

Research article

Open Access

## Modelling the evolution of the archaeal tryptophan synthase

Rainer Merkl\*

Address: Institut für Biophysik und Physikalische Biochemie, Universität Regensburg, 93040 Regensburg, Germany

Email: Rainer Merkl\* - Rainer.Merkl@Biologie.uni-regensburg.de

\* Corresponding author

Published: 10 April 2007

Received: 13 February 2007

BMC Evolutionary Biology 2007, 7:59 doi:10.1186/1471-2148-7-59

Accepted: 10 April 2007

This article is available from: <http://www.biomedcentral.com/1471-2148/7/59>

© 2007 Merkl; licensee BioMed Central Ltd.

This is an Open Access article distributed under the terms of the Creative Commons Attribution License (<http://creativecommons.org/licenses/by/2.0>), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

### Abstract

**Background:** Microorganisms and plants are able to produce tryptophan. Enzymes catalysing the last seven steps of tryptophan biosynthesis are encoded in the canonical *trp* operon. Among the *trp* genes are most frequently *trpA* and *trpB*, which code for the alpha and beta subunit of tryptophan synthase. In several prokaryotic genomes, two variants of *trpB* (named *trpB1* or *trpB2*) occur in different combinations. The evolutionary history of these *trpB* genes is under debate.

**Results:** In order to study the evolution of *trp* genes, completely sequenced archaeal and bacterial genomes containing *trpB* were analysed. Phylogenetic trees indicated that TrpB sequences constitute four distinct groups; their composition is in agreement with the location of respective genes. The first group consisted exclusively of *trpB1* genes most of which belonged to *trp* operons. Groups two to four contained *trpB2* genes. The largest group (*trpB2\_o*) contained *trpB2* genes all located outside of operons. Most of these genes originated from species possessing an operon-based *trpB1* in addition. Groups three and four pertain to *trpB2* genes of those genomes containing exclusively one or two *trpB2* genes, but no *trpB1*. One group (*trpB2\_i*) consisted of *trpB2* genes located inside, the other (*trpB2\_a*) of *trpB2* genes located outside the *trp* operon. TrpA and TrpB form a heterodimer and cooperate biochemically. In order to characterise *trpB* variants and stages of TrpA/TrpB cooperation *in silico*, several approaches were combined. Phylogenetic trees were constructed for all *trp* genes; their structure was assessed *via* bootstrapping. Alternative models of *trpB* evolution were evaluated with parsimony arguments. The four groups of *trpB* variants were correlated with archaeal speciation. Several stages of TrpA/TrpB cooperation were identified and *trpB* variants were characterised. Most plausibly, *trpB2* represents the predecessor of the modern *trpB* gene, and *trpB1* evolved in an ancestral bacterium.

**Conclusion:** In archaeal genomes, several stages of *trpB* evolution, TrpA/TrpB cooperation, and operon formation can be observed. Thus, archaeal *trp* genes may serve as a model system for studying the evolution of protein-protein interactions and operon formation.

### Background

The synthesis of tryptophan is a common metabolic capability of microorganisms and higher plants, which is not provided by mammals. The prokaryotic *trp* operon encodes the enzymes catalysing the final and pathway-

specific steps from chorismate to L-tryptophan. For more than 40 years, the enterobacterial operon has now been the classical model system for studying the evolutionary relation of genes and enzymes (see [1,2] and references therein) as well as gene regulation. Considering gene reg-

ulation, several, conceptually quite different mechanisms have been described for the *trp* operon. Most of them were elucidated in bacterial species (see *e.g.* [3-5], and references therein). However, regulation of *trp* operon expression has also been shown for the archaea *Methanothermobacter thermoautotrophicus* [6,7] and *Thermococcus kodakaraensis* [8]. The reason for an elaborated regulation may be the fact that tryptophan is one of the amino acids, whose biochemical synthesis is very expensive [9]. Besides regulation, other features of tryptophan biosynthesis have been studied extensively. The composition of the operon and several aspects of its evolution have been analysed [10], and for each enzyme, at least one 3D-structure has been determined. Taken together, the *trp* operon is besides the ribosomal protein operons one of the best-characterised gene clusters occurring in microorganisms. Its investigation has provided fundamental insights into many aspects of bacterial genetics and enzymology; see [2].

The canonical *trp* operon encodes seven enzymes responsible for the synthesis of L-tryptophan from chorismate. The first reaction is catalysed by the anthranilate synthase, a glutamine amidotransferase, which is a complex consisting of the larger synthase (TrpE) and a smaller glutaminase (TrpG) subunit. The anthranilate phosphoribosyl transferase (TrpD) provides the glutamine amidotransferase function that allows glutamine to serve as the amino donor in anthranilate formation. The two subsequent enzymes, TrpF and TrpC, catalyse the isomerisation of phosphoribosylanthranilate and the synthesis of indole-3-glycerol phosphate, respectively.

TrpA and TrpB constitute the  $\alpha\beta\beta\alpha$  tryptophan synthase complex which catalyses the final reaction from indole-3-glycerol phosphate + L-serine to L-tryptophan + H<sub>2</sub>O. The  $\alpha$  subunit (TrpA) cleaves indoleglycerol-3-phosphate to glyceraldehyde-3-phosphate and indole. The latter is transported through a hydrophobic tunnel to the associated  $\beta$  subunit (TrpB), where it is condensed with L-serine to yield L-tryptophan [11]. A sophisticated mechanism of allostery links the  $\alpha$  and  $\beta$  monomers of the synthase; see *e.g.* [12].

Several Trp enzymes represent paradigmatically larger classes of proteins having similar function or protein architecture: TrpG is similar to HisH (an enzyme involved in histidine biosynthesis) and other glutaminases of type I glutamine amidotransferases [13]. TrpF, TrpC and TrpA are all  $(\beta\alpha)_8$  barrels possessing similar phosphate binding sites [14]. The basic  $(\beta\alpha)_8$  barrel is the most common enzyme fold in the PDB database of known protein structures [15].

For the bacterial *trp* genes, the following order was determined: large anthranilate synthase subunit (*trpE*), small anthranilate synthase subunit (*trpG*), anthranilate phosphoribosyl transferase (*trpD*), indole-3-glycerol phosphate synthase (*trpC*), phosphoribosyl anthranilate isomerase (*trpF*), tryptophan synthase  $\beta$  subunit (*trpB*) and tryptophan synthase  $\alpha$  subunit (*trpA*), or abbreviated *trpEGDCFBA* [16]. The gene-fusions *trpGD* and *trpEG* have been observed in several species; moreover, in other genomes, the operon is broken up into several gene clusters. In archaeal genomes, order of *trp* genes is highly variable. In *Sulfolobus solfataricus*, an intact operon *trpBADFEGC* is observed. In *Haloferax volcanii*, the *trp* operon is divided into two isolated clusters, *trpCBA* and *trpDFEG*, separated by more than 1200 kb. In the genome of *Natronomonas pharaonis*, there exist three homologs of *trpD* and two homologs of *trpB*, *trpE* and *trpG* each. *Pyrococcus horikoshii* completely lacks the genes for tryptophan synthesis (and for other aromatic amino acids).

The genes *trpB*, *trpA* and *trpE*, *trpG* are frequently in the same order and in close proximity, *i.e.* they comprise the linkage groups *trpBA* and *trpEG*. In both cases, the gene products constitute a holoenzyme complex, whose active centres interact with each other. Because they occur in both bacterial and archaeal genomes, these linkage groups have been identified as ancestral [16]. A reconstruction of the tentative ancestral *trp* operon is hampered by the observation that *trp* genes are poor phylogenetic reporters. Different rates of evolution, multiple gene duplications and convergent evolution, as a consequence of specific adaptation to environmental demands, may be the reason for inconsistencies seen in comparisons of phylogenies deduced from *trp* genes or rRNA [16]. Therefore, the evolution of each element of the *trp* operon has to be examined separately.

For evolutionary studies, tryptophan synthase is an especially interesting candidate. This enzyme has been analysed for decades in order to understand the structural basis and functional consequences of protein-protein interactions [17]. The isolated TrpA and TrpB proteins form stable, however poorly active  $\alpha$  monomers and  $\beta\beta$  homodimers, respectively [18,19]. Their assembly to the native  $\alpha\beta\beta\alpha$  complex induces conformational changes in both subunit types, as shown by X-ray crystallography for the *Pyrococcus furiosus* synthase [18]. The result of this communication between the  $\alpha$  and  $\beta$  subunits is a reciprocal activation by one to two orders of magnitude [20]. Conformational changes crucial for the allosteric communication between the active sites of the  $\alpha$ - and  $\beta$ -subunits have been analysed in detail for the *Salmonella typhimurium* tryptophan synthase; see *e.g.* [21-24].

The role of the  $\beta$ -subunit is of particular importance for the evolution of Trp synthase. For archaea and bacteria, it is known that two variants of *trpB* genes occur, which can clearly be distinguished by their protein sequences [25]. The major group, harbouring proteins of type TrpB1 includes the enzymes of enterobacteria and *Bacillus subtilis*. The minor group (denoted TrpB2) contains many archaeal proteins. Most prokaryotes like *E. coli* possess a single *trpB1* gene. However, in several bacterial and archaeal genomes, a combination of one *trpB1* and one *trpB2* gene occurs. In addition, some species exist, which have only one or two *trpB2*, but no *trpB1* gene. This variety prompted us to characterise the evolution of TrpB and its interaction with TrpA in detail, both biochemically and *in silico*.

Based on biochemical findings, a model for the evolution of the tryptophan synthase complex has recently been introduced [26]. This model assumes the existence of an ancient and non operon-based *trpB2*. After duplication, only one *trpB2* gene presumably has been integrated into the *trp* operon. Differences in evolutionary pressure may have been responsible for the divergence of non operon- and operon-based *trpB* genes. The coevolution with *trpA* may have led to a better adapted *trpB1*. The data on complex formation and subunit activation led us consider existing *trpB* variants as representatives of evolutionary steps in the postulated model.

In this study, I have assessed this model by phylogenetic methods. Two basic questions have been addressed: *i*) What is the evolutionary relationship of *trpB1* and *trpB2*? *ii*) How did extant archaeal *trp* operons evolve? Extending previous work [25], I will discuss novel hypotheses concerning the properties of TrpB2 and operon formation. Based on the content of 26 completely sequenced archaeal genomes, comparative analyses of *trp* sequences, and their locations in genomes will be reported in order to reconstruct the evolution of TrpB-type subunits and of the coevolution of TrpA/TrpB. It will be shown that TrpB2 variants represent different stages of TrpA/TrpB cooperation and that TrpB2 is favoured over TrpB1 in certain environments. Moreover, TrpB2 has features of a more ancient TrpB variant.

## Results and Discussion

### Assessing the composition of *trp* gene clusters

In order to describe the composition of *trp* regulons in a quantitative manner and to compare their content in archaeal and bacterial genomes, AMIGOS [27] was used. By comparing genomes, this program identifies gene clusters and rates each individual cluster element with a *cons<sub>CL</sub>*-score. The *cons<sub>CL</sub>*-score of an individual gene depends on *i*) the occurrence of this gene in a given gene cluster and *ii*) the global similarity of the genomes harbouring

these clusters. Thus, individual scores assess both the relatedness of genomes and the frequency with which individual genes are members of a cluster. The higher a score, the more pronounced is the occurrence of an individual gene in a given gene cluster. Table 1 lists *cons<sub>CL</sub>*-scores for elements of archaeal and bacterial *trp* operons. The numbers indicate that in bacteria the clustering of *trpA* and *trpB1* was stronger than that of all other *trp* genes. In archaeal genomes, the clustering of *trpE* and *trpG* was most prominent. A reason for the lower score of *trpB* in archaeal *trp* operons was the occurrence of two *trpB* variants (*trpB1* and *trpB2*) in these species. The scores signalled that *trpB1* was more frequently part of an *trp* operon than *trpB2*. Moreover, the score for *trpA* was lower than that of *trpE* or *trpG*. It follows for archaea that *trpA* and *trpB* are less strictly integrated into *trp* operons than in bacteria. This suggests that either evolutionary pressure responsible for operon formation is less pronounced or that additional selective forces disfavour the integration of *trpA* and *trpB* into certain archaeal *trp* operons.

It has been hypothesised that TrpB2 possesses a second function and acts as a serine deaminase [25]. This prediction has been deduced from the analysis of phyletic patterns, *i.e.* the absence of an encoded serine deaminase function in certain genomes. However, it has been shown that TrpB1 of *Thermotoga maritima* and TrpB2<sub>o</sub> proteins of *Sulfolobus solfataricus* and *T. maritima* have poor serine deaminase activities [26]. An alternative method of non-homologous gene annotation is the exploitation of gene neighbourhoods [28], as *e.g.* implemented with AMIGOS. For *trpB2*, AMIGOS did not detect a second conserved gene neighbourhood besides the one constituting *trp* operons. Thus, no clues for an additional function besides tryptophan synthesis have been deduced for *trpB2* by this approach.

### A naming code for *trpB* genes

The two variants of *trpB* occur in various genomes in different combinations [25]. In order to facilitate the analysis of phylogenetic trees, a naming scheme was introduced. Names of genes and gene products were generated according to the scheme *SPECIES\_LOC|TYPE|TAX*. Here, *SPECIES* is an abbreviation of the species name (see Materials). *LOC* indicates the position of the specific *trpB* gene relative to a putative *trp* operon (more precisely: relative to a *trpA* gene). If two *trpB* genes occur in a genome, they were labelled *\_i* (if the gene was located inside the *trp* operon) or *\_o* (if located outside the operon). If only a single *trpB* gene occurred in the genome, it was labelled *\_s*, if the gene was linked to *trpA*, and it was labelled *\_S*, if it was separate from *trpA*. *TYPE* indicates the gene type. It is 1 for *trpB1* and 2 for *trpB2*. Finally, *TAX* gives the taxonomical classification. It is C for *Crenarchaeota*, E for *Euryarchaeota* and B for *Bacteria*. The following examples

**Table 1:  $cons_{cl}$  scores for *trp* genes**

$cons_{cl}$ - values		Protein	COG #	Function
Archaea	Bacteria			
2.0	2.8	TrpE	COG0147	anthranilate/para-aminobenzoate synthases comp. I
2.1	2.7	TrpG	COG0512	anthranilate/para-aminobenzoate synthases comp. II
1.8	2.3	TrpF	COG0135	phosphoribosylanthranilate isomerase
1.8	2.9	TrpC	COG0134	indole-3-glycerol phosphate synthase
1.9	3.0	TrpA	COG0159	tryptophan synthase alpha chain
1.4	3.0	TrpB1	COG0133	tryptophan synthase beta chain
0.6	-	TrpB2	COG1350	paralogue of TrpB
1.9	2.6	TrpD	COG0547	anthranilate phosphoribosyltransferase

Note: The first two columns list score values deduced from representative sets of archaeal and bacterial genomes. Columns three and four list protein names, COG numbers, and protein function. The  $cons_{cl}$ -scores were determined by using AMIGOS [27]. COG numbers indicate orthologous gene clusters as defined in the COG database [29].

explain how to resolve sequence names: Aperi\_o2C was used to name a *trpB* gene in the genome of *Aeropyrum pernix* (Aperi), which occurred outside the *trp* operon (\_o) and was of type *trpB2* (2). As *A. pernix* is a *Crenarchaeota*, the name ends with a C. The \_o notation indicates that a second *trpB* gene exists in *A. pernix*. This gene was consequently named Aperi\_i2C, as it is a *trpB2* gene inside the *trp* operon. Note that also pairs like Tmarit\_i1B and Tmarit\_o2B exist indicating the occurrence of a *trpB1* gene inside and a *trpB2* gene outside the *trp* operon. Sacido\_s2C is the designation of a *trpB2* gene located inside the *trp* operon. As *Sulfolobus acidocaldarius* possesses only one *trpB* gene, it was labelled with a \_s. Since *Thermoplasma volcanium* possesses only one *trpB* gene, which is non operon-based and of type *trpB2*, this gene was named Tvolc\_S2E. Designations of the encoded proteins were assigned in a corresponding way.

#### Determining the occurrence of *trpB* genes

In order to determine the distribution of *trpB* variants, the COG [29] and the STRING database [30] were used. For all completely sequenced archaeal and bacterial genomes, their occurrence was determined and their location was identified. Depending on the occurrence of *trpB* variants, archaeal species were grouped into five categories, named species-types in the following. Note that these species-types characterise the content of genomes. Links to the above naming scheme for genes are gene location and type.

As Table 2 shows, there were six archaeal genomes possessing a single *trpB* gene of class *trpB1* (*s1* or *S1* species), four genomes with a single *trpB* gene of class *trpB2* (*s2* or *S2* species), five genomes harbouring one operon-based and one additional, non operon-based *trpB2* each (*i2\_o2* species), ten species of type *i1\_o2* (one operon-based *trpB1* and one additional *trpB2* gene) and one species possessing one operon-based and at least one non operon-

based *trpB1* gene (*i1\_o1* species). The most frequent combination (10 out of 26) was an operon-based *trpB1* and a non operon-based *trpB2* gene (*i1\_o2* species). *N. pharaonis* was the only archaeal species of type *i1\_o1*. All five completely sequenced *Crenarchaeota* possess exclusively genes of class *trpB2*.

Bacterial species did not contribute species-types noticeably different from those observed among archaea (data not shown). Both *Geobacter* species represent special cases most plausibly explained by ongoing genomic rearrangements: Gsulfu\_i2B is an operon-based *trpB2* gene of type *TrpB2\_o*. The *trp* operon of *G. sulfurreducens* harbours both a *trpB1* and a *trpB2* gene. According to the annotation, the *trpB1* gene (Locus tag GSU2375) contains a frameshift and is annotated as a pseudogene [31]. A direct neighbour of *trpB1* in *G. metallireducens* is a transposase, making a recent transfer of this gene plausible. In comparison to archaea, the occurrence of *trpB2* was less frequent in bacterial genomes and none contained exclusively *trpB2* genes.

#### Assessing phylogenetic relationship of *trp* genes

Sequences originating from all archaea and several representative bacteria were selected for a phylogenetic classification of *trp* genes. Multiple sequence alignments were created by using M-Coffee [32], and trees were constructed and evaluated using SplitsTrees [33]. Figures 1, 2, 3, 4 are plots of unrooted trees generated for protein sequences of TrpA, TrpB, TrpD, TrpE, and TrpG. In order to assess the statistical strength of individual edges, bootstrap resampling was used. For relevant edges, bootstrap values were plotted; see Figures 1, 2, 3, 4. The trees were analysed in detail, as follows.

#### *TrpB*

In agreement with previous findings [25], TrpB1 and TrpB2 clearly fall into two distinct groups. This distinction

**Table 2: Classifying known archaeal genomes according to the occurrence of *trpB* genes**

S2 (3), s2 (1)	i2_o2 (5)	i1_o2 (10)	i1_o1 (1)	S1 (1), s1 (5)
<i>S. acidocaldarius</i> , s2C 3, TA <i>T. volcanium</i> , S2E 3, TA <i>T. acidophilum</i> , S2E 3, TA <i>P. horikoshii</i> , S2E 2, HT	<i>A. pernix</i> , i2C 4, o2C 2, HT <i>P. aerophilum</i> , i2C 3, o2C 4, HT <i>P. torridus</i> , i2E 2, o2E 2, TA <i>S. solfataricus</i> , i2C 3, o2C 3, TA <i>S. tokodaii</i> , i2C 3, o2C 4, TA	<i>A. fulgidus</i> , i1E 2, o2E 6, HT <i>M. acetivorans</i> , i1E 1, o2E 5, MS <i>M. barkeri</i> , i1E 1, o2E 4, MS <i>M. burtonii</i> , i1E 1, o2E 4, MS <i>M. hungatei</i> , i1E 1, o2E 4, MS <i>M. mazei</i> , i1E 1, o2E 5, MS <i>M. thermoautotrophicus</i> , i1E 1, o2E 3, TP <i>P. abyssi</i> , i1E 3, o2E 2, HT <i>P. furiosus</i> , i1E 3, o2E 2, HT <i>T. kodakaraensis</i> , i1E 2, o2E 2, HT	<i>N. pharaonis</i> , i1E 1, o1E 0, HP	<i>M. kandleri</i> , S1E 4, YP <i>Halobacterium</i> , s1E 1, HP <i>H. marismortui</i> , s1E 1, HP <i>M. maripaludis</i> , s1E 1, MS <i>M. stadtmanae</i> , s1E 1, MS <i>M. jannaschii</i> , s1E 2, YP
2.75	i2: 3.0, o2: 3.0	i1: 1.6, o2: 3.7	i1: 1.0, o1: 0.0	1.6

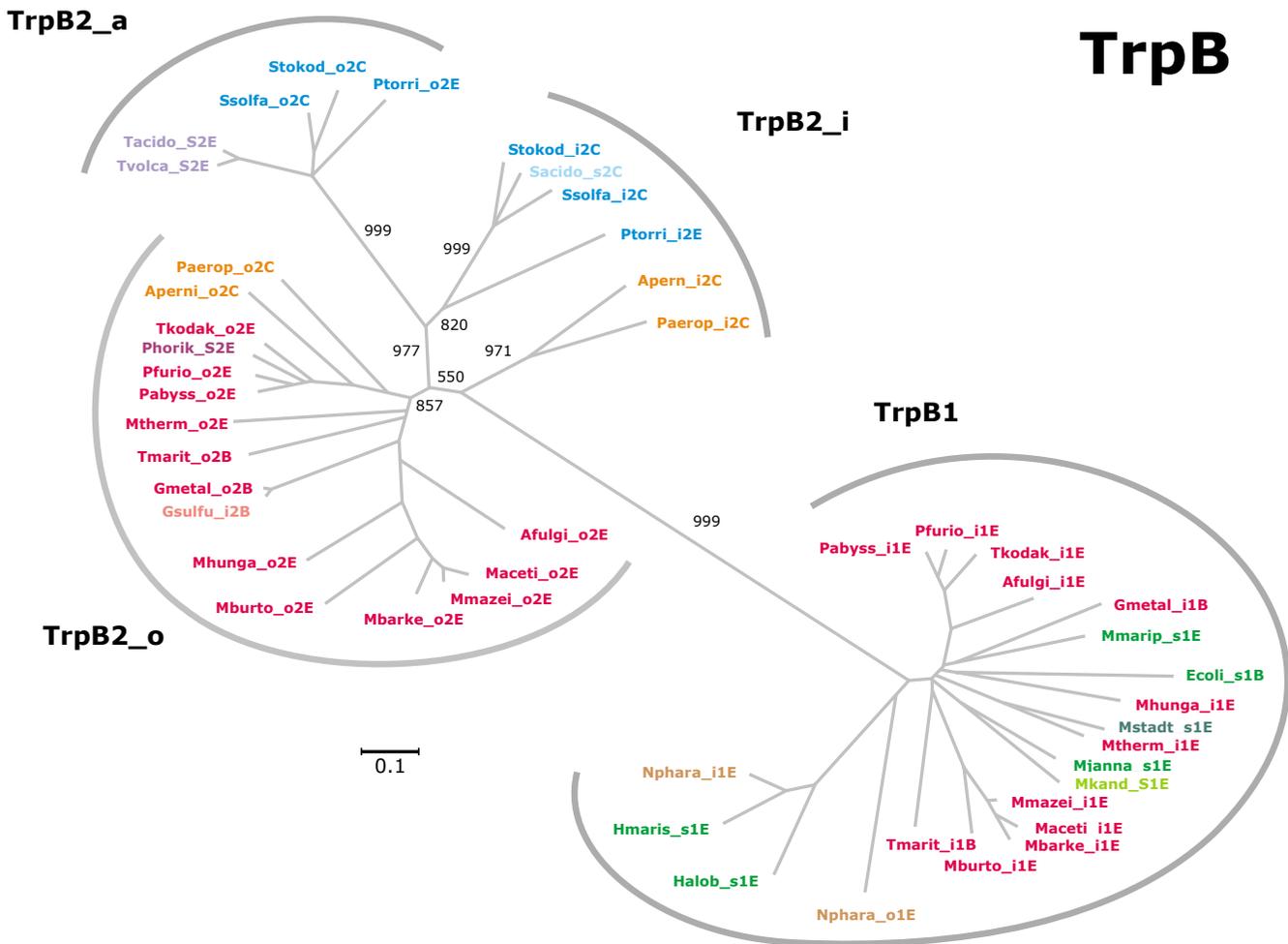
Note: The occurrence and the location of *trpB* genes were coded according to the following scheme used in the top line: S2 species possess exactly one, non operon-based *trpB2* gene, s2: ditto, the gene is located inside the *trp* operon. *trpB1* was treated analogously. i2\_o2 are species possessing an *trpB2* gene inside and a second *trpB2* outside the operon, i1\_o2 are species with an operon-based *trpB1* and a non operon-based *trpB2*, and i1\_o1 are species possessing an operon-based and at least one non operon-based *trpB1*. The number of genomes having the same species-type is given in brackets. The acronyms following the species name classify *trpB* genes; see Results. Numbers give tryptophan codons occurring in the respective gene. The abbreviations indicate hyperthermophilic (HT), thermoacidophilic (TA), thermophilic (TP), mesophilic (MS), halophilic (HP), or hyperthermophilic + halophilic species (YP). The last line of the table gives mean values for the occurrence of tryptophan codons.

was supported by a high bootstrap value; see Figure 1. Moreover, among TrpB2 sequences a finer sub-clustering could be deduced, which was in agreement with the location of the genes. One group (labelled TrpB2\_o) consisted of products of *trpB2* genes not located in operons. 14 out of 16 elements were TrpB2 sequences originating from i1\_o2 species, i.e. species possessing besides an isolated lying *trpB2* an additional, operon-based *trpB1*. The genes Paerip\_o2C and Aperni\_o2C of the two i2\_o2 species *Pyrobaculum aerophilum* and *A. pernix* belonged to this group too. These two species possess a *trp* operon containing a *trpB2* gene. Bacterial TrpB2\_o sequences, which originated from the i1\_o2 species *T. maritima* and *G. metallireducens* did not form an isolated subtree. This finding argues for a common origin of bacterial and archaeal *trpB2\_o* genes.

The other two subgroups of TrpB2 variants were clearly distinct from the TrpB2\_o cluster. The sequences of these clusters originated from archaeal S2 (*Thermoplasmataceae*), s2 or i2\_o2 species (*Sulfolobaceae*, *Picrophilus torridus*, *A. pernix*, *P. aerophilum*), i.e. species possessing exclusively one or two *trpB2* genes. These sequences formed two clearly separated sets. The first set, named TrpB2\_i, subsumes operon-based *trpB2* genes, and harboured Stokod\_i2C, Sacido\_s2C, Ssolfa\_i2C, Ptorri\_i2E, Apern\_i2C, and Paerop\_i2C. The second set, named TrpB2\_a, consisted of Ptorri\_o2E, Stokod\_o2C, Ssolfo\_o2C, Tacido\_S2E, and Tvolca\_S2E, and subsumed *trpB2* genes located outside *trp* operons. For *Thermoplasma volcanium* and *Thermoplasma acidophilum*, these *trpB2*

genes were the only *trpB* genes, for *S. solfataricus*, *S. tokodaii* and *P. torridus*, a second, however distinguishable *trpB2* gene of type *trpB2\_i* was part of the *trp* operon. Proteins of type TrpB2\_i formed two finer subgroups: Those of *P. torridus* and the *Sulfolobaceae* resembled more sequences of TrpB2\_a. Those of *A. pernix* and *P. aerophilum*, which possess a non operon-based *trpB2\_o* gene, were different both from TrpB2\_a and from TrpB2\_o sequences; see Figure 1. All relevant edges separating these groups are due to their high bootstrap value statistically highly significant.

As a single exception, the genome of *P. horikoshii* did not follow the general classification scheme. It possesses a single *trpB2* gene, which is of type *trpB2\_o* and not – as expected – of type *trpB2\_a*. However, this genome lacks all the other *trp* genes, which has been previously interpreted as reductive evolution [10]. The occurrence of a *trpB2\_o* gene might be due to the loss of the complete *trp* operon after speciation of *trpB2\_i* and *trpB2\_o*. The fact that the *P. horikoshii* *trpB2\_o* gene was not affected by the reduction has been considered as an argument for assigning to it an other selective function [25], which has not been identified yet. As noted above, the two bacterial *Geobacter* species represent special cases associated with the presumptive rearrangement of *trp* genes. Briefly, the *trpB* variants can be characterised as follows: *trpB1* genes occur exclusively in *trp* operons. *trpB2\_o* variants represent genes occurring outside operons in those species that have an operon-based *trpB1*. Several archaeal species possess



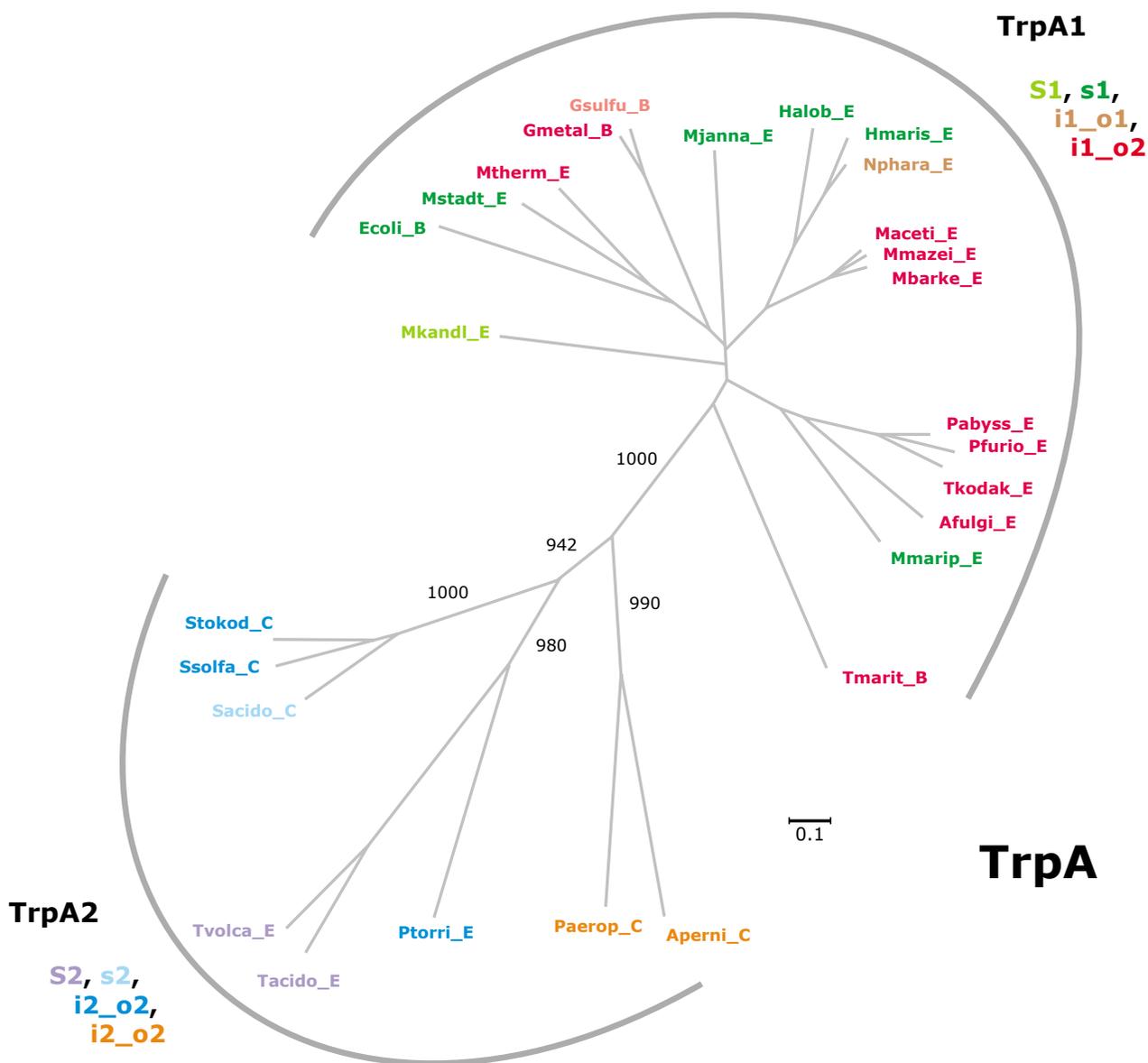
**Figure 1**  
**Phylogenetic tree of TrpB sequences.** Using archaeal and bacterial TrpB sequences, a multiple sequence alignment was generated and an unrooted phylogenetic tree was constructed. Proteins were labelled according to the naming scheme introduced in the Results section. Subtrees were marked according to the sequence type (TrpB1 or TrpB2). TrpB2 sequences span three subtrees; clustering is in agreement with the location of genes. TrpB2<sub>o</sub> proteins are all encoded outside operons; 14 out of 16 originate from species that possess an operon-based *trpB1* in addition. TrpB2<sub>i</sub> proteins are encoded inside operons. Each of these genes is accompanied by a non operon-based *trpB2*. TrpB2<sub>a</sub> sequences occur exclusively in genomes that have a single *trpB2* gene or occur as a second *trpB2* outside an operon in combination with a *trpB2<sub>i</sub>* gene. The numbers are bootstrap values resulting from 1000 replications. Gene names are colour-coded. Blue colours indicate genes occurring in S2 (violet), s2 (light blue) and those *i2<sub>o2</sub>* species, which possess *trpB2<sub>a</sub>* or *trpB2<sub>i</sub>* genes (dark blue). Orange colours designate *trpB2<sub>i</sub>* and *trpB2<sub>o</sub>* genes. Red colours signify genes of *il<sub>o2</sub>*, S2, or s2 species, and green colours mark genes of *sI* (light green) or *S1* (dark green) species. The names of the two *trpB1* copies occurring in *N. pharaonis* are printed in brown. For acronyms of species-types, see legend of Table 2. The length of the horizontal bar corresponds to 0.1 substitutions per site.

exclusively *trpB2* genes: If only one *trpB2* gene exists, it is of type *trpB2<sub>a</sub>*, if two *trpB2* genes occur, one is an operon-based *trpB2<sub>i</sub>*, the second a *trpB2<sub>a</sub>*, or a *trpB2<sub>o</sub>* gene.

**TrpA**

Correlated with TrpB speciation, TrpA proteins showed a division into two, statistically highly significant subgroups; see Figure 2. The larger TrpA1 group consisted of TrpA sequences originating from genomes that possess a

*trpB1* gene. Most likely, TrpA1 proteins interact with the operon encoded TrpB1 and thus fall into the same class. The smaller TrpA2 group contained exclusively TrpA proteins of species-types S2, s2, or *i2<sub>o2</sub>*, *i.e.* TrpA proteins whose putative interaction partner is exclusively a TrpB2 protein. The high bootstrap value of 1000 (100%) for the central edge emphasises the distinction made between TrpA1 and TrpA2. S2, s2, *i2<sub>o2</sub>* species formed three statistically significant subtrees; compare Figure 2. These har-

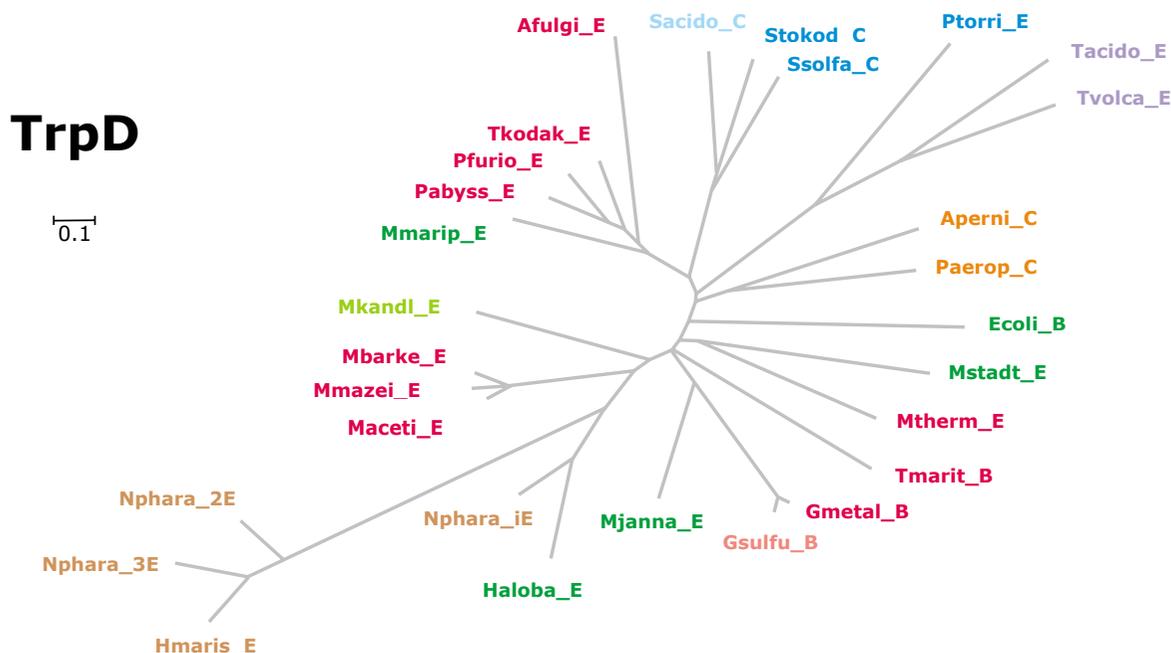


**Figure 2**  
**Phylogenetic tree of archaeal and bacterial TrpA sequences.** The two subtrees cluster genomes, which encode at least one *trpB1* gene (*S1*, *s1*, *il\_o1*, or *il\_o2* species) or which possess only genes of type *trpB2* (*S2*, *s2*, *i2\_o2* species). The clusters were named TrpA1 or TrpA2, respectively. For abbreviations of sequence names, see Results. For colour code, see legend of Figure 1. For the acronyms of species-types, see legend of Table 2.

boured the TrpA sequences of (i) *Sulfolobaceae*, (ii) *Thermoplasmatales* (*T. acidophilum*, *T. volcanium*, *P. torridus*) and (iii) *P. aerophilum*, and *A. pernix*. The composition of these groups is in agreement with the TrpB2\_a and TrpB2\_i groups in Figure 1 and indicates the coevolution of *trpB2* variants with *trpA*.

*TrpD, TrpE, and TrpG*

In all three trees (see Figures 3 and 4), both the proteins of *Thermoplasmatales* and of the three *Sulfolobaceae* constituted sub-clusters. The edges determined for TrpD or TrpE entries of these species have similar lengths as those calculated for TrpA or TrpB. Especially for the *trpA* and *trpB* genes of these species, an increased rate of evolution has been previously postulated [25]. However, the comparison of trees and edge lengths showed that in these species



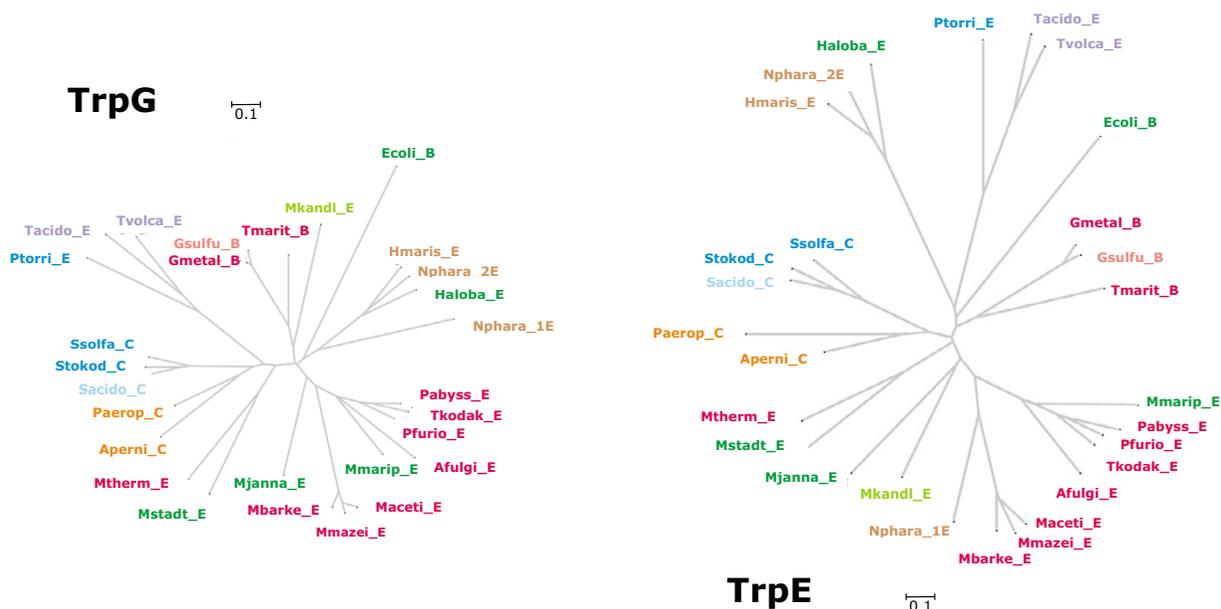
**Figure 3**  
**Phylogenetic tree of TrpD sequences.** Archaeal and bacterial protein sequences were used to construct the unrooted tree. The last letter of the acronyms indicates the taxonomical position of the species. "E" marks *Euryarchaeota*, "C" *Crenarchaeota*, and "B" bacterial species. The three TrpD sequences of *N. pharaonis* are designated as \_1E, \_2E, and \_3E. For colour code and abbreviations, see legend of Figure 1.

evolutionary divergence is similarly high for several proteins encoded by the *trp* operon. These findings argue against a specifically increased rate of *trpA* and *trpB* evolution. In general, smaller genomes evolve faster [34]. Therefore, a higher evolutionary rate in the *trp* genes of *Thermoplasmatales* is more plausibly explained by a general trend, which is due to their smaller genome size.

Interestingly, no sub-clustering into smaller, distinctly separated groups was observed in TrpE and TrpG, which form like TrpA and TrpB a heteromeric complex. The above finding distinguishes the subunits of tryptophan synthase from those of anthranilate synthase. TrpG was characterised as the evolutionary most stable *trp* protein by the compactness of its phylogenetic tree; see Figure 4.

The three *Euryarchaeota* *Halobacterium* (*s1*), *Haloarcula marismortui* (*s1*) and *Natronomonas pharaonis* (*i1\_s1* species) constituted an isolated group in all five trees (Figures 1, 2, 3, 4); edge lengths were comparable to those of *s2* or *i2\_o2* species. This congruence indicates an elevated evolutionary rate for all elements of these *trp* operons. Note that these operons harbour *trpB1* genes.

**Analysing typical differences in TrpA and TrpB sequences**  
 The phylogenetic tree depicted in Figure 1 illustrates that all TrpB variants can be sorted into four, clearly separated groups. The tree did however not allow to deduce the degree of sequence similarity and to infer whether these differences were subtle sequence variations broadly distributed in the whole sequence or larger indels (inserts or deletions). Table 3 lists the results of pairwise sequence comparisons generated by using BLAST [35]. The selected



**Figure 4**  
**Phylogenetic trees for TrpG and TrpE sequences.** Archaeal and bacterial protein sequences were used to construct the unrooted tree. The last letter of the acronyms indicates the taxonomical position of the species. "E" marks *Euryarchaeota*, "C" *Crenarchaeota*, and "B" bacterial species. The two TrpE sequences of *N. pharaonis* are designated as \_1E, \_2E. For colour code and abbreviations, see legend of Figure 1.

sequences represent the species-types S2, *i2\_o2*, *i1\_o2*, *s1*, and *S1*. As expected, sequence similarity values are in agreement with tree composition. Importantly, for all pairwise comparisons, more than 25% identical residues were determined. Therefore, all TrpB variants should most probably have the same overall 3D-structure [36].

In order to characterise sequence differences in detail, multiple sequence alignments (MSAs) were generated on the basis of a representative selection of TrpA and TrpB sequences. Figure 5 lists for TrpB the MSA, residue conservation, secondary structure and the location of the interface area. Residues interacting with ligands and residues, which are characteristic for TrpB1 and TrpB2 respectively, were labelled. 3D-data were deduced from the X-ray structure of Pfurio\_i1E, *i.e.* the operon-based TrpB1 protein of *P. furiosus* [18], which has PDB code 1WDW. For Ssolfa\_o2C, the 2D-structure was predicted by using Jpred [37]. SDPPred [38] was employed to identify those residues, which separated TrpB1 and TrpB2 due to their skewed or bimodal distribution. In the following, positions and residues are referenced according to the sequence Pfurio\_i1E. Annotations referring active site residues and the interface originate from the PDBsum page

and the Macromolecular Structure Database of the EMBL-EBI.

The MSA shows that nearly all differences between TrpB1 and TrpB2 are due to larger indels, in agreement with [25]. Interestingly, an insertion of 2 to 6 residues between positions 243 and 244 occurred coincidentally in TrpB2\_a and TrpB2\_o sequences, *i.e.* exclusively in non operon-based proteins. All considered TrpB1 and TrpB2\_i sequences lack this subsequence, which was not predicted as a well-defined 2D-element by Jpred. Several representatives belonging to these two sets of operon-based proteins were shown to interact with TrpA [26,39]. Therefore, it is probable that this putative loop influences the allosteric communication with TrpA. Most residues, which are in contact with ligands in the known TrpB1 structure, were strictly conserved among all TrpB1 and TrpB2 sequences. The only exception is residue C225, which is V225 in TrpB2\_a sequences. The active site residues H81, K82, and S371 were strictly conserved, whereas active site residue K162 was conserved only in TrpB1 proteins and active site residue D300 (TrpB1) was an arginine in TrpB2. Several residues of the interface regions, adjacent to active sites and near sites interacting with ligands had a bimodal occurrence pattern distinguishing TrpB1 and TrpB2.

**Table 3: Pairwise sequence similarity values of TrpB proteins**

	Ssolfa_i2C	Ssolfa_o2C	Paerop_i2C	Paerop_o2C	Afulgi_i1E	Afulgi_o2E	Tmarit_i1B	Tmarit_o2B	Mmarip_s1E	Mkand_S1E	Ecoli_s1B
Tacido_S2E	49, 72, 2	76, 88, 0	47, 65, 2	46, 67, 1	26, 44, 17	46, 64, 2	32, 43, 18	47, 66, 2	28, 43, 12	30, 46, 11	26, 42, 6
Ssolfa_i2C	-	54, 75, 2	56, 71, 3	53, 72, 1	34, 48, 14	57, 72, 1	34, 47, 12	54, 73, 1	30, 46, 15	30, 45, 12	28, 42, 14
Ssolfa_o2C		-	50, 66, 2	52, 71, 1	33, 50, 13	47, 68, 1	32, 46, 13	48, 70, 1	28, 43, 16	30, 45, 11	27, 40, 10
Paerop_i2C			-	57, 68, 3	35, 49, 14	54, 63, 3	35, 50, 12	54, 65, 3	32, 48, 12	34, 46, 15	31, 43, 13
Paerop_o2C				-	30, 45, 13	63, 78, 1	32, 46, 13	60, 75, 1	28, 44, 16	33, 46, 11	28, 41, 16
Afulgi_i1E					-	31, 44, 11	65, 78, 0	36, 48, 10	65, 85, 0	65, 78, 0	59, 76, 1
Afulgi_o2E						-	34, 47, 12	64, 76, 0	30, 45, 9	32, 45, 9	29, 40, 12
Tmarit_i1B							-	35, 47, 11	61, 79, 1	64, 79, 1	58, 75, 1
Tmarit_o2B								-	34, 46, 9	32, 46, 8	29, 40, 12
Mmarip_s1E									-	59, 77, 0	58, 77, 0
Mkand_S1E										-	57, 72, 0

Note: The sequences represent species-types S2, i2\_o2, i1\_o2, s1, and S1. All pairs were compared by using BLAST with default parameters. For each pair, the fraction of identical residues, similar residues, and inserted gaps is given in percent. For the generation of protein names, see Results.

Among these were residues 2 and 110, which were strictly conserved tryptophan residues in all TrpB2 proteins. Given its position near the gene start, W2 may assume a function in translation control. W110 succeeds a cluster of strictly conserved residues suggesting a role in stability or protein function.

Figure 6 lists the MSA generated for TrpA sequences. It shows that the active site residues E36, D47, and Y161 are strictly conserved in the TrpA sequences studied. Most evident was a three-residue insertion into TrpA2 sequences following position 125 (numbering deduced from TrpA of *P. furiosus*) as well as deletions at position 162 and between positions 172 and 174. Moreover, most positions showing a bimodal or skewed distribution specific for a *trpA* variant were located near interface regions. In summary, the deviations characterising the two TrpA variants were not as pronounced as those observed in TrpB sequences, however three indels distinguished TrpA1 from TrpA2.

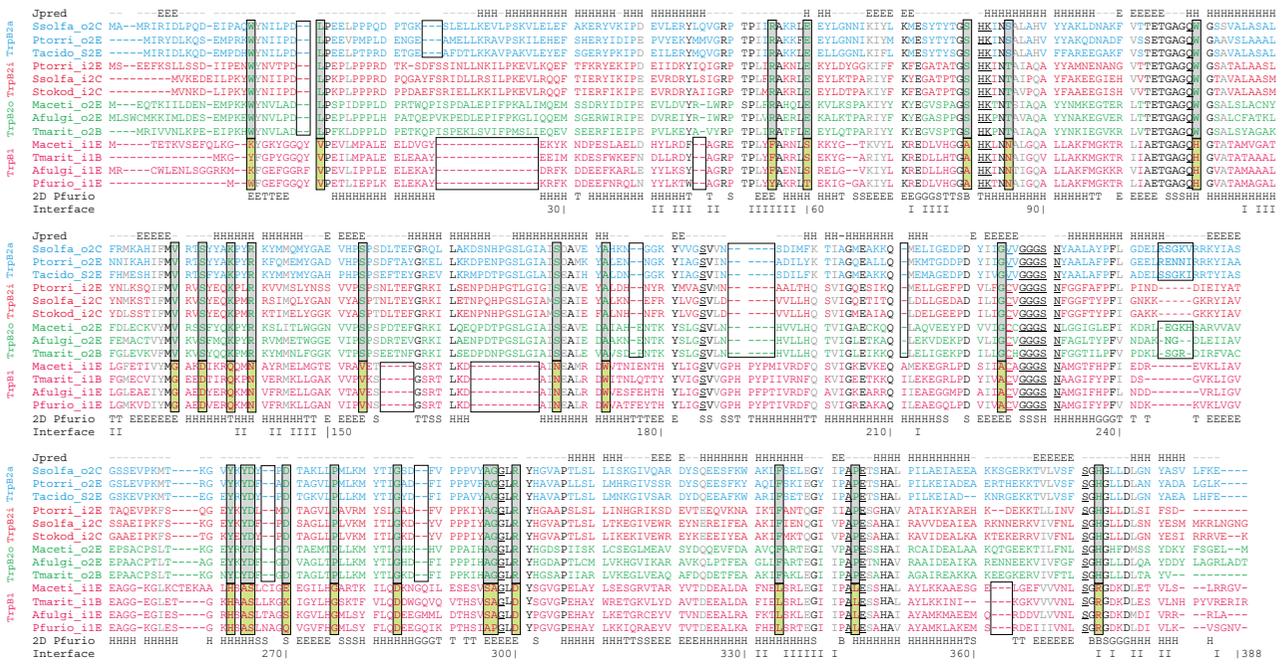
#### Frequency of Trp codons in trpB genes

It has been postulated that the avoidance of tryptophan residues in enzymes for tryptophan synthesis provides a selective advantage [7] as has been shown for a number of amino acid biosynthetic enzymes [40]. This criterion was also applied to the *trpB* genes by assessing the frequency of tryptophan codons (Table 2). *trpB1* genes contained one or two tryptophan codons with a mean value of 1.6 both for S1, s1, and i1\_o2 species. *trpB2* genes contained two tryptophan codons or more with a mean of 2.75 for S2 species, and 3.0 for i2\_o2 species. Most pronounced was the difference for i1\_o2 species. Here, *trpB2* genes had a mean of 3.7, whereas *trpB1* genes had a mean of 1.6 tryptophan codons. These *trpB1* genes showed a habitat-specific imbalance of tryptophan codon occurrence with one in mesophilic species and at least two tryptophan codons in hyperthermophiles. In summary and according to the notion of tryptophan codon avoidance, *trpB2* genes are less optimised than *trpB1* genes.

#### The composition of archaeal trp gene clusters

The evolution of individual genes and operon formation proceed in parallel. For the combined analysis of both processes, gene orders of relevant archaeal and some bacterial *trp* operons were determined and plotted in Figure 7. In most operons, the gene orders *trpBA* and *trpEG*, respectively, were conserved; however, the arrangement of the linkage groups varied. Figure 7 is organised as six panels A – F. Panel A depicts the *trp* clusters of *Thermoplasmataceae*, which are of type *trpA2DFEGC*; *trpB2* lies isolated. In *Sulfolobaceae* and *P. torridus* (Panel B), *trpB2* is the first gene of the gene cluster *trpB2ADFEGC*, which matches the above *trpA2DFEGC* in all positions following *trpB2*. Panel C gives the gene clusters of *A. pernix* and *P. aerophilum*, which possess a *trpB2\_i* and a *trpB2\_o* gene. In the genome of *A. pernix*, two linkage groups *trpA2B2FC* and *trpDEG* occur; *P. aerophilum* possesses the cluster *trpB2DEGA2*. Panel D lists archaeal genomes containing linkage groups *trpCB1A1* and *trpDFEG*. In *Methanosarcina mazei*, these genes form a single cluster, resulting in *trpCB1A1DFEG*. In *N. pharaonis*, these groups are separated by more than 69kb. In panel E, operons are listed where *trpB1* lies close to the 3'-terminal end. For *Thermococcus kodakaraensis*, *Methanococcus maripaludis*, *Archaeoglobus fulgidus* and *Pyrococcus abyssi*, gene order is *trpCDEGFB1A1*. The gene orders in Panel E resemble bacterial operons; two representative examples are plotted in panel F.

It has been argued that simple *trp* clusters may have been unstable until the complexity of regulation and the foundation of a metabolic theme had reached a certain level [10]. Gene clusters observed in s2 and i2\_o2 species can be considered the less evolved stages of cluster organisation; compare Panels A – C. Moreover, the only archaeal *trp* gene regulatory systems identified so far are part of the *trp* operons of *M. thermoautotrophicus* [7] and *T. kodakaraensis* [8], which both have a bacterial-like composition.



**Figure 5**  
**Multiple sequence alignment of TrpB sequences.** Representatives of the four groups of TrpB sequences were aligned. *Ssolfa\_o2C*, *Ptorri\_o2E*, and *Tacido\_S2E* represent TrpB2\_a sequences. *Ptorri\_i2E*, *Ssolfa\_i2C*, and *Stokod\_i2C* represent TrpB2\_i, *Maceti\_o2E*, *Afulgi\_o2E*, and *Tmarit\_o2B* represent TrpB2\_o sequences. *Maceti\_i1E*, *Tmarit\_i1B*, *Afulgi\_i1E*, and *Pfurio\_i1E* represent TrpB1 sequences. The 2D-structural elements of *Pfurio\_i1E*, as deduced from the PDB file IWDW, are shown below the sequences, and residues involved in protein interaction with TrpA (I) are assigned under 'Interface'. The line *Jpred* (top) lists a 2D-prediction of *Ssolfa\_o2C* generated by using the *Jpred* server [37]. Residues in bold face printing are conserved; black residues are strictly, grey residues are less strictly conserved. Active site residues are plotted in italics; residues in contact with ligands are underlined. These data were deduced from the PDBsum pages [56] and the PISA server [58] of the EMBL-EBI. Residues printed in boxes were predicted by SDPpred [38] as being specific for TrpB1 or TrpB2. See legend of Figure 1 for an explanation of sequence acronyms.

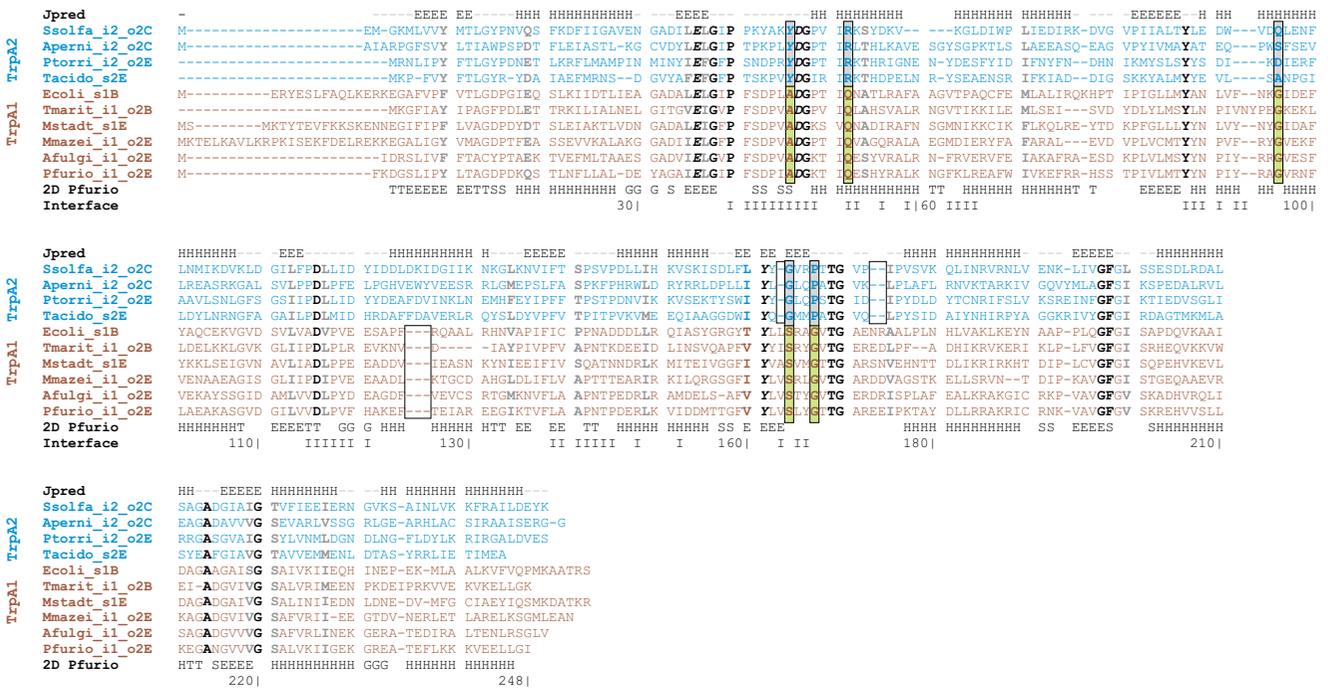
Besides *Nanoarchaeum equitans*, *Thermoplasma* (*T. volcanium*, *T. acidophilum*, and *P. torridus*) possess the smallest archaeal genomes sequenced so far. Most plausibly, strong selective pressure associated with the colonised habitat enforces the minimisation of genome size. However, both *Thermoplasma* species possess the gene cluster *trpA2DFEGC*. Therefore, the need for tryptophan synthesis can be taken for granted. The separation of *trpB2* from the remaining *trp* genes is consistent with a demand for individual gene regulation and expression presumably to an additional function of TrpB2. Most plausibly, under these constraints, *trpB2* is the more optimal variant, which is in a specific environment favoured over *trpB1*.

**What is the origin of trpB genes?**

Recently, TrpA, Tmari\_i1B and Tmari\_o2B of *T. maritima* have been produced in *E. coli*, purified, and characterised [39]. It has been shown that recombinant TrpA forms an  $\alpha$ -monomer, and that both recombinant TrpB proteins form  $\beta_2$ -homodimers. However, only the operon-

encoded Tmari\_i1B – but not Tmari\_o2B – associated with TrpA to constitute the conventional  $\alpha\beta\beta\alpha$  tryptophan synthase complex in which both subunits reciprocally activate each other. An analogous experiment has been carried out for genes of *S. solfataricus* [26]. The results have shown that *Ssolfa\_i2C* – but not *Ssolfa\_o2C* – associates transiently with TrpA during catalysis to form a functional tryptophan synthase complex. However, in contrast to regular tryptophan synthases, the affinity between the two subunit-types was weak, and activation has been unidirectional from *Ssolfo\_i2C* to TrpA. These results indicate the following ranking for the binding-affinity to TrpA: TrpB2\_o < TrpB2\_i < TrpB1.

In the course of modelling *trpB* evolution, the relationship between the *trpB* variants has to be made plausible. A possible explanation for the existence of two *trpB* variants would be convergent evolution, i.e. the independent development of *trpB1* and *trpB2* towards a *trpB* gene. In this case, few residues, which are critical for function,



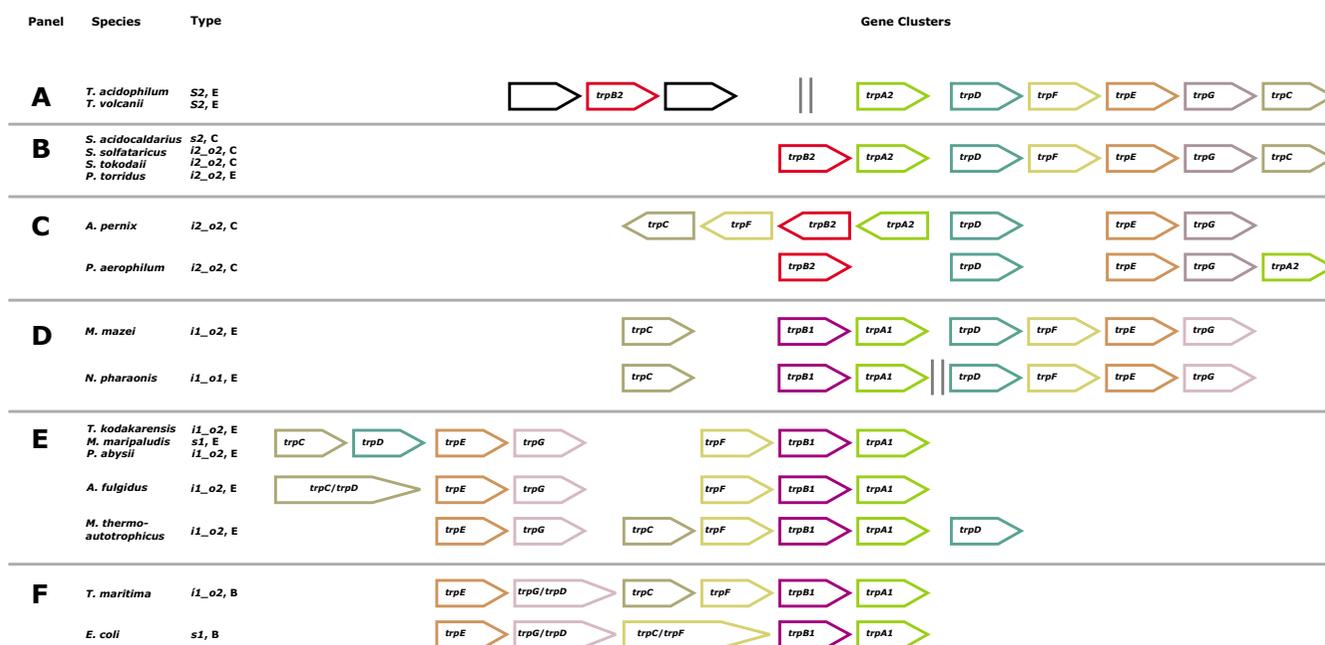
**Figure 6**  
**Multiple sequence alignment of TrpA sequences.** Ssolfa\_i2\_o2C, Aperni\_i2\_o2C, and Ptorri\_i2\_o2C represent TrpA sequences from i2\_o2 species possessing both an operon-based and a non operon-based *trpB2*. Tacido\_s2E represents a species having exclusively an operon-based *trpB2*. Ecoli\_s1B and Tmarit\_i1\_o2B represent bacterial TrpA proteins. Mstادت\_s1E is from a species possessing exclusively an operon-based *trpB1* gene, Mmazei\_i1\_o2E, Afulgi\_i1\_o2E and Pfurio\_i1\_o2E are TrpA sequences from i1\_o2 species possessing an operon-based *trpB1* and a non operon-based *trpB2* gene. Presumably, these TrpA1 proteins interact with a protein of type TrpB1. Below the alignment, the 2D-structure of TrpA of *P. furiosus* (Pfurio\_i1\_o2E), and residues involved in protein interaction with its TrpB1 (I) are given. The line named Jpred lists a 2D-prediction of Ssolfa\_i2\_o2C generated by using the Jpred server [37]. Residues printed in bold are conserved; black residues are strictly, grey residues are less strictly conserved. Active site residues are plotted in italics. These data were deduced from the PDBsum pages [56] and the PISA server [58] of the EBI. Residues printed in boxes were predicted by SDPpred [38] as being specific for the two TrpA species.

should correspond. However, one would expect these residues embedded into polypeptides, which are relatively dissimilar on the sequence level. In contrast, comparison of TrpB1 and TrpB2 sequences shows that on average 30% of the residues are identical and 40% are similar; compare Table 3. This finding and the conservation of indels makes convergent evolution highly improbable and argues for a common origin of *trpB1* and *trpB2* genes.

The most-widely accepted model for the evolution of novel protein functions postulates gene duplication and the generation of a redundant gene copy [41]. It is assumed that evolutionary stress for a copy is largely reduced thus facilitating the evolution of a paralogue with a novel function. This model is based on the notion that negative trade-offs dominate evolutionary processes [42]. According to this model of evolution, one of the *trpB* genes originates from a copy of the ancestral variant.

Which of the two existing variants represents the more ancient gene? The arguments listed below suggest that *trpB2* is the ancestral *trpB* gene.

- i) *trpB1* is not universally distributed among archaea. *Crenarchaeota* possess exclusively *trpB2* genes.
- ii) A low frequency of amino acids in enzymes required for their synthesis provides selective advantage [40]. In general, *trpB1* genes contain fewer tryptophan codons than *trpB2* genes; in *i1\_o2* species, the ratio is 1.6/3.7 *i.e.* less than 0.5. Therefore, *trpB1* is the more evolved gene.
- iii) The sophisticated inter-subunit communication suggests that the products of *trpB1* and *trpA1* of species-types *s1* or *i1\_o2* are the most efficient enzymes; see [39] and references therein. Hence, TrpB1 is the more optimised and later evolved TrpB variant.
- iv) It has been postulated that ancient enzymes possess broad specificities [43]. The occurrence of *trpB2* outside *trp* operons argues for either a



**Figure 7**  
**Gene organisation of archaeal and bacterial *trp* gene clusters.** Each panel A – F represents the occurrence and orientation of *trp* genes in the genomes of organisms listed in the second column. The third column gives the species-type and the taxonomical lineage. "E" marks *Euryarchaeota*, "C" *Crenarchaeota*, and "B" bacteria. A vertical double line (Panels A and D) indicates borders of gene clusters separately located in the genome. Open arrows represent hypothetical genes. The arrows are not to scale; gaps of arbitrary length were inserted between genes to allow the alignment of arrows. For acronyms of species-types, see legend of Table 2.

new function or a broader specificity. In summary, it is plausible to regard *trpB2* as representing the more ancient variant of *trpB*.

**Modelling the evolution of *TrpB***

In order to reduce the number of possible alternative scenarios that have to be discussed for modelling the evolution of *trpB*, the following assumptions were made:

i) For bacteria, an ancestral *trp* operon of type *trpEGDCFB1A1* is most likely [10]. Therefore, the existence of a *trpB1* gene in the bacterial predecessor was taken for granted. In addition, it has been concluded for bacterial *trp* operons that horizontal gene transfer (HGT) did not affect the path of evolutionary history [44].

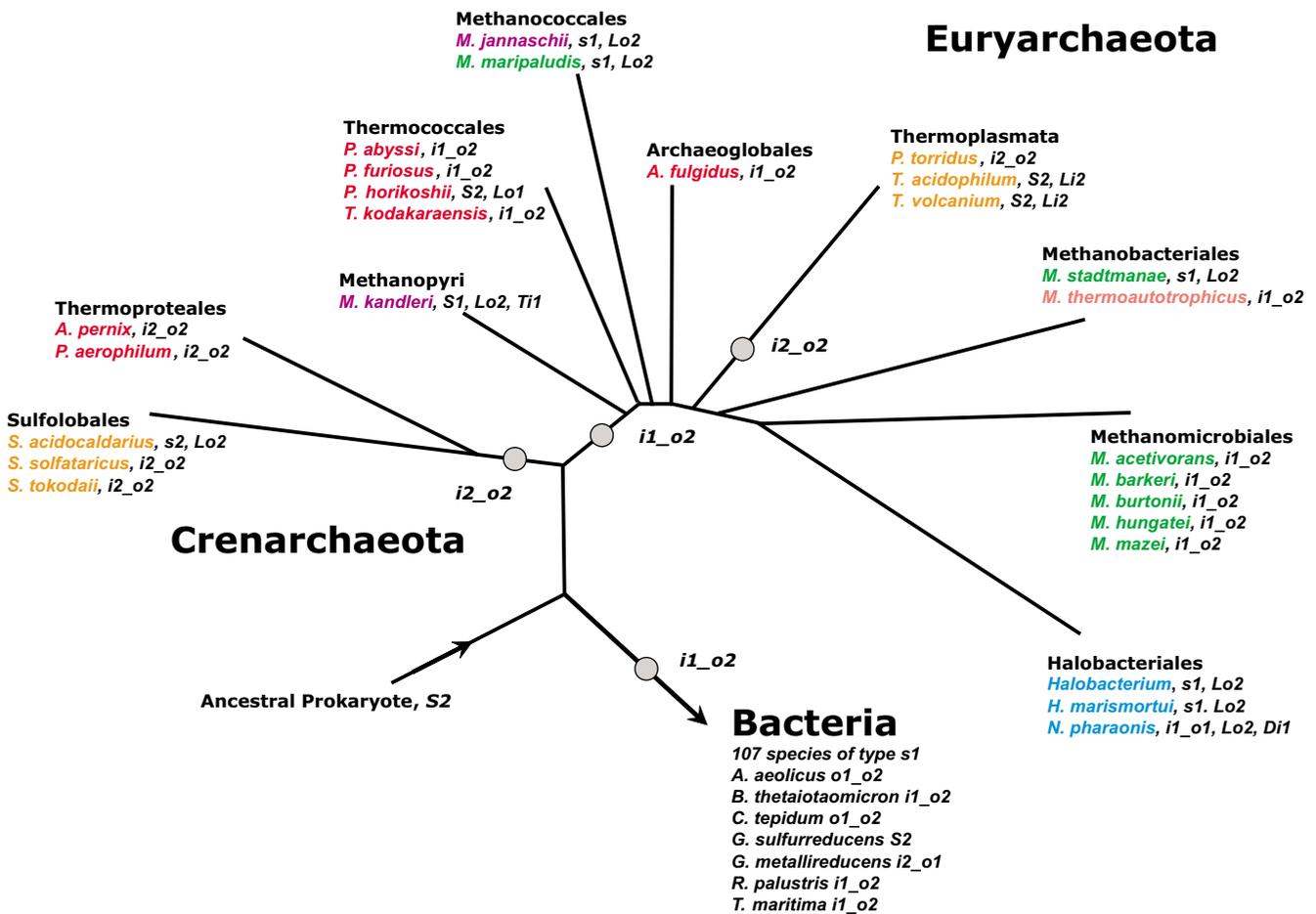
ii) *trpB1*, *trpB2*, *trpA1* and *trpA2* have been invented only once. The analysis of multiple sequence alignments (see Figures 5 and 6) shows that the main differences distinguishing the variants are conserved indels. It has been convincingly argued that conserved indels result less likely than e.g. point mutations from independent mutational events and provide useful milestones for the identification of evolutionary phases [45]. In addition, the strong coherence seen in the *TrpB* subtree argues against an independ-

ent evolution occurring in parallel for bacteria and archaea. Due to the existence of conserved indels, an evolutionary process *trpB2<sub>i</sub> → trpB1 → trpB2<sub>o</sub>* or *vice versa* is unlikely too.

iii) As has been deduced previously [46], the following order of importance was taken for the processes of genome evolution: gene loss > gene genesis > gene duplication > HGT.

iv) The integration of a *trpB* gene into the *trp* operon (or linkage group) was rated less probable than other translocations, gene duplications, gene loss, and mutations. It is presumably very rare that a particular gene gets integrated into a specific gene cluster [47], which is the *trp* operon in the considered case.

v) It is unlikely that several recent events of HGT explain the taxonomically widespread occurrence of *trpB2* genes in bacteria. In bacteria, *trpB2* genes were found in hyperthermophilic (*Aquificae* and *Thermotogae*) and mesophilic bacteria belonging to the taxonomical groups of *Alpha*- and *Gammaproteobacteria* and *Bacteroides*. The program SIGI [48] identifies genomic islands, i.e. gene clusters having a conspicuous codon usage indicating recent HGT



**Figure 8**

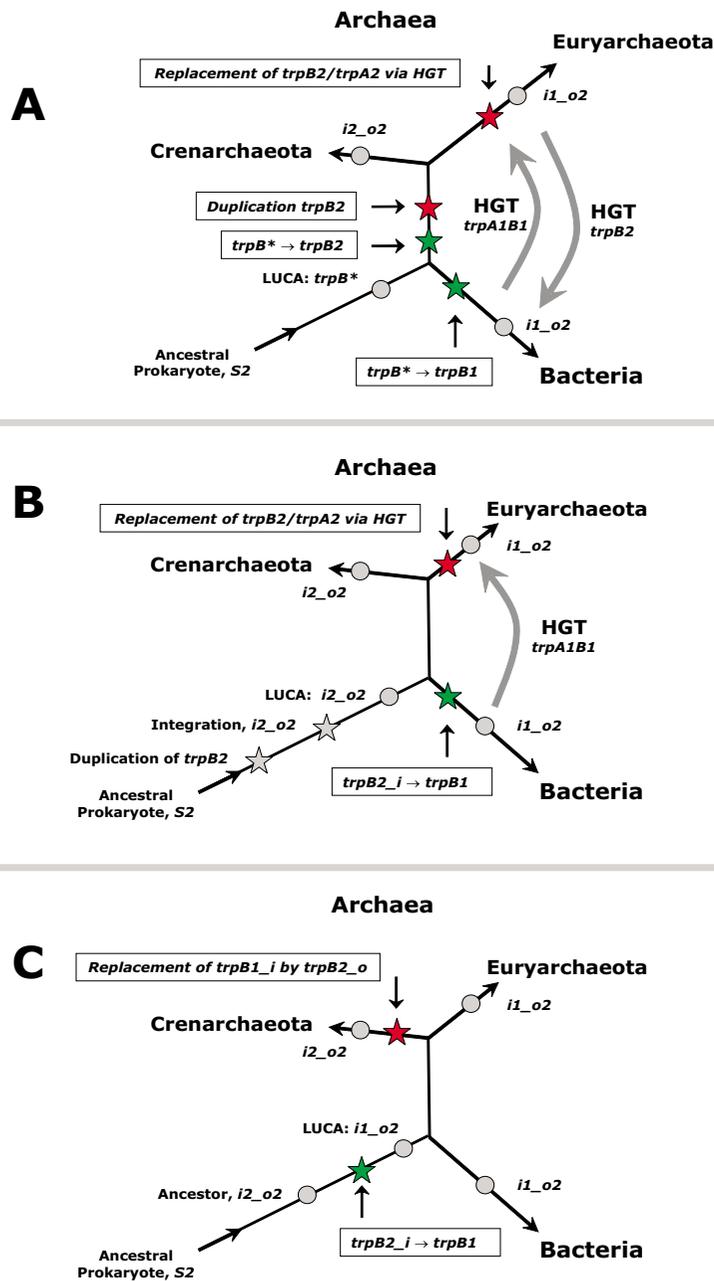
**A parsimonious reconstruction of predecessors.** The phylogenetic tree is based on 16S rRNA sequence comparisons (after Fig. 2 of [49]). For all modern species, their species-type and evolutionary events leading most plausibly from the ancestral predecessor to the current genome content are added. The most probable species-type of predecessors for *Crenarchaeota*, *Euryarchaeota*, and *Bacteria* is given next to the grey circles. Abbreviations for events changing genomic content: Li1 or Lo2, (L)oss of the operon-based *trpB1* or the non operon-based *trpB2* gene, respectively. Ti1 (T)ranslocation of the operon based *trpB1*, Di1 (D)uplication of the operon-based *trpB1*. The colour of species names indicates the habitat: Hyperthermophiles are given in red, thermoacidophiles in orange, thermophiles in pink, mesophiles in green, halophiles in blue, and species living in a both hyperthermophilic and halophilic environment are given in purple.

events. In none of the considered genomes were *trpB1* or *trpB2* genes (both inside and outside operons) elements of such islands.

In order to model the evolution of tryptophan synthase, a phylogenetic tree based on archaeal 16S rRNA sequence comparisons was plotted according to Fig. 2 from [49]. All considered species and their species-types were added. Using the above premises, the most plausible sequence-types of predecessors were determined. These types and evolutionary events needed to infer the modern species-

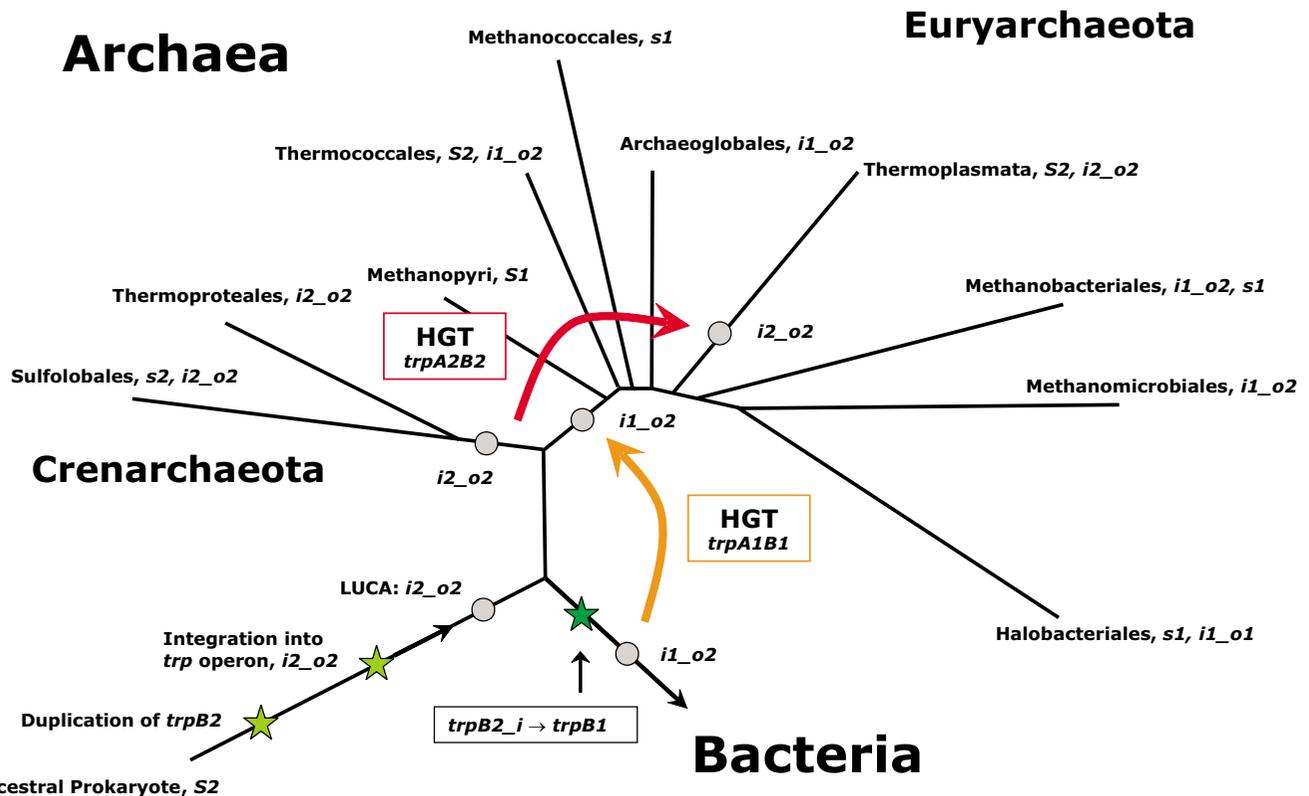
types from the predecessors were added to the tree; see Figure 8.

The most plausible predecessor of all *Crenarchaeota* is of type *i2\_o2*; for *Bacteria* and for *Euryarchaeota* it is of type *i1\_o2*. Assuming this and excluding *Thermoplasmata* (see below), of the 23 modern archaeal species, 14 have the same species-type as their ancestor. Of the 9 species possessing a deviating type, 7 can be explained with a single gene loss, and for only 2 modern species a more complicated genomic rearrangement has to be postulated: Loss of *trpB2* and dislocation of *trpB1* has to be postulated for



**Figure 9**

**Alternative models of *trpB* evolution.** Model A assumes that a single and intermediate *trpB\** gene existed in the last universal common ancestor (LUCA) of bacteria and archaea. The evolution of the *trpB2* gene is considered an archael and that of the *trpB1* gene is considered a bacterial invention. The occurrence of *trpA1* and *trpB1* genes in archaea and of *trpB2* genes in bacteria are explained by a twofold horizontal gene transfer (HGT). A duplication of *trpB2* in an ancient archael genome has been postulated to explain the existence of the non operon-based *trpB2*. Models B and C propose two alternatives for the evolution of the LUCA. Model B assumes that the evolution *trpB2* → *trpB1* occurred in an early bacterial species after the divergence of bacteria and archaea. The replacement of linkage group *trpB2A2* by *trpB1A1* via HGT was postulated to account for the euryarchaeal predecessor of type *i1\_o2*. Model C assumes that the evolution *trpB2* → *trpB1* occurred before the divergence of bacteria and archaea. Hence, the replacement of an operon-based *trpB1* by a *trpB2* gene and the evolution *trpA1* → *trpA2* was postulated for the crenarchaeal ancestor. For acronyms of species-types, see legend of Table 2. Distances are arbitrary and do not represent evolutionary time intervals. Stars indicate events of genomic rearrangements, circles filled in grey represent ancient predecessors.



**Figure 10**  
**Composite model of *trpB* evolution.** Upon duplication and integration of an ancient *trpB2* gene into the *trp* operon, the last universal common ancestor (LUCA) of bacteria and archaea was of species-type *i2\_o2*. In a bacterial ancestor, the evolution of a linkage group *trpB1A1* occurred. Via horizontal gene transfer (HGT), an euryarchaeal ancestor acquired this linkage group, which gave rise to a predecessor of type *i1\_o2*. *Thermoplasmata* acquired *trpA2* and *trpB2* genes in an ancient event of HGT. For all taxonomical orders, species-types of current species are given. S2 species possess exactly one, non operon-based *trpB2* gene, s2: ditto, the gene is located inside the *trp* operon. *trpB1* was treated analogously. *i2\_o2* are species possessing a *trpB2* gene inside and a second *trpB2* outside the operon, *i1\_o2* are species with an operon-based *trpB1* and a non operon-based *trpB2*, and *i1\_o1* are species possessing an operon-based and at least one non operon-based *trpB1*.

*M. kandleri* (representing *Methanopyri*), which is a S1 species. The replacement of *trpB2\_o* with a copy of *trpB1* is necessary to explain the *i1\_o1* genome of *N. pharaonis*. The only euryarchaeal class requiring a more complex explanation than gene loss and translocation subsumes *Thermoplasmata*, which possess exclusively *trpB2* and *trpA2* genes. The composition of congruency groups (compare Figures 1 and 2) makes a common evolution with *Sulfolobales* or the acquisition of the same *trp* genes probable. The similarity of operon structures supports this assumption: operon structures of *P. torridus* and *Sulfolobales* are identical (compare Panel B of Figure 7). For *T. acidophilum*, a large amount of HGT with *S. solfataricus*, which is found in the same habitat, has been made plausible [50]. In summary, a common evolutionary history of *trpB2* and *trpA2* genes of *Sulfolobales* and *Thermoplasmata* is highly plausible, proposing for both taxonomical classes an ancestor of species-type *i2\_o2*. Assuming an *i2\_o2* ances-

tor, gene loss is sufficient to explain the genome composition of all modern *Thermoplasmata*.

Based on these predecessors, three alternatives explaining the evolution and distribution of *trpB* species starting from the last universal common ancestor (LUCA) of bacteria and archaea were deduced (Figure 9, Panels A – C). In the following paragraph, the plausibility of these alternatives will be discussed. The rest of this paragraph is used to elucidate the three alternatives.

In Panel A of Figure 9, the existence of an ancestral *trpB\**, an intermediate of *trpB1* and *trpB2* was postulated for the LUCA. *trpB\** might then have diverged into a bacterial *trpB1* and an archaeal *trpB2* variant. To explain the existence of a non operon-based *trpB2* in archaea, a duplication of *trpB2* is necessary. The advent of an euryarchaeal *i1\_o2* predecessor requires the replacement of linkage

group *trpB2A2* with *trpB1A1* via HGT from bacteria to archaea. The occurrence of *trpB2* in bacterial genomes demands an early transfer of *trpB2* from an archaeal to a bacterial predecessor.

Panel B of Figure 9 depicts an alternative model for the evolution of the LUCA towards the bacterial and archaeal ancestors. As introduced above, gene duplication is regarded the first step for evolving a novel gene function. In addition, *trpB2* must be considered to represent the more ancient variant of *trpB*. Therefore, the evolution towards the LUCA of bacteria and archaea is most plausibly explained by the duplication of a non operon-based *trpB2* gene, which was subsequently integrated into the *trp* operon and constituted an ancient linkage group *trpB2A2*. This makes a common ancestor of type *i2\_o2* plausible. These considerations are the basis for further reconstructing the evolution of predecessors. In Panels B and C, two alternatives are given.

In Panel B, it is assumed that the LUCA was of type *i2\_o2* and that the evolution *trpB2* → *trpB1* occurred in an early bacterial species. In this case, species-types of the LUCA and the crenarchaeal predecessor are identical. To explain the advent of an euryarchaeal predecessor of type *i1\_o2*, an ancient event of HGT from *Bacteria* to *Archaea* has to be postulated for the acquisition of the linkage group *trpB1A1*, which replaced *trpB2A2*.

In Panel C, it is assumed that the LUCA was of type *i1\_o2*, i.e. the evolution *trpB2* → *trpB1* occurred earlier than the speciation of *Bacteria* and *Archaea*. In this case, the species-types of the LUCA and the predecessors of *Bacteria* and *Euryarchaeota* are identical. However, a replacement of *trpB2\_i* by *trpB2\_o* is necessary to constitute the crenarchaeal predecessor.

#### How plausible are these three models?

Model A requires at least two ancient events of HGT to explain the occurrence of *trpB2* in *Bacteria* and of *trpA1B1* in *Euryarchaeota*. The phenomenon of non-orthologous displacement *in situ* is well-characterised [51,52]. In addition to HGT, a duplication of the *trpB2* gene is needed for the predecessor of *Archaea*. This model is not the most parsimonious one: Model B demands only one HGT event, the ancient acquisition of the linkage group *trpB1A1* by an euryarchaeal predecessor.

Model C postulates a LUCA of species-type *i1\_o2*. The sophisticated inter-subunit communication clearly suggests that products of *trpB1* and *trpA1* genes are the most specialised and most recently evolved tryptophan synthases; see [39] and references therein. Thus, the replacement of *trpB1* with *trpB2*, which is needed to explain the existence of a crenarchaeal predecessor of type *i2\_o2*,

would – with respect to protein-protein interaction – lead to a less optimal tryptophan synthase. This seems unlikely, if one presumes the sustained need for tryptophan synthesis in *Crenarchaeota*.

In contrast, model B postulates the replacement of a (less evolved) *trpB2\_i* by a *trpB1* for the euryarchaeal predecessor and does not require the replacement of a *trpB1* by a *trpB2* for the crenarchaeal predecessor. The case of *Thermoplasma* makes clear that in a thermophilic or hyperthermophilic environment *trpB2* and *trpA2* genes are favoured over *trpB1* and *trpA1*. There is evidence that the LUCA was a thermophilic or hyperthermophilic species [34,53,54]. Therefore, it is more probable to expect a LUCA of species-type *i2\_o2*. In summary, considering parsimony arguments and the assumption that negative trade-offs dominate evolutionary processes [42], model B is the likelier one. Figure 10 summarises the most parsimonious scenario explaining the composition of modern archaeal *trp* operons: Assuming that the LUCA was of type *i2\_o2*, and that *trpB1* was a bacterial invention, besides gene loss, which is a frequent evolutionary event, two cases of ancient HGT are sufficient to explain the distribution of *trpA* and *trpB* species in current archaeal genomes.

#### Conclusion

In archaeal genomes, various stages of *trpB* function have been conserved. Most plausibly, *trpB2* represents the ancestral variant of *trpB* genes. With respect to TrpA/TrpB communication and cooperativity, the situation observed in *S2* species (*T. acidophilum* and *T. volcanium*) is probably the least complex one. Similarly archaic are the non operon-based *trpB2* genes of *Sulfolobaceae*, whereas the operon-based *trpB* genes are more evolved. *s1* and *i1\_o2* species possess highly cooperative synthases. Thus, the archaeal tryptophan synthase (especially *trpB* variants) constitutes a model system for the study of protein complex formation. Due to different environmental conditions, several stages of cooperativity have been conserved, which allow to characterise the progress of *trpA* – *trpB* coevolution based on gene expression and on functional cooperativity.

#### Materials

##### Genomes and protein sequences

Genomic content was determined by analysing version 6.2 of the STRING database [30].

All protein sequences were downloaded via the "Genome Project" database of the NCBI [55], which allows to access completely sequenced genomes. Respective COG tables were consulted to determine the COG group of genes [29] and to download sequences. Genes originating from the following completely sequenced genomes were analysed

(abbreviations used for Figures and accession numbers of genomes in brackets):

#### Crenarchaeota

*Aeropyrum pernix* K1 (Aperni, NC\_00854), *Pyrobaculum aerophilum* str. IM2 (Paerop, NC\_003364), *Sulfolobus acidocaldarius* DSM 639 (Sacido, NC\_007181), *Sulfolobus solfataricus* P2 (Ssolfa, NC\_002754), *Sulfolobus tokodaii* str. 7 (Stokod, NC\_003106).

#### Euryarchaeota

*Archaeoglobus fulgidus* DSM 4304 (Afulgi, NC\_000917), *Haloarcula marismortui* ATCC 43049 (Hmaris, NC\_006396), *Halobacterium* sp. NRC-1 (Halob, NC\_002607), *Methanocaldococcus jannaschii* DSM 2661 (Mjanna, NC\_000909), *Methanococcoides burtonii* DSM 6242 (Mburto, NC\_007955), *Methanococcus maripaludis* S2 (Mmarip, NC\_005791), *Methanopyrus kandleri* AV19 (Mkandl, NC\_003551), *Methanosarcina acetivorans* C2A (Maceti, NC\_003552), *Methanosarcina barkeri* str. Fusaro (Mbarke, NC\_007355), *Methanosarcina mazei* Go1 (Mmazei, NC\_003901), *Methanosphaera stadtmanae* DSM 3091 (Mstadt, NC\_007681), *Methanospirillum hungatei* JF-1 (Mhunga, NC\_007796) *Methanothermobacter thermautotrophicus* str. Delta H. (Mtherm, NC\_000916), *Natronomonas pharaonis* DSM 2160 (Nphara, NC\_007426), *Picrophilus torridus* DSM9790 (Ptorri, NC\_005877), *Pyrococcus abyssi* GE5 (Pabyss, NC\_000868), *Pyrococcus furiosus* DSM 3638 (Pfurio, NC\_003413), *Pyrococcus horikoshii* OT3 (Phorik, NC\_000961), *Thermococcus kodakaraensis* KOD1 (Tkodak, NC\_006624), *Thermoplasma acidophilum* DSM1728 (Tacido, NC\_002578), *Thermoplasma volcanium* GSS1 (Tvolca, NC\_002689).

#### Bacteria

*Escherichia coli* K-12 (Ecoli, NC\_000913), *Geobacter metalireducens* GS-15 (Gmetal, NC\_007517), *Geobacter sulfurreducens* PCA (Gsulfu, NC\_002939), *Thermotoga maritima* (Tmarit, NC\_00853).

#### Methods

##### Generating multiple sequence alignments

For the generation of multiple sequence alignments (MSAs) the program M-Coffee [32] was used. It combines the output of nine individual MSA methods for the generation of a "meta"-MSA. M-Coffee has been shown to outperform all individual methods of MSA generation [32].

##### Annotating multiple sequence alignments

For each position in a MSA, residue conservation, secondary structure, the location of the interface area, active sites and residues, which are characteristic for sequence types, were determined and plotted. 3D-data were deduced from the PDB-file 1WDW, describing the TrpA/TrpB complex of *P. furiosus* [18]. For 2D-structure prediction, Jpred [37]

was used. SDPpred [38] was utilised to identify those residues, which distinguished sequence groups due to their skewed or bimodal distribution. Annotations referring active site residues were deduced from the PDBsum page [56,57], interface residues were annotated according to the *Protein interfaces, surfaces and assemblies service PISA* [58,59]. Both services were located at the webserver of the European Bioinformatics Institute (EMBL-EBI).

##### Creating and evaluating phylogenetic trees

SplitsTrees4 [33], a frame-work for phylogenetic analyses, was used to generate and analyse phylogenetic trees. MSAs originating from M-Coffee were utilised to calculate maximum likelihood protein distance estimates based on a JTT [60] model. The bio-neighbour joining approach [61] was used to generate trees. Resulting trees were analysed by bootstrapping (1000 replications each).

##### Acknowledgements

I want to thank Reinhard Sterner and Rüdiger Schmitt for stimulating discussions and their help in preparing this manuscript.

##### References

1. Yanofsky C: **Using studies on tryptophan metabolism to answer basic biological questions.** *J Biol Chem* 2003, **278(13)**:10859-10878.
2. Yanofsky C: **Advancing our knowledge in biochemistry, genetics, and microbiology through studies on tryptophan metabolism.** *Annu Rev Biochem* 2001, **70**:1-37.
3. Yanofsky C: **Attenuation in the control of expression of bacterial operons.** *Nature* 1981, **289(5800)**:751-758.
4. Yanofsky C: **The different roles of tryptophan transfer RNA in regulating trp operon expression in E. coli versus B. subtilis.** *Trends Genet* 2004, **20(8)**:367-374.
5. Gutierrez-Preciado A, Jensen RA, Yanofsky C, Merino E: **New insights into regulation of the tryptophan biosynthetic operon in Gram-positive bacteria.** *Trends Genet* 2005, **21(8)**:432-436.
6. Gast DA, Jenal U, Wasserfallen A, Leisinger T: **Regulation of tryptophan biosynthesis in Methanobacterium thermoautotrophicum Marburg.** *J Bacteriol* 1994, **176(15)**:4590-4596.
7. Xie Y, Reeve JN: **Regulation of tryptophan operon expression in the archaeon Methanothermobacter thermautotrophicus.** *J Bacteriol* 2005, **187(18)**:6419-6429.
8. Tang X, Ezaki S, Fujiwara S, Takagi M, Atomi H, Imanaka T: **The tryptophan biosynthesis gene cluster trpCDEGFBA from Pyrococcus kodakaraensis KOD1 is regulated at the transcriptional level and expressed as a single mRNA.** *Mol Gen Genet* 1999, **262(4-5)**:815-821.
9. Bentley R: **The shikimate pathway--a metabolic tree with many branches.** *Crit Rev Biochem Mol Biol* 1990, **25(5)**:307-384.
10. Xie G, Keyhani NO, Bonner CA, Jensen RA: **Ancient origin of the tryptophan operon and the dynamics of evolutionary change.** *Microbiol Mol Biol Rev* 2003, **67(3)**:303-342.
11. Dunn MF, Aguilar V, Brzovic P, Drewe WF Jr., Houben KF, Leja CA, Roy M: **The tryptophan synthase holoenzyme complex transfers indole between the a- and b-sites via a 25-30 Å long tunnel.** *Biochemistry* 1990, **29(37)**:8598-8607.
12. Weber-Ban E, Hur O, Bagwell C, Banik U, Yang LH, Miles EW, Dunn MF: **Investigation of allosteric linkages in the regulation of tryptophan synthase: the roles of salt bridges and monovalent cations probed by site-directed mutation, optical spectroscopy, and kinetics.** *Biochemistry* 2001, **40(12)**:3497-3511.
13. Zalkin H, Smith JL: **Enzymes utilizing glutamine as an amide donor.** *Adv Enzymol Relat Areas Mol Biol* 1998, **72**:87-144.
14. Wilmanns M, Hyde CC, Davies DR, Kirschner K, Jansonius JN: **Structural conservation in parallel b/a-barrel enzymes that**

- catalyze three sequential reactions in the pathway of tryptophan biosynthesis. *Biochemistry* 1991, **30(38)**:9161-9169.
15. Sterner R, Höcker B: **Catalytic versatility, stability, and evolution of the (ba)8-barrel enzyme fold.** *Chem Rev* 2005, **105(11)**:4038-4055.
  16. Brown JR, Doolittle WF: **Archaea and the prokaryote-to-eukaryote transition.** *Microbiol Mol Biol Rev* 1997, **61(4)**:456-502.
  17. Miles EW: **Tryptophan synthase: a multienzyme complex with an intramolecular tunnel.** *Chem Rec* 2001, **1(2)**:140-151.
  18. Lee SJ, Ogasahara K, Ma J, Nishio K, Ishida M, Yamagata Y, Tsukihara T, Yutani K: **Conformational changes in the tryptophan synthase from a hyperthermophile upon a2b2 complex formation: crystal structure of the complex.** *Biochemistry* 2005, **44(34)**:11417-11427.
  19. Hioki Y, Ogasahara K, Lee SJ, Ma J, Ishida M, Yamagata Y, Matsuura Y, Ota M, Ikeguchi M, Kuramitsu S, Yutani K: **The crystal structure of the tryptophan synthase b2 subunit from the hyperthermophile *Pyrococcus furiosus*. Investigation of stabilization factors.** *Eur J Biochem* 2004, **271(13)**:2624-2635.
  20. Kirschner K, Lane AN, Strasser AW: **Reciprocal communication between the lyase and synthase active sites of the tryptophan synthase holoenzyme complex.** *Biochemistry* 1991, **30(2)**:472-478.
  21. Miles EW, Rhee S, Davies DR: **The molecular basis of substrate channeling.** *J Biol Chem* 1999, **274(18)**:12193-12196.
  22. Schneider TR, Gerhardt E, Lee M, Liang PH, Anderson KS, Schlichting I: **Loop closure and intersubunit communication in tryptophan synthase.** *Biochemistry* 1998, **37(16)**:5394-5406.
  23. Weyand M, Schlichting I: **Crystal structure of wild-type tryptophan synthase complexed with the natural substrate indole-3-glycerol phosphate.** *Biochemistry* 1999, **38(50)**:16469-16480.
  24. Osborne A, Teng Q, Miles EW, Phillips RS: **Detection of open and closed conformations of tryptophan synthase by 15N-heteronuclear single-quantum coherence nuclear magnetic resonance of bound L-tryptophan.** *J Biol Chem* 2003, **278(45)**:44083-44090.
  25. Xie G, Forst C, Bonner C, Jensen RA: **Significance of two distinct types of tryptophan synthase beta chain in bacteria, archaea and higher plants.** *Genome Biol* 2002, **3(1)**:RESEARCH0004.
  26. Leopoldseder S, Hettwer S, Sterner R: **Evolution of multi-enzyme complexes: the case of tryptophan synthase.** *Biochemistry* 2006, **45(47)**:14111-14119.
  27. Merkl R: **AMIGOS: a method for the inspection of genomic organisation or structure and its application to characterise and dissect conserved gene arrangements.** *In Silico Biol* 2006, **6**:281-306.
  28. Overbeek R, Fonstein M, D'Souza M, Pusch GD, Maltsev N: **The use of gene clusters to infer functional coupling.** *Proc Natl Acad Sci U S A* 1999, **96(6)**:2896-2901.
  29. Tatusov RL, Fedorova ND, Jackson JD, Jacobs AR, Kiryutin B, Koonin EV, Krylov DM, Mazumder R, Mekhedov SL, Nikolskaya AN, Rao BS, Smirnov S, Sverdlov AV, Vasudevan S, Wolf YI, Yin JJ, Natale DA: **The COG database: an updated version includes Eukaryotes.** *BMC Bioinformatics* 2003, **4(1)**:41.
  30. von Mering C, Jensen LJ, Snel B, Hooper SD, Krupp M, Foglierini M, Jouffre N, Huynen MA, Bork P: **STRING: known and predicted protein-protein associations, integrated and transferred across organisms.** *Nucleic Acids Res* 2005, **33(Database issue)**:D433-7.
  31. Methe BA, Nelson KE, Eisen JA, Paulsen IT, Nelson WW, Heidelberg JF, Wu D, Wu M, Ward N, Beanan MJ, Dodson RJ, Madupu R, Brinkac LM, Daugherty SC, DeBoy RT, Durkin AS, Gwinn M, Kolonay JF, Sullivan SA, Haft DH, Selengut J, Davidsen TM, Zafar N, White O, Tran B, Romero C, Forberger HA, Weidman J, Khouri H, Feldblyum TV, Utterback TR, Van Aken SE, Lovley DR, Fraser CM: **Genome of *Geobacter sulfurreducens*: metal reduction in subsurface environments.** *Science* 2003, **302(5652)**:1967-1969.
  32. Wallace IM, O'Sullivan O, Higgins DG, Notredame C: **M-Coffee: combining multiple sequence alignment methods with T-Coffee.** *Nucleic Acids Res* 2006, **34(6)**:1692-1699.
  33. Huson DH, Bryant D: **Application of phylogenetic networks in evolutionary studies.** *Mol Biol Evol* 2006, **23(2)**:254-267.
  34. Ciccarelli FD, Doerks T, von Mering C, Creevey CJ, Snel B, Bork P: **Toward automatic reconstruction of a highly resolved tree of life.** *Science* 2006, **311(5765)**:1283-1287.
  35. Altschul SF, Madden TL, Schaffer AA, Zhang J, Zhang Z, Miller W, Lipman DJ: **Gapped BLAST and PSI-BLAST: a new generation of protein database search programs.** *Nucleic Acids Res* 1997, **25(17)**:3389-3402.
  36. Sander C, Schneider R: **Database of homology-derived protein structures and the structural meaning of sequence alignment.** *Proteins* 1991, **9(1)**:56-68.
  37. Cuff JA, Clamp ME, Siddiqui AS, Finlay M, Barton GJ: **JPred: a consensus secondary structure prediction server.** *Bioinformatics* 1998, **14(10)**:892-893.
  38. Kalinina OV, Novichkov PS, Mironov AA, Gelfand MS, Rakhmaninova AB: **SDPPred: a tool for prediction of amino acid residues that determine differences in functional specificity of homologous proteins.** *Nucleic Acids Res* 2004, **32(Web Server issue)**:W424-8.
  39. Hettwer S, Sterner R: **A novel tryptophan synthase b-subunit from the hyperthermophile *Thermotoga maritima*. Quaternary structure, steady-state kinetics, and putative physiological role.** *J Biol Chem* 2002, **277(10)**:8194-8201.
  40. Alves R, Savageau MA: **Evidence of selection for low cognate amino acid bias in amino acid biosynthetic enzymes.** *Mol Microbiol* 2005, **56(4)**:1017-1034.
  41. Ohno S: **Evolution by gene duplication.** Heidelberg, Springer; 1970.
  42. Kondrashov FA: **In search of the limits of evolution.** *Nat Genet* 2005, **37(1)**:9-10.
  43. Jensen RA: **Enzyme recruitment in evolution of new function.** *Annu Rev Microbiol* 1976, **30**:409-425.
  44. Xie G, Bonner CA, Song J, Keyhani NO, Jensen RA: **Inter-genomic displacement via lateral gene transfer of bacterial trp operons in an overall context of vertical genealogy.** *BMC Biol* 2004, **2**:15.
  45. Gupta RS: **Protein phylogenies and signature sequences: A reappraisal of evolutionary relationships among archaeobacteria, eubacteria, and eukaryotes.** *Microbiol Mol Biol Rev* 1998, **62(4)**:1435-1491.
  46. Snel B, Bork P, Huynen MA: **Genomes in flux: The evolution of archaeal and proteobacterial gene content.** *Genome Res* 2002, **12(1)**:17-25.
  47. Price MN, Arkin AP, Alm EJ: **The life-cycle of operons.** *PLOS Genetics* 2006, **2(6)**:e96.
  48. Merkl R: **SIGI: score-based identification of genomic islands.** *BMC Bioinformatics* 2004, **5**:22.
  49. Huber H, Hohn MJ, Stetter KO, Rachel R: **The phylum Nanoarchaeota: present knowledge and future perspectives of a unique form of life.** *Res Microbiol* 2003, **154(3)**:165-171.
  50. Ruepp A, Graml W, Santos-Martinez ML, Koretke KK, Volker C, Mewes HW, Frishman D, Stocker S, Lupas AN, Baumeister W: **The genome sequence of the thermoacidophilic scavenger *Thermoplasma acidophilum*.** *Nature* 2000, **407(6803)**:508-513.
  51. Omelchenko MV, Makarova KS, Wolf YI, Rogozin IB, Koonin EV: **Evolution of mosaic operons by horizontal gene transfer and gene displacement in situ.** *Genome Biol* 2003, **4(9)**:R55.
  52. Makarova KS, Ponomarev VA, Koonin EV: **Two C or not two C: recurrent disruption of Zn-ribbons, gene duplication, lineage-specific gene loss, and horizontal gene transfer in evolution of bacterial ribosomal proteins.** *Genome Biol* 2001, **2(9)**:RESEARCH 0033.
  53. Di Giulio M: **The universal ancestor and the ancestor of bacteria were hyperthermophiles.** *J Mol Evol* 2003, **57(6)**:721-730.
  54. Gribaldo S, Brochier-Armanet C: **The origin and evolution of Archaea: a state of the art.** *Philos Trans R Soc Lond B Biol Sci* 2006, **361(1470)**:1007-1022.
  55. NCBI Genomes: **NCBI complete microbial genomes.** [<http://www.ncbi.nlm.nih.gov/genomes/proks.cgi>].
  56. Laskowski RA, Chistyakov VV, Thornton JM: **PDBsum more: new summaries and analyses of the known 3D structures of proteins and nucleic acids.** *Nucleic Acids Res* 2005, **33(Database issue)**:D266-8.
  57. PDBsum: **PDBsum home page.** [<http://www.ebi.ac.uk/thornton-srv/databases/pdbsum/>].
  58. Krissinel E, Henrick K: **Detection of protein assemblies in crystals.** Edited by: Berthold MR. Springer, Heidelberg; 2005:163-174.
  59. EBI PISA: **EBI PISA home page.** [[http://www.ebi.ac.uk/msd-srv/prot\\_int/pistart.html](http://www.ebi.ac.uk/msd-srv/prot_int/pistart.html)].

60. Jones DT, Taylor WR, Thornton JM: **The rapid generation of mutation data matrices from protein sequences.** *Comput Appl Biosci* 1992, **8(3)**:275-282.
61. Gascuel O: **BIONJ: an improved version of the NJ algorithm based on a simple model of sequence data.** *Mol Biol Evol* 1997, **14(7)**:685-695.

Publish with **BioMed Central** and every scientist can read your work free of charge

*"BioMed Central will be the most significant development for disseminating the results of biomedical research in our lifetime."*

Sir Paul Nurse, Cancer Research UK

Your research papers will be:

- available free of charge to the entire biomedical community
- peer reviewed and published immediately upon acceptance
- cited in PubMed and archived on PubMed Central
- yours — you keep the copyright

Submit your manuscript here:  
[http://www.biomedcentral.com/info/publishing\\_adv.asp](http://www.biomedcentral.com/info/publishing_adv.asp)

