calibration. Photograph of *Eptatretus stoutii* is courtesy Douglas Fudge

earliest Mesozoic, only 20 to 30 million years after the first putative hagfishes appear in the fossil record [38]. This old age for hagfish diversity highlights a hidden period of jawless fish radiation that followed the extinction of 'ostracoderms,' a grade of armored, jawless fishes that formed the dominant assemblage of vertebrates until the Devonian [35, 36, 53, 54, 105].

The body plan of hagfishes remains highly conserved and includes specializations such as deskeletonization [37, 106], a rudimentary visual system [107, 108], burrowing and knotting feeding behavior [56, 63], and tolerance to high ammonia concentrations, such as those from carrion [109]. The low rate of morphological change observed in hagfishes provides a notable contrast to similarly old clades of vertebrates that exhibit high phenotypic diversity and species richness, such as teleosts, chondrichthyans, and lissamphibians (Fig. 3a-c) [24, 25, 110–115]. The low morphological disparity observed among living hagfishes and the ancient age inferred for the crown also imply that the specialized anatomy of this clade appeared by the end of the Paleozoic.

The tempo of hagfish diversification contrasts with the pattern observed in lampreys, the only other living clade of jawless vertebrates [36, 39], which include numerous regional radiations that have diversified over the past

100 million years [36]. In contrast, our time-calibrated phylogeny of hagfishes infers an average evolutionary interval of 31.6 million years of common ancestry for individual hagfish species, which is considerably higher than the corresponding values for cartilaginous and bony fishes [115] or the one-to-two-million-year divergences of most lamprey species pairs [36]. The most isolated single branch on the hagfish tree leads to Eptatretus cheni (Fig. 1a), which we estimate diverged from all other species of *Eptatretus* during the Jurassic Period (Fig. 1: median MRCA age=167.24 Ma, 95% HPD: 101.77, 255.65 Ma). The age of this single branch is comparable to the most evolutionarily isolated species among sharks, rays, and chimeras [115], as well as long branches like the Tuatara Sphenodon punctatus [111, 116] and the Salamanderfish Lepidogalaxias salamandroides [26, 28, 30].

Hagfishes are ancient inhabitants of continental slope settings

The phylogenetic hypothesis of hagfishes presented in this paper highlights them as a trove of ancient vertebrate evolutionary history hidden in oceanic demersal habitats. The true diversity of living hagfishes remains a frontier of biological research but is challenged by their deep marine habitats. For example, *Rubicundus* is the living sister lineage of all other hagfishes (Fig. 1, Figure S1-S3) and has a nearly cosmopolitan geographic distribution, but was identified as a distinct genus and described in the last decade [40]. In addition, species discovery in hag-fishes continues at a pace as 15% of recognized species in the clade were described over the past ten years [117]; several new forms were recovered at depths in the thousands of meters [60].

Ancestral state reconstructions of habitat along the time-calibrated hagfish phylogeny that we present in this paper (Fig. 1) demonstrate an ancient history of continental slope habitat use in Myxiniformes. Despite levels of uncertainty introduced by the variability of habitat use among hagfish subclades (including among species; Fig. 1; [62]), we infer that hagfishes have accessed continental slope settings in the deep sea since at least the Permian period (Fig. 1), with several subclades in Myxine and Eptatretus diversifying recently in continental shelf settings. This makes hagfishes the vertebrate clade with the most ancient history in the deep sea, far outpacing the ages of deep-sea diversifications in teleost fishes [23, 24, 26, 31, 32, 34, 93] or chondrichthyans [118]. The deep-sea habitats reconstructed as ancestral for hagfishes complicate the current picture of early vertebrate diversification. Although evidence from the fossil record, including fossil hagfishes [38], indicate that early vertebrates primarily diversified in coastal environments [21, 119, 120], our results suggest that exploration of the deep sea was occurring in at least one clade during the Paleozoic.

Deep sea cradles and refugia of hagfish diversity

One possibility is that demersal marine habitats have served as a refugium for the diversity of hagfishes. Although similarly old marine vertebrate clades such as cartilaginous fishes did experience declines in phenotypic disparity, they show less pronounced species turnover than other lineages during major events like the Cretaceous-Paleogene mass extinction [121-124]. Some of the oldest surviving chondrichthyan clades, such as the goblin shark, frilled and sevengill sharks, and the chimeras and ratfishes, occupy the deep sea, underscoring these habitats as a refugium of ancient diversity in cartilaginous fishes [115]. Consequently, the evolutionary history of hagfishes provides an intriguing parallel with longlived, species-poor lineages from freshwater ecosystems often referred to as living fossils, which are classically thought to have been buffered from extinction by their environments [1, 92, 125-128].

At the same time, our time-calibrated phylogeny suggests that considerable speciation (Fig. 1) has taken place in continental slope environments over the past 100 million years. Despite the old age of the hagfish crown group and the high proportion of species with ancient unique evolutionary histories, our results highlight that Myxiniformes includes ancient species diversity that has also undergone significant diversification since first appearing in the marine benthos.

Conclusions

Vertebrate diversity in the deep sea is mostly young (<100 million years old) relative to the ancient ages of many continental slope and abyssal invertebrate radiations. This observation contrasts with the hypothesis that the deep sea has been a refugium for animal diversity, and also begs the question of whether any deep sea vertebrates are ancient endemics. Here, by reconstructing the phylogeny and divergence times of hagfishes, we substantiate this clade as a truly ancient lineage with a long history in continental slope settings (>200 m depth). The evolutionary history of hagfishes reconstructed in this paper markedly contrasts with that of the only other surviving jawless vertebrate clade, the lampreys, and implies that vertebrate colonization of habitats classically considered part of the deep sea [94] has taken place since the Paleozoic.. Our results highlight hagfishes as a unique vector of ancient vertebrate diversity that has persisted and diversified in the deep sea.

Supplementary Information

The online version contains supplementary material available at https://doi.org/10.1186/s12862-024-02253-y.

Supplementary Material 1.

Acknowledgements

We thank Douglas Fudge for providing the image of the hagfish used in Figure 1. We also wish to thank several anonymous reviewers for their comments on a previous draft of this paper.

Authors' contributions

C.D.B. collected the data, ran and interpreted the analyses, and wrote and edited drafts of the paper. T.J.N. administered the project, interpreted the analyses, and wrote and edited drafts of the paper.

Funding

C.D.B. was supported by the Yale College Richter Fellowship (2023). T.J.N. was supported by the Bingham Oceanographic Fund of the Yale Peabody Museum.

Availability of data and materials

All data is in the supplementary information submitted with this article. No new sequence data were generated for this study.

Declarations

Ethics approval and consent to participate

N/A.

Consent for publication

N/A.

Competing interests

The authors declare no competing interests.

Received: 16 January 2024 Accepted: 7 May 2024 Published online: 13 June 2024

References

- Darwin C, Kebler I. On the origin of species by means of natural selection, or, The preservation of favoured races in the struggle for life. London: J. Murray; 1859.
- Amemiya CT, Alföldi J, Lee AP, Fan S, Philippe H, MacCallum I, et al. The African coelacanth genome provides insights into tetrapod evolution. Nature. 2013;496:311–6.
- Tanner AR, Fuchs D, Winkelmann IE, Gilbert MTP, Pankey MS, Ribeiro ÂM, et al. Molecular clocks indicate turnover and diversification of modern coleoid cephalopods during the Mesozoic marine revolution. Proc Royal Soc B: Biol Sci. 2017;284:20162818.
- 4. Whalen CD, Landman NH. Fossil coleoid cephalopod from the Mississippian Bear Gulch Lagerstätte sheds light on early vampyropod evolution. Nat Commun. 2022;13:1107.
- Mongiardino Koch N, Thompson JR, Hiley AS, McCowin MF, Armstrong AF, Coppard SE, et al. Phylogenomic analyses of echinoid diversification prompt a re-evaluation of their fossil record. eLife. 2022;11:e72460.
- Thuy B, Gale AS, Kroh A, Kucera M, Numberger-Thuy LD, Reich M, et al. Ancient origin of the modern deep-sea fauna. PLoS ONE. 2012;7:e46913.
- Bribiesca-Contreras G, Verbruggen H, Hugall AF, O'Hara TD. The importance of offshore origination revealed through ophiuroid phylogenomics. Proc Royal Soc B: Biol Sci. 2017;284:20170160.
- 8. Horowitz J, Quattrini AM, Brugler MR, Miller DJ, Pahang K, Bridge TCL, et al. Bathymetric evolution of black corals through deep time. Proc Royal Soc B: Biol Sci. 2023;290:20231107.
- Simakov O, Kawashima T, Marlétaz F, Jenkins J, Koyanagi R, Mitros T, et al. Hemichordate genomes and deuterostome origins. Nature. 2015;527:459–65.
- Nanglu K, Caron J-B, Cameron CB. Cambrian Tentaculate Worms and the Origin of the Hemichordate Body Plan. Curr Biol. 2020;30:4238-4244. e1.
- 11. Briggs DEG, Koch NM. A Silurian pseudocolonial pterobranch. Curr Biol. 2023;33:5225-5232.e3.
- Nanglu K, Waskom ME, Richards JC, Ortega-Hernández J. Rhabdopleurid epibionts from the Ordovician Fezouata Shale biota and the longevity of cross-phylum interactions. Commun Biol. 2023;6:1–10.
- Raupach MJ, Mayer C, Malyutina M, Wägele J-W. Multiple origins of deep-sea Asellota (Crustacea: Isopoda) from shallow waters revealed by molecular data. Proc Royal Soc B: Biol Sci. 2008;276:799–808.
- 14. Wilson GDF. Some of the Deep-Sea Fauna Is Ancient. Crustaceana. 1999;72:1019–30.
- 15. Lins LSF, Ho SYW, Wilson GDF, Lo N. Evidence for Permo-Triassic colonization of the deep sea by isopods. Biol Let. 2012;8:979–82.
- Rex MA, McClain CR, Johnson NA, Etter RJ, Allen JA, Bouchet P, Warén A. A source-sink hypothesis for abyssal biodiversity. Am Nat. 2005;165(2):163–78. https://doi.org/10.1086/427226.
- Campoy AN, Rivadeneira MM, Hernández CE, Meade A, Venditti C. Deep-sea origin and depth colonization associated with phenotypic innovations in scleractinian corals. Nat Commun. 2023;14:7458.
- Lindner A, Cairns SD, Cunningham CW. From offshore to onshore: multiple origins of shallow-water corals from deep-sea ancestors. PLoS ONE. 2008;3:e2429.
- 19. Brazeau MD, Friedman M. The origin and early phylogenetic history of jawed vertebrates. Nature. 2015;520:490–7.

- Sallan LC, Coates MI. End-Devonian extinction and a bottleneck in the early evolution of modern jawed vertebrates. Proc Natl Acad Sci U S A. 2010;107:10131–5.
- 21. Sallan L, Friedman M, Sansom RS, Bird CM, Sansom IJ. The nearshore cradle of early vertebrate diversification. Science. 2018;362:460–4.
- Baucon A, Ferretti A, Fioroni C, Pandolfi L, Serpagli E, Piccinini A, et al. The earliest evidence of deep-sea vertebrates. Proc Natl Acad Sci. 2023;120:e2306164120.
- Miya M, Pietsch TW, Orr JW, Arnold RJ, Satoh TP, Shedlock AM, et al. Evolutionary history of anglerfishes (Teleostei: Lophiiformes): a mitogenomic perspective. BMC Evol Biol. 2010;10:58.
- Ghezelayagh A, Harrington RC, Burress ED, Campbell MA, Buckner JC, Chakrabarty P, et al. Prolonged morphological expansion of spiny-rayed fishes following the end-Cretaceous. Nat Ecol Evol. 2022;6:1211–20.
- Near TJ, Dornburg A, Eytan RI, Keck BP, Smith WL, Kuhn KL, et al. Phylogeny and tempo of diversification in the superradiation of spiny-rayed fishes. Proc Natl Acad Sci. 2013;110:12738–43.
- Near TJ, Eytan RI, Dornburg A, Kuhn KL, Moore JA, Davis MP, et al. Resolution of ray-finned fish phylogeny and timing of diversification. Proc Natl Acad Sci USA. 2012;109:13698–703.
- Alfaro ME, Faircloth BC, Harrington RC, Sorenson L, Friedman M, Thacker CE, et al. Explosive diversification of marine fishes at the Cretaceous-Palaeogene boundary. Nat Ecol Evol. 2018;2:688–96.
- Betancur-R R, Broughton RE, Wiley EO, Carpenter K, López JA, Li C, et al. The tree of life and a new classification of bony fishes. PLoS Curr. 2013. https://doi.org/10.1371/currents.tol.53ba26640df0ccaee75bb165c 8c26288.
- 29. Betancur-R R, Wiley EO, Arratia G, Acero A, Bailly N, Miya M, et al. Phylogenetic classification of bony fishes. BMC Evol Biol. 2017;17:162.
- Hughes LC, Ortí G, Huang Y, Sun Y, Baldwin CC, Thompson AW, et al. Comprehensive phylogeny of ray-finned fishes (Actinopterygii) based on transcriptomic and genomic data. Proc Natl Acad Sci U S A. 2018;115:6249–54.
- Kenaley CP, DeVaney SC, Fjeran TT. The complex evolutionary history of seeing red: molecular phylogeny and the evolution of an adaptive visual system in deep-sea dragonfishes (stomiiformes: Stomiidae). Evolution. 2014;68:996–1013.
- Davis MP, Fielitz C. Estimating divergence times of lizardfishes and their allies (Euteleostei: Aulopiformes) and the timing of deep-sea adaptations. Mol Phylogenet Evol. 2010;57:1194–208.
- Inoue JG, Miya M, Miller MJ, Sado T, Hanel R, Hatooka K, et al. Deepocean origin of the freshwater eels. Biol Lett. 2010;6:363–6.
- Denton JSS. Diversification Patterns of Lanternfishes Reveal Multiple Rate Shifts in a Critical Mesopelagic Clade Targeted for Human Exploitation. Curr Biol. 2018;28:933-940.e4.
- Forey P, Janvier P. Agnathans and the origin of jawed vertebrates. Nature. 1993;361:129–34.
- 36. Brownstein CD, Near TJ. Phylogenetics and the Cenozoic radiation of lampreys. Curr Biol. 2023;33:397-404.e3.
- Miyashita T, Coates MI, Farrar R, Larson P, Manning PL, Wogelius RA, et al. Hagfish from the Cretaceous Tethys sea and a reconciliation of the morphological–molecular conflict in early vertebrate phylogeny. Proc Natl Acad Sci. 2019;116:2146–51.
- Miyashita T. A Paleozoic stem hagfish Myxinikela siroka revised anatomy and implications for evolution of the living jawless vertebrate lineages. Can J Zool. 2020;98:850–65.
- 39. Wu F, Janvier P, Zhang C. The rise of predation in Jurassic lampreys. Nat Commun. 2023;14:6652.
- Fernholm B, Norén M, Kullander SO, Quattrini AM, Zintzen V, Roberts CD, et al. Hagfish phylogeny and taxonomy, with description of the new genus Rubicundus (Craniata, Myxinidae). J Zool Syst Evol Res. 2013;51:296–307.
- Pascual-Anaya J, Sato I, Sugahara F, Higuchi S, Paps J, Ren Y, et al. Hagfish and lamprey Hox genes reveal conservation of temporal colinearity in vertebrates. Nat Ecol Evol. 2018;2:859–66.
- 42. Near TJ. Conflict and resolution between phylogenies inferred from molecular and phenotypic data sets for hagfish, lampreys, and gnathostomes. J Exp Zool B Mol Dev Evol. 2009;312:749–61.
- Bardack D. Relationships of Living and Fossil Hagfishes. In: Jørgensen JM, Lomholt JP, Weber RE, Malte H, editors. The Biology of Hagfishes. Dordrecht: Springer, Netherlands; 1998. p. 3–14.

- 44. Delarbre C, Gallut C, Barriel V, Janvier P, Gachelin G. Complete mitochondrial DNA of the hagfish, Eptatretus burgeri: the comparative analysis of mitochondrial DNA sequences strongly supports the cyclostome monophyly. Mol Phylogenet Evol. 2002;22:184–92.
- 45. Yu D, Ren Y, Uesaka M, Beavan AJ, Muffato M, Shen J, Li Y, Sato I, Wan W, Clark JW, Keating JN. Hagfish genome elucidates vertebrate wholegenome duplication events and their evolutionary consequences. Nat Ecol Evol. 2024:1–17. in press.
- Heimberg AM, Cowper-Sallari R, Sémon M, Donoghue PCJ, Peterson KJ. microRNAs reveal the interrelationships of hagfish, lampreys, and gnathostomes and the nature of the ancestral vertebrate. Proc Natl Acad Sci. 2010;107:19379–83.
- Kuraku S, Hoshiyama D, Katoh K, Suga H, Miyata T. Monophyly of lampreys and hagfishes supported by nuclear DNA-coded genes. J Mol Evol. 1999;49:729–35.
- Marlétaz F, Timoshevskaya N, Timoshevskiy VA, Parey E, Simakov O, Gavriouchkina D, Suzuki M, Kubokawa K, Brenner S, Smith JJ, Rokhsar DS. The hagfish genome and the evolution of vertebrates. Nature. 2024:1–3. in press.
- Tian Q, Zhao F, Zeng H, Zhu M, Jiang B. Ultrastructure reveals ancestral vertebrate pharyngeal skeleton in yunnanozoans. Science. 2022;377:218–22.
- 50. Hardisty MW. Biology of the Cyclostomes. Boston: Springer, US; 1979.
- Kuraku S, Kuratani S. Time scale for cyclostome evolution inferred with a phylogenetic diagnosis of hagfish and lamprey cDNA sequences. Zoolog Sci. 2006;23:1053–64.
- Delsuc F, Brinkmann H, Chourrout D, Philippe H. Tunicates and not cephalochordates are the closest living relatives of vertebrates. Nature. 2006;439:965–8.
- 53. Janvier P. The Phylogeny of the Craniata, with Particular Reference to the Significance of Fossil "Agnathans." J Vertebr Paleontol. 1981;1:121–59.
- Janvier P. Facts and fancies about early fossil chordates and vertebrates. Nature. 2015;520:483–9.
- 55. Donoghue PC, Forey PL, Aldridge RJ. Conodont affinity and chordate phylogeny. Biol Rev Camb Philos Soc. 2000;75:191–251.
- Martini FH. The Ecology of Hagfishes. In: Jørgensen JM, Lomholt JP, Weber RE, Malte H, editors. The biology of hagfishes. Dordrecht: Springer, Netherlands; 1998. p. 57–77.
- Zintzen V, Roberts CD, Shepherd L, Stewart AL, Struthers CD, Anderson MJ, et al. Review and phylogeny of the New Zealand hagfishes (Myxiniformes: Myxinidae), with a description of three new species. Zool J Linn Soc. 2015;174:363–93.
- Martini F, Lesser M, Heiser JB. Ecology of the hagfish, Myxine glutinosa L., in the gulf of Maine: II. Potential impact on benthic communities and commercial fisheries. J Exper Marine Biol Ecol. 1997;214:97–106.
- Møller PR, Jones WJ. Eptatretus strickrotti n. sp. (Myxinidae): first hagfish captured from a hydrothermal vent. Biol Bull. 2007;212:55–66.
- Fernholm B, Kumar AB, Norān M. First record of hagfish (Cyclostomata: Myxinidae) in Indian waters. J Threatened Taxa. 2017;9:10365–8.
- 61. Priede IG, Froese R. Colonization of the deep sea by fishes. J Fish Biol. 2013;83:1528–50.
- 62. Ichikawa T, Kobayashi H, Nozaki M. Seasonal Migration of the Hagfish, Eptatretus burgeri. Girard jzoo. 2000;17:217–23.
- Zintzen V, Roberts CD, Anderson MJ, Stewart AL, Struthers CD, Harvey ES. Hagfish predatory behaviour and slime defence mechanism. Sci Rep. 2011;1:131.
- 64. Gess RW, Coates MI, Rubidge BS. A lamprey from the Devonian period of South Africa. Nature. 2006;443:981–4.
- 65. Miyashita T, Gess RW, Tietjen K, Coates Ml. Non-ammocoete larvae of Palaeozoic stem lampreys. Nature. 2021;591:408–12.
- Dearden RP, Lanzetti A, Giles S, Johanson Z, Jones AS, Lautenschlager S, et al. The oldest three-dimensionally preserved vertebrate neurocranium. Nature. 2023;621:782–7.
- Reeves JC, Wogelius RA, Keating JN, Sansom RS. Lasanius, an exceptionally preserved Silurian jawless fish from Scotland. Palaeontology. 2023;66:e12643.
- Delsuc F, Philippe H, Tsagkogeorga G, Simion P, Tilak M-K, Turon X, et al. A phylogenomic framework and timescale for comparative studies of tunicates. BMC Biol. 2018;16:39.

- Kuo C-H, Huang S, Lee S-C. Phylogeny of hagfish based on the mitochondrial 16S rRNA gene. Mol Phylogenet Evol. 2003;28:448–57.
- Chang M, Zhang J, Miao D. A lamprey from the Cretaceous Jehol biota of China. Nature. 2006;441:972–4.
- Chang M, Wu F, Miao D, Zhang J. Discovery of fossil lamprey larva from the Lower Cretaceous reveals its three-phased life cycle. Proc Natl Acad Sci. 2014;111:15486–90.
- 72. Huang W. A new species of fossil lamprey (Petromyzontida: Petromyzontiformes) from Hebei, China. Histor Biol. 2023;0:1–13.
- McCoy VE, Saupe EE, Lamsdell JC, Tarhan LG, McMahon S, Lidgard S, et al. The 'Tully monster' is a vertebrate. Nature. 2016;532:496–9.
- McCoy VE, Wittry J, Sadabadi H, Mayer P. A reappraisal of Nemavermes mackeei from the Mazon Creek fossil site expands Carboniferous cyclostome diversity. J Paleontol. 2023;97(5):1116–32. https://doi.org/ 10.1017/jpa.2023.72.
- Hirasawa T, Oisi Y, Kuratani S. Palaeospondylus as a primitive hagfish. Zool Lett. 2016;2:20.
- Joss J, Johanson Z. Is Palaeospondylus gunni a fossil larval lungfish? Insights from Neoceratodus forsteri development. J Exp Zool B Mol Dev Evol. 2007;308:163–71.
- 77. Brownstein CD. Palaeospondylus and the early evolution of gnathostomes. Nature. 2023;620:E20–2.
- Johanson Z, Smith M, Sanchez S, Senden T, Trinajstic K, Pfaff C. Questioning hagfish affinities of the enigmatic Devonian vertebrate Palaeospondylus. Royal Soc Open Sci. 2017;4:170214.
- Song YS, Kim J-K. Molecular phylogeny and classification of the family Myxinidae (Cyclostomata: Myxiniformes) using the supermatrix method. J Asia-Pac Biodivers. 2020;13:533–8.
- Mincarone M. Further description of the hagfish Nemamyxine kreffti McMillan & Wisner, 1982 (AGNATHA, MYXINIDAE). Mare Magnum. 2001;1(1):19–22.
- Minh BQ, Schmidt HA, Chernomor O, Schrempf D, Woodhams MD, von Haeseler A, et al. IQ-TREE 2: new models and efficient methods for phylogenetic inference in the genomic era. Mol Biol Evol. 2020;37:1530–4.
- Kalyaanamoorthy S, Minh BQ, Wong TKF, von Haeseler A, Jermiin LS. ModelFinder: fast model selection for accurate phylogenetic estimates. Nat Methods. 2017;14:587–9.
- Ronquist F, Teslenko M, Van Der Mark P, Ayres DL, Darling A, Höhna S, et al. MrBayes 3.2: Efficient bayesian phylogenetic inference and model choice across a large model space. Syst Biol. 2012;61:539–42.
- Bouckaert R, Heled J, Kühnert D, Vaughan T, Wu C-H, Xie D, et al. BEAST 2: a software platform for Bayesian evolutionary analysis. PLoS Comput Biol. 2014;10:e1003537.
- Bouckaert R, Vaughan TG, Barido-Sottani J, Duchêne S, Fourment M, Gavryushkina A, et al. BEAST 2.5: An advanced software platform for Bayesian evolutionary analysis. PLOS Comput Biol. 2019;15:e1006650.
- Gavryushkina A, Heath TA, Ksepka DT, Stadler T, Welch D, Drummond AJ. Bayesian total-evidence dating reveals the recent crown radiation of penguins. Syst Biol. 2017;66(1):57–73.
- Andreev PS, Sansom IJ, Li Q, Zhao W, Wang J, Wang C-C, et al. Spiny chondrichthyan from the lower Silurian of South China. Nature. 2022;609:969–74.
- Rambaut A, Drummond AJ, Xie D, Baele G, Suchard MA. Posterior Summarization in Bayesian Phylogenetics Using Tracer 1.7. Syst Biol. 2018;67:901–4.
- 89. Townsend JP. Profiling phylogenetic informativeness. Syst Biol. 2007;56:222–31.
- López-Giráldez F, Townsend JP. PhyDesign: an online application for profiling phylogenetic informativeness. BMC Evol Biol. 2011;11:152.
- Kumar S, Suleski M, Craig JM, Kasprowicz AE, Sanderford M, Li M, et al. TimeTree 5: an expanded resource for species divergence times. Mol Biol Evol. 2022;39:msac174.
- 92. Doran Brownstein C, Yang L, Friedman M, Near TJ. Phylogenomics of the ancient and species-Depauperate gars tracks 150 million years of continental fragmentation in the northern hemisphere. Syst Biol. 2023;72:213–27.
- 93. Maile AJ, May ZA, DeArmon ES, Martin RP, Davis MP. Marine habitat transitions and body-shape evolution in lizardfishes and their allies (Aulopiformes). Cope. 2020;108:820–32.
- 94. Angel MV. 1 What is the deep sea? In: Randall DJ, Farrell AP, editors. Fish physiology. Academic; 1997. p. 1–41.

- Qi L, Xu B, Kong L, Li Q. Improved phylogenetic resolution within Neritidae (Gastropoda, Nertimorpha) with implications for the evolution of shell traits and habitat. Zoolog Scr. 2023;52:46–57.
- Rousset V, Plaisance L, Erséus C, Siddall ME, Rouse GW. Evolution of habitat preference in Clitellata (Annelida). Biol J Linnean Soc. 2008;95:447–64.
- 97. Wilberg EW, Turner AH, Brochu CA. Evolutionary structure and timing of major habitat shifts in Crocodylomorpha. Sci Rep. 2019;9:514.
- Habitat use affects morphological diversification in dragon lizards -COLLAR - 2010 – J. Evol Biol. Wiley Online Library. https://onlinelibrary. wiley.com/doi/full/10.1111/j.1420-9101.2010.01971.x. Accessed 23 Dec 2023.
- 99. Hardy CR. Reconstructing ancestral ecologies: challenges and possible solutions. Divers Distrib. 2006;12:7–19.
- 100. Revell LJ. phytools: an R package for phylogenetic comparative biology (and other things). Methods Ecol Evol. 2012;3:217–23.
- FitzJohn RG, Maddison WP, Otto SP. Estimating trait-dependent speciation and extinction rates from incompletely resolved phylogenies. Syst Biol. 2009;58:595–611.
- 102. Sansom RS, Gabbott SE, Purnell MA. Non-random decay of chordate characters causes bias in fossil interpretation. Nature. 2010;463:797–800.
- 103. Sansom RS, Wills MA. Fossilization causes organisms to appear erroneously primitive by distorting evolutionary trees. Sci Rep. 2013;3:2545.
- Dornburg A, Townsend JP, Friedman M, Near TJ. Phylogenetic informativeness reconciles ray-finned fish molecular divergence times. BMC Evol Biol. 2014;14:169.
- Sansom RS, Randle E, Donoghue PCJ. Discriminating signal from noise in the fossil record of early vertebrates reveals cryptic evolutionary history. Proc Royal Soc B: Biol Sci. 2015;282:20142245.
- Ota KG, Fujimoto S, Oisi Y, Kuratani S. Late development of hagfish vertebral elements. J Exp Zool B Mol Dev Evol. 2013;320:129–39.
- Dong EM, Allison WT. Vertebrate features revealed in the rudimentary eye of the Pacific hagfish (Eptatretus stoutii). Proc Biol Sci. 2021;288:20202187.
- Locket NA, Jørgensen JM. The Eyes of Hagfishes. In: Jørgensen JM, Lomholt JP, Weber RE, Malte H, editors. The Biology of Hagfishes. Dordrecht: Springer, Netherlands; 1998. p. 541–56.
- 109. Clifford AM, Goss GG, Wilkie MP. Adaptations of a deep sea scavenger: high ammonia tolerance and active NH₄⁺ excretion by the Pacific hagfish (Eptatretus stoutii). Comp Biochem Physiol A Mol Integr Physiol. 2015;182:64–74.
- Pyron RA, Burbrink FT, Wiens JJ. A phylogeny and revised classification of Squamata, including 4161 species of lizards and snakes. BMC Evol Biol. 2013;13:93.
- 111. Burbrink FT, Grazziotin FG, Pyron RA, Cundall D, Donnellan S, Irish F, et al. Interrogating genomic-scale data for squamata (Lizards, Snakes, and Amphisbaenians) shows no support for key traditional morphological relationships. Syst Biol. 2020;69:502–20.
- 112. Friedman M. Explosive morphological diversification of spiny-finned teleost fishes in the aftermath of the end-Cretaceous extinction. Proc Royal Soc B: Biol Sci. 2010;277:1675–83.
- 113. Pyron RA. Divergence time estimation using fossils as terminal taxa and the origins of lissamphibia. Syst Biol. 2011;60:466–81.
- 114. Pimiento C, Cantalapiedra JL, Shimada K, Field DJ, Smaers JB. Evolutionary pathways toward gigantism in sharks and rays. Evolution. 2019;73:588–99.
- 115. Stein RW, Mull CG, Kuhn TS, Aschliman NC, Davidson LNK, Joy JB, et al. Global priorities for conserving the evolutionary history of sharks, rays and chimaeras. Nat Ecol Evol. 2018;2:288–98.
- Gemmell NJ, Rutherford K, Prost S, Tollis M, Winter D, Macey JR, et al. The tuatara genome reveals ancient features of amniote evolution. Nature. 2020;584:403–9.
- 117. Eschmeyer's Catalog of Fishes | California Academy of Sciences. https:// www.calacademy.org/scientists/projects/eschmeyers-catalog-of-fishes. Accessed 14 Feb 2024.
- Inoue JG, Miya M, Lam K, Tay B-H, Danks JA, Bell J, et al. Evolutionary origin and phylogeny of the modern holocephalans (Chondrichthyes: Chimaeriformes): a mitogenomic perspective. Mol Biol Evol. 2010;27:2576–86.
- 119. Wen-Jin Z, Min Z. Diversification and faunal shift of Siluro-Devonian vertebrates of China. Geol J. 2007;42:351–69.

- 120. Zhu Y, Li Q, Lu J, Chen Y, Wang J, Gai Z, et al. The oldest complete jawed vertebrates from the early Silurian of China. Nature. 2022;609:954–8.
- 121. Friedman M, Sallan LC. Five hundred million years of extinction and recovery: a phanerozoic survey of large-scale diversity patterns in fishes. Palaeontology. 2012;55:707–42.
- Bazzi M, Kear BP, Blom H, Ahlberg PE, Campione NE. Static dental disparity and morphological turnover in sharks across the end-cretaceous mass extinction. Curr Biol. 2018;28:2607-2615.e3.
- Bazzi M, Campione NE, Ahlberg PE, Blom H, Kear BP. Tooth morphology elucidates shark evolution across the end-Cretaceous mass extinction. PLoS Biol. 2021;19:e3001108.
- 124. Thomas E, Thomas E. Late Cretaceous-early Eocene mass extinctions in the deep sea. Geol Soc Am Spec Pap. 1990;247:481–95.
- 125. García-Girón J, Chiarenza AA, Alahuhta J, DeMar DG, Heino J, Mannion PD, et al. Shifts in food webs and niche stability shaped survivorship and extinction at the end-Cretaceous. Sci Adv. 2022;8:eadd5040.
- 126. Wilson GP, DeMar Jr DG, Carter G. Extinction and survival of salamander and salamander-like amphibians across the Cretaceous-Paleogene boundary in northeastern Montana, USA. Through the end of the Cretaceous in the type locality of the Hell Creek Formation in Montana and adjacent areas. Geological Soc Am Spec Pap. 2014;503:271–97.
- 127. Brownstein CD, Lyson TR. Giant gar from directly above the Cretaceous-Palaeogene boundary suggests healthy freshwater ecosystems existed within thousands of years of the asteroid impact. Biol Let. 2022;18:20220118.
- 128. Hilton EJ, Grande L. Late Cretaceous sturgeons (Acipenseridae) from North America, with two new species from the Tanis site in the Hell Creek Formation of North Dakota. J Paleontol. 2023;97:189–217.

Publisher's Note

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.